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Testosterone increases bone mineral density in female-to-male transsexuals: a case series of 15 subjects

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Summary

OBJECTIVE—Testosterone therapy for osteoporosis has not been studied extensively in women because of its potential to cause virilization. Female-to-male transsexuals are genetic females who suffer from gender dysphoria and thus take supra-physiologic doses of testosterone to change from the female to male phenotype. The aim of this study is to examine the effects of testosterone treatment on the genetic female skeleton.

PATIENTS AND DESIGN—A group of 15 female-to-male transsexuals was prospectively enrolled for observation over a 2-year period. The subjects had a mean age of 37·0 ± 3·0 years. All of the subjects self-administered testosterone esters intramuscularly at a mean dose of 70·7 ± 4·5 mg weekly.

MEASUREMENTS—The subjects had measurements of bone mineral density (BMD) by dual X-ray absorptiometry (DXA) of the femoral neck and spine (L2–L4) at 12-month intervals. They had determinations of serum oestradiol, testosterone, soluble RANKL (sRANKL), osteoprotegerin (OPG) and urine N-telopeptide (NTX) at the date of enrolment and at the end of 2 years.

RESULTS—There was a significant positive increase in mean BMD of 7·8% at the femoral neck and a nonsignificant increase in mean BMD of 3·1% at the spine over 2 years. The levels of testosterone reached the upper normal range for males and the levels of oestradiol declined to near the postmenopausal range. sRANKL levels decreased significantly in female-to-male transsexuals who newly initiated testosterone therapy. There was no significant change in urine NTX or serum OPG during the study.

CONCLUSIONS—We conclude that supra-physiologic testosterone therapy increases BMD at the hip while maintaining BMD at the spine in female-to-male transsexuals. The effects of testosterone may be the result of testosterone hormone directly acting on the bone or indirectly through aromatization to oestradiol. Lower RANKL levels coupled with unchanged OPG levels results in an increased OPG/RANKL ratio, which may be beneficial to the bone by inhibiting osteoclastogenesis.
Oestrogen is regarded as the dominant sex steroid hormone in maintaining bone mineral density (BMD) in males and females. Oestrogen therapy in postmenopausal women prevents bone loss (Wells et al., 2002), increases BMD (Lindsay & Thome, 1990) and prevents fractures (Women’s Health Initiative, 2002). Oestradiol rather than testosterone in men correlates more strongly with BMD (Greendale et al., 1997; Khosla et al., 1998; Amin et al., 2000; Szulc et al., 2003) and is a stronger determinant of peak bone mass (Khosla et al., 2001). Men who have either defective aromatase (Bilezikian et al., 1998) or oestradiol receptor (Smith et al., 1994) develop severe osteoporosis, which further supports the notion that oestradiol is the more dominant sex steroid in preserving bone density.

Previously, testosterone was considered the more dominant sex steroid in men. With ageing, men experience declining levels of testosterone (Morley et al., 1997) and decreasing BMD (Meier et al., 1984; Jones et al., 1994; Fatayerji et al., 1999; Melton et al., 2000). Testosterone treatment of elderly men with hypogonadism increases BMD (Katznelson et al., 1996; Behre et al., 1997, 1999; Snyder et al., 1999; Amory et al., 2004). However, a study of elderly men rendered hypogonadal with GnRH agonists demonstrated that men receiving oestradiol had significantly lower bone turnover than men receiving testosterone (Falahati-Nini et al., 2000). Thus, it is unclear whether the beneficial effects of testosterone on bone physiology are the direct effect of testosterone or an indirect effect by the aromatization of testosterone to oestradiol, either peripherally or locally by osteoblasts (Shozu & Simpson, 1998).

In the present study, we evaluated the effect of supra-physiologic testosterone administration on BMD and markers of bone turnover in female-to-male (FTM) transsexuals. FTM transsexuals are biologic females who take testosterone in order to change their phenotype to the male gender (Tangpricha et al., 2003). Our initial hypothesis was that testosterone would not be sparing of BMD because oestradiol levels would decline secondary to the antigonadotropic effects of testosterone. We measured BMD at the hip and spine annually over a period of 2 years in addition to levels of sex hormones and markers of bone turnover.

**Subjects and methods**

**Subjects**

Eligible participants for the study were FTM transsexuals who had been prescribed testosterone treatment for gender reassignment. Subjects were excluded if they were taking any medications known to impact BMD other than calcium and multivitamins or if they were currently pregnant.

The study was approved by the Institutional Review Board at Boston University School of Medicine and conducted through the General Clinical Research Center (GCRC) at Boston University School of Medicine. All of the study participants gave written informed consent prior to participation in the study.

**Study protocol**

Subjects attended the GCRC at Boston University School of Medicine for the initial screening and returned to the GCRC every 6 months. Subjects were instructed to return for their visits 1–2 days prior to an injection of testosterone for trough measurements of serum testosterone. The principal investigator performed the initial history and physical examination and interviewed all of the subjects at each subsequent visit.
Bone mineral measurements

BMD at the femoral neck of the hip and spine (L2–L4) were performed at baseline, 12 months and 24 months using a Norland Ellipse dual-energy X-ray absorptiometer. The coefficient of variation (CV) was obtained by daily measurements using a standard phantom. The CV was 1.2% at the femoral neck of the hip and 1.0% at L2–L4 of the spine.

Biochemical measurements

Blood specimens were obtained for serum oestradiol and total testosterone at enrollment, 1 year and 2 years. Serum osteoprotegerin (OPG) and soluble receptor activator of NF-kappaB ligand (sRANKL) were determined at enrollment and at 2 years. A urine sample was collected for determination of urine N-telopeptide (NTX) at each of the annual visits. The levels of serum testosterone and urine NTX were determined by Quest Diagnostics Incorporated (Cambridge, MA, USA). Levels of OPG and sRANKL were determined using a commercially available kit for OPG and sRANKL from ALPCO Diagnostics (Windham, NH, USA) and performed according to the instructions by the manufacturer. Levels of oestradiol were determined using a commercially available kit from Diagnostic Systems Laboratory (Webster, TX, USA) and performed according to protocols supplied by the manufacturer.

Statistical analysis

Descriptive statistics were used to summarize the BMD and biochemical measurements and were expressed as mean ± SEM. Changes in biochemical measurements were evaluated for significance using a paired t-test comparing enrollment baseline and final measurements. Changes in BMD measurements at enrollment, 1 year and 2 years were further analysed with repeated measures by ANOVA. Pair-wise comparisons with adjustment for multiple comparisons (Bonferroni) were performed if the ANOVA was significant. All of the analyses were carried out at the level of α = 0.05. The statistical analyses were performed using Microsoft Excel Office 2003™ (Seattle, WA, USA) and Analyse-It™ (Leeds, UK) statistical software.

Results

Subjects

Twenty-five eligible subjects agreed to participate in the study. Ten subjects failed to complete the 2-year study (three moved from the area and seven failed to return for the 2-year visit). Therefore, 15 subjects completed the 2-year study (Table 1). All of the subjects took intramuscular testosterone esters for their gender reassignment regimen.

There were no differences between the 15 who completed the study and the 10 who failed to complete the study in regards to initial spine BMD (P = 0.65), initial hip BMD (P = 0.88), oestradiol level (P = 0.49), testosterone level (P = 0.66), testosterone dose (P = 0.60) and urine N-telopeptide (P = 0.14). The only significant difference that was found was in age. The dropouts were statistically younger than the subjects who remained in the study, 27.3 ± 3.0 years old vs. 37.0 ± 3.0 years old, P = 0.03.

Seven subjects were taking testosterone for a mean of 4.1 ± 1.0 years prior to enrolling into the study. An additional eight subjects enrolled in the study prior to starting testosterone therapy. Five out of 15 subjects underwent ovariectomy 2.3 ± 0.5 years prior to enrolment in the study (Table 1). All of these subjects were already receiving testosterone therapy.
Serum oestradiol

Seven out of eight subjects had levels of serum oestradiol determined prior to their gender reassignment and before the initiation of testosterone therapy. Their levels of oestradiol significantly decreased by 48% after 2 years of testosterone therapy (Table 2; \(P = 0.02\)). There was no statistically significant change in oestradiol levels in subjects who had already been treated with testosterone (Table 2).

Serum testosterone

Seven out eight subjects had levels of serum testosterone determined prior to their gender reassignment and before the initiation of testosterone therapy. The levels of testosterone significantly increased greater than 14-fold after 2 years of testosterone therapy (Table 2; \(P = 0.006\)). The subjects who were already being treated with testosterone had no significant change in their serum testosterone level (Table 2).

Markers of bone turnover

The mean urine NTX was low at enrollment for both groups of transsexuals, pre- and posthormone treatment for gender reassignment (Table 2). There was no statistically significant change in urine NTX after 2 years of follow-up.

Osteoprotegerin/RANKL signalling

Levels of OPG decreased by 38% in FTM transsexuals from baseline (prior to gender reassignment) to 2-year measurements (\(P = 0.28\); Table 2). There was no change in OPG levels in subjects who were already taking testosterone at enrollment (Table 2).

Levels of sRANKL significantly decreased by 32% in the first 2 years of testosterone therapy in newly initiated transsexuals (\(P = 0.048\); Table 2). Transsexuals who had previously been treated with testosterone prior to entry had lower mean sRANKL than newly treated transsexuals at enrollment and after 2 years. Their sRANKL levels decreased by a further 13% after the 2-year follow-up period (\(P = 0.58\)).

Bone density measurements

Fourteen out of 15 transsexual subjects had increases in BMD at the hip over the 2-year observation period. Repeated ANOVA measurements were significant (\(P < 0.001\)) for changes in hip BMD. There was a significant 7.8% increase after 2 years of observation in mean hip BMD from enrollment (0.984 ± 0.05 g/cm\(^2\)–1.060 ± 0.04 g/cm\(^2\); \(t\)-test with Bonferroni correction, \(P \leq 0.01\); Fig. 1).

Eleven out of 15 transsexual subjects had increases in BMD at the spine over the 2-year observation period. There was a 3.1% increase after 2 years of observation in mean spine BMD from enrollment (1.172 ± 0.04 g/cm\(^2\)–1.209 ± 0.04 g/cm\(^2\); \(t\)-test with Bonferroni correction, \(P \leq 0.01\); Fig. 1). Post hoc analysis with \(t\)-test with Bonferroni correction did not reveal any statistically significant change in enrollment spine BMD measurements to 2-year measurements.

The increases in BMD were analysed in the two subgroups: previously treated with testosterone and testosterone naïve. The previously treated group had a mean increase of 8.4% and 4.4% at the hip and spine, respectively (Fig. 2). The testosterone naïve group had mean increase of 7.2% and 2.0% at the hip and spine, respectively (Fig. 2).

Discussion

This case series demonstrates that testosterone treatment in FTM transsexuals results in increased hip BMD and maintenance of spinal BMD over an observation period of 2 years.
The increase in BMD occurred despite serum oestradiol levels decreasing to near the menopausal range, suggesting a potentially beneficial effect of testosterone. The increase in BMD may be the result of the direct effect of testosterone on bone or an indirect effect of testosterone after aromatization to oestradiol.

One previous study of 19 FTM transsexuals from Holland examined the effects of testosterone administration in regards to spinal BMD only. They demonstrated a loss of BMD at the spine over a 2- to 5-year period (van Kesteren et al., 1998). All of the subjects in this Dutch study underwent ovariectomy within 12–18 months of starting testosterone therapy. They were treated with intramuscular (i.m.) testosterone esters 250 mg every 2 weeks and were continued on a lower dose of parental testosterone postoperatively, testosterone esters 250 mg i.m. every 2–3 weeks, or changed to oral testosterone undeconatoe 160 mg/day. The lower dose of testosterone resulted in higher levels of the gonadotropins LH and FSH (21.6 ± 4.10 and 46.2 ± 10.6, respectively) at the end of the study compared to preoperative levels on higher doses of testosterone (1.9 ± 0.4 and 2.7 ± 0.4, respectively). In our study, only five out of 15 subjects underwent ovariectomy prior to enrolling in our study and no subjects had ovariectomy during the study. The subjects in our study did not reduce their dose of testosterone during the 2-year period. We sampled the levels of LH and FSH from 12 of our subjects at the end of our 2-year study and found that they were more uniformly suppressed (5.4 ± 2.6 and 3.2 ± 1.2, respectively). This may explain why our subjects had positive increases in BMD at the hip and preservation of BMD at the spine compared to the study by van Kesteren et al. (1998).

In a cross-sectional analysis of FTM transsexuals in Singapore, a group of FTM transsexuals who were treated for a period of 1–3 years with similar doses of testosterone used in our study had higher spine BMD compared to a group of testosterone-naive FTM transsexuals. Subjects who were noncompliant with testosterone therapy after ovariectomy had significantly lower BMD than testosterone naïve FTM transsexuals. The BMD of five non-compliant subjects improved with re-initiation of testosterone therapy. The authors concluded that testosterone therapy improves BMD in FTM transsexuals especially in a hormone-deficient state such as after ovariectomy (Goh & Ratnam, 1997). These findings are similar to our study. We demonstrated that continuous testosterone therapy in FTM transsexuals significantly increases the BMD at the hip while maintaining BMD at the spine compared to baseline measurements.

Our subjects were receiving supra-physiologic dosages of testosterone, making comparison to other studies using female replacement doses of testosterone more problematic. These studies examining the effects of testosterone replacement in preserving BMD in females have revealed conflicting findings. A randomized 2-year study of 34 postmenopausal women demonstrated that combination therapy of testosterone plus oestrogen pellets was superior in improving BMD at the hip and spine than oestrogen alone (Davis et al., 1995). Another trial of 66 post-menopausal women demonstrated that oral combination therapy of oestrogen and testosterone therapy was not superior to oestrogen alone in improving spine BMD (Watts et al., 1995). A previous study using oestrogen and testosterone pellets showed no additional benefit of testosterone over oestrogen on hip and spine BMD (Garnett et al., 1992).

Our study is consistent with previous studies that androgens inhibit OPG. Hofbauer et al. (2002) demonstrated that androgens inhibit mRNA and protein secretion in osteoblast cultures. Elderly men rendered hypogonadal with GnRH agonists had decreases in OPG after testosterone therapy (Khosla et al., 2002). Oestrogen appears to have the opposite effect on OPG production. When the same elderly men rendered hypogonadal with GnRH agonists were treated with oestrogen, there were increases in OPG (Khosla et al., 2002).
Several in vitro studies have demonstrated that oestrogen increases OPG levels in bone marrow stromal and osteoblastic cells (Hofbauer et al., 1999; Chen et al., 2000; Saika et al., 2001). Women who take oral contraceptives containing oestrogen have higher serum OPG levels compared to nonusers (Hofbauer et al., 2004). We noted a decrease in soluble RANKL levels and with no increase in circulating OPG levels in the newly treated testosterone subjects. This would result in an increased OPG/RANKL ratio which might be potentially favourable for bone preservation by decreasing osteoclast formation.

We saw a greater increase in BMD at the hip compared to the spine. Because the hip has more cortical bone than the spine, which has more trabecular bone, one potential explanation for this differential effect could be that testosterone has more positive effects on cortical bone than trabecular bone as suggested by Lips et al. (1996). In his study, he performed iliac crest biopsies on FTM transsexuals and found greater cortical thickness compared to women and healthy young men, whereas there was no difference in trabecular bone thickness.

The increase in BMD occurred with no significant decrease in bone turnover measured by urine NTX. The most plausible explanation for this discrepant finding could be that the subjects already had very low bone turnover by baseline measurements of urine NTX prior to initiating testosterone therapy. This would suggest that testosterone may exert beneficial effects on the bone independent from reducing bone turnover.

In summary, this small case series of genetic FTM transsexuals suggests that testosterone therapy in genetic females in supra-physiologic dosages may improve BMD at the hip and maintain BMD at the spine. This improvement occurs in spite of lowered serum oestradiol levels. Larger controlled studies using this human model of androgen supplementation in genetic females may be difficult to conduct due to its extremely low prevalence. Studies using either lower doses of testosterone or less androgenic testosterone derivatives should be conducted to investigate the role of androgens in female skeletal health.

Acknowledgments

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References


Chen XW, Garner SC, Anderson JJB. Effects of 17b-estradiol (E2), genistein (GEN) and daidzein (DIZ) on osteoclastogenesis inhibitory factor (OCIF) and osteoclast differentiation factor (ODF)


Fig. 1.
Mean bone mineral density (BMD) changes in female-to-male transsexuals on testosterone. The mean hip BMD increased by 7.8% after 2 years from initial enrollment in the study ($P < 0.01$, $t$-test with Bonferroni correction for repeated measures). The mean spine BMD increased by 3.1% after 2 years from initial enrollment in the study. *$P < 0.05$. Error bars represent ± SEM.
Fig. 2.
Mean individual change (%) in bone mineral density (BMD) in female-to-male transsexuals on testosterone therapy. The mean percentage increase after 2 years in hip BMD was 8·4% and 7·2% in the previously treated and testosterone naïve groups (left panel). The mean percentage increase after 2 years in spine BMD was 4·4% and 2·0% in the previously treated and testosterone naïve groups (left panel).
Table 1
Baseline characteristics of female-to-male transsexual study participants

<table>
<thead>
<tr>
<th></th>
<th>Testosterone-naïve (n = 8)</th>
<th>Testosterone-treated (n = 7)</th>
<th>All</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>33·1 ± 3·0</td>
<td>40·5 ± 4·0</td>
<td>37·0 ± 3·0</td>
</tr>
<tr>
<td>Years on testosterone</td>
<td>0</td>
<td>4·2 ± 1·0</td>
<td></td>
</tr>
<tr>
<td>Dose of testosterone * (mg testosterone ester/week)</td>
<td>66·0 ± 6·0</td>
<td>76·0 ± 7·0</td>
<td>70·7 ± 4·5</td>
</tr>
<tr>
<td>Current smokers (%)</td>
<td>5/8 (63%)</td>
<td>3/7 (43%)</td>
<td>8 (53·3%)</td>
</tr>
<tr>
<td>Prior ovariectomy (%)</td>
<td>0/8</td>
<td>5/7 (71%)</td>
<td>5/15 (33%)</td>
</tr>
<tr>
<td>Spine L2-L4 (gm/cm(^2))</td>
<td>1·211 ± 0·06</td>
<td>1·134 ± 0·06</td>
<td>1·172 ± 0·04</td>
</tr>
<tr>
<td>Femoral neck (gm/cm(^2))</td>
<td>1·072 ± 0·07</td>
<td>0·861 ± 0·03</td>
<td>0·984 ± 0·05</td>
</tr>
</tbody>
</table>

Data expressed as mean ± SEM.

\* Subjects took either testosterone cypionate or enanthate.
<table>
<thead>
<tr>
<th></th>
<th>Baseline</th>
<th>2 years</th>
<th>%</th>
<th>P-value</th>
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<tbody>
<tr>
<td>Previously treated transsexuals</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Oestradiol * (pmol/l) (n = 4)</td>
<td>161 ± 15</td>
<td>148 ± 20</td>
<td>8.0%</td>
<td>0.67</td>
</tr>
<tr>
<td>Testosterone † (nmol/l) (n = 7)</td>
<td>20.4 ± 3.0</td>
<td>26.4 ± 6.0</td>
<td>+29%</td>
<td>0.36</td>
</tr>
<tr>
<td>OPG (pmol/l) (n = 6)</td>
<td>5.3 ± 0.7</td>
<td>5.5 ± 1.0</td>
<td>+3.8%</td>
<td>0.88</td>
</tr>
<tr>
<td>sRANKL (pmol/l) (n = 6)</td>
<td>0.4 ± 0.7</td>
<td>0.3 ± 0.7</td>
<td>13.0%</td>
<td>0.58</td>
</tr>
<tr>
<td>OPG/RANKL ratio (n = 6)</td>
<td>17.0 ± 5.0</td>
<td>18.4 ± 5.0</td>
<td>+8.0%</td>
<td></td>
</tr>
<tr>
<td>Urine N-telopeptide (n = 7) (BCE/mmol creatinine)</td>
<td>31.7 ± 7</td>
<td>28.3 ± 3</td>
<td>10.5%</td>
<td>0.57</td>
</tr>
<tr>
<td>Newly treated transsexuals</td>
<td></td>
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<td></td>
<td></td>
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<tr>
<td>Oestradiol * (pmol/l) (n = 7)</td>
<td>400 ± 74</td>
<td>206 ± 50</td>
<td>48.65%</td>
<td>0.02</td>
</tr>
<tr>
<td>Testosterone † (nmol/l) (n = 7)</td>
<td>1.52 ± 0.46</td>
<td>22.5 ± 4.8</td>
<td>+1470%</td>
<td>0.006</td>
</tr>
<tr>
<td>OPG (pmol/l) (n = 8)</td>
<td>5.06 ± 0.89</td>
<td>4.30 ± 0.55</td>
<td>38.0%</td>
<td>0.28</td>
</tr>
<tr>
<td>sRANKL (pmol/l) (n = 8)</td>
<td>0.80 ± 0.26</td>
<td>0.45 ± 0.18</td>
<td>32.0%</td>
<td>0.048</td>
</tr>
<tr>
<td>OPG/RANKL ratio</td>
<td>12.3 ± 4</td>
<td>18.1 ± 5</td>
<td>+47.0%</td>
<td></td>
</tr>
<tr>
<td>Urine N-telopeptide (n = 8) (BCE/mmol creatinine)</td>
<td>32.9 ± 8</td>
<td>30.9 ± 6</td>
<td>6.0%</td>
<td>0.84</td>
</tr>
</tbody>
</table>


† Reference range for testosterone, males 9.0–34.7 nmol/l, females < 3.5 nmol/l.