Non-motor symptoms of Parkinson's disease revealed in an animal model with reduced monoamine storage capacity

Tonya N. Taylor, Emory University
William Michael Caudle, Emory University
Kennie R. Shepherd, Emory University
AliReza Noorian, Emory University
Chad R. Jackson, Emory University
P Michael Iuvone, Emory University
David Weinshenker, Emory University
James G Greene, Emory University
Gary W Miller, Emory University

Journal Title: Journal of Neuroscience Nursing
Volume: Volume 29, Number 25
Publisher: Lippincott, Williams & Wilkins | 2009-06-24
Type of Work: Article | Post-print: After Peer Review
Publisher DOI: 10.1523/JNEUROSCI.1495-09.2009
Permanent URL: http://pid.emory.edu/ark:/25593/fhmf4

Final published version: http://www.jneurosci.org/content/29/25/8103

Copyright information:
© 2009 Society for Neuroscience
Accessed August 27, 2020 10:26 PM EDT
Non-motor symptoms of Parkinson’s disease revealed in an animal model with reduced monoamine storage capacity

Tonya N. Taylor1,2, W. Michael Caudle3, Kennie R. Shepherd1,2, AliReza Noorian4, Chad R. Jackson5, P. Michael Iuvone5,6, David Weinshenker7, James G. Greene4,5, and Gary W. Miller1,2

1 Center for Neurodegenerative Disease, Emory University, Atlanta, GA 30322
2 Department of Environmental and Occupational Health, Rollins School of Public Health, Emory University, Atlanta, GA 30322
3 Department of Pathology, University of Washington, Seattle, WA 98104
4 Department of Neurology, School of Medicine, Emory University, Atlanta, GA 30322
5 Department of Pharmacology, School of Medicine, Emory University, Atlanta, GA 30322
6 Department of Ophthalmology, School of Medicine, Emory University, Atlanta, GA 30322
7 Department of Human Genetics, School of Medicine, Emory University, Atlanta, GA 30322

Abstract

Parkinson’s disease (PD) is a progressive neurodegenerative disorder that is characterized by the loss of dopamine neurons in the substantia nigra pars compacta, culminating in severe motor symptoms, including: resting tremor, rigidity, bradykinesia, and postural instability. In addition to motor deficits, there are a variety of non-motor symptoms associated with PD. These symptoms generally precede the onset of motor symptoms, sometimes by years, and include anosmia, problems with gastrointestinal motility, sleep disturbances, sympathetic denervation, anxiety, and depression. Previously, we have shown that mice with a 95% genetic reduction in vesicular monoamine transporter expression (VMAT2-deficient, VMAT2 LO) display progressive loss of striatal dopamine, L-DOPA responsive motor deficits, α-synuclein accumulation, and nigral dopaminergic cell loss. We hypothesized that since these animals exhibit deficits in other monoamine systems (norepinephrine, serotonin), which are known to regulate some of these behaviors that the VMAT2-deficient mice may display some of the non-motor symptoms associated with PD. Here we report that the VMAT2-deficient mice demonstrate progressive deficits in olfactory discrimination, delayed gastric emptying, altered sleep latency, anxiety-like behavior, and age-dependent depressive behavior. These results suggest that the VMAT2-deficient mice may be a useful model of the non-motor symptoms of PD. Furthermore, monoamine dysfunction may contribute to many of the non-motor symptoms of PD and interventions aimed at restoring monoamine function may be beneficial in treating the disease.

Keywords

VMAT2; dopamine; norepinephrine; serotonin; depression; olfaction
Introduction

Parkinson’s disease (PD) is a neurodegenerative disease that has long been considered to be a disorder of the dopamine (DA) system. Pathophysiologically, PD is believed to be caused by the death of neuromelanin-containing DA neurons in the substantia nigra pars compacta (SNpc) and the appearance of proteinaceous intracellular inclusions known as Lewy bodies (Olanow and Tatton, 1999; Jenner and Olanow, 2006). Motor disturbances do not present clinically until approximately 70–80% of striatal dopamine has already been lost; however, other non-motor symptoms are evident before the onset of motor disturbances. These include hyposmia/anosmia, gastrointestinal disturbances, sleep abnormalities, autonomic dysfunction, anxiety, depression, and, at later stages, impaired cognition (Braak et al., 2003; Langston, 2006).

Moreover, a careful reading of James Parkinson’s description of his patients indicates that many of the non-motor symptoms that are of great interest today were observed nearly 200 years ago (Parkinson, 1817). It is possible that other neurotransmitters such as norepinephrine (NE) and serotonin (5-HT) may significantly contribute to these symptoms, as the locus coeruleus (LC) and raphe nucleus have also been shown to degenerate in PD (Dauer and Przedborski, 2003; Fahn and Sulzer, 2004; Rommelfanger and Weinshenker, 2007). This is supported by studies that show neurodegeneration in the LC, the primary source of NE, in addition to the SNpc, and evidence that perturbation of the noradrenergic system can lead to serious alteration in DA neurotransmission (Antelman and Caggiula, 1977; Gesi et al., 2000; Marien et al., 2004; Rommelfanger and Weinshenker, 2007). In contrast to DA and NE, serotonin has been virtually absent from PD research, even though the raphe nuclei has been shown to undergo degeneration in PD patients (Braak et al., 2002; Braak et al., 2003). Although the degeneration of the raphe is not as prominent as that of the SNpc or LC, 5-HT is widely recognized in the development of many other diseases such as depression, psychiatric, and sleep disorders, all of which have been observed in PD patients (Halliday et al., 1990; Jellinger, 1991; Murai et al., 2001).

Recently, we have characterized a new potential model of PD based on reduced vesicular storage of monoamines. Animals expressing 5% of normal vesicular monoamine transporter 2 (VMAT2-deficient) exhibit increased oxidative stress, progressive loss of DA terminals and cell bodies in the SNpc, as well as α-synuclein accumulation (Caudle et al., 2007). Monoaminergic dysfunction in the VMAT2-deficient animals arises from reduced expression of VMAT2, resulting in severely diminished levels of DA, NE, and 5-HT (Mooslehner et al., 2001; Caudle et al., 2007). Because VMAT2-deficient mice demonstrate a significant reduction in NE and 5-HT levels in multiple brain regions, our laboratory has characterized the behavioral manifestations of reduced monoaminergic innervation in the VMAT2-deficient mice. In this study, we report that the VMAT2-deficient mice display many of the non-motor symptoms of PD.

Materials and Methods

Animals

Male and female VMAT2-deficient mice were generated as previously described (Mooslehner et al., 2001; Caudle et al., 2007). Briefly, the mouse VMAT2 locus was cloned from the 129/Sv genomic library and a 2.2 kb PvuII fragment from the third intron of the VMAT2 gene, and cloned into the blunted NotI site of the construct. The targeting vector was introduced into 129/Ola CGR 8.8 embryonic stem (ES) cells and injected into blastocytes of C57BL/6 mice. Highly chimeric males were bred with C57BL/6 females; genotype was confirmed by Southern blot analysis. A recent report uncovered that the C57BL/6 inbred strain of mice originally used to establish the VMAT2-deficient mouse line contained a spontaneous chromosomal deletion spanning the α-synuclein gene locus (Specht and Schoepfer, 2001), which was confirmed in the original strain (Mooslehner et al., 2001; Patel et al., 2003; Colebrooke et al., 2006). Through
diligent breeding, we eliminated all traces of this mutation from our strain of mice and routinely verify the presence of α-synuclein via Southern blot analysis. This report represents the second set of data on VMAT2-deficient mice with a normal α-synuclein background. All mice were generated through redundant breeding of mice that were heterozygous for the VMAT2 allele and wild type (WT) for the α-synuclein allele. The genotype of all mice was confirmed by PCR of DNA extracted from tail samples. For all behavioral tests, VMAT2 WT littermate controls were used. All procedures were conducted in accordance with the National Institutes of Health Guide for Care and Use of Laboratory Animals and previously approved by the Institutional Animal Care and Use Committee at Emory University.

HPLC determination of monoamines and metabolites

HPLC-electrochemical analysis of neurochemistry was performed as previously described (Richardson and Miller, 2004; Caudle et al., 2006; Caudle et al., 2007). Briefly, dissected right striata, cortex, and hippocampus were sonicated in 0.1 M perchloric acid containing 347 μM sodium bisulfate and 134 μM EDTA. Homogenates were centrifuged at 10,000 × g for 10 min at 4°C; the supernatant was removed and filtered through a 0.22 μm filter by centrifugation at 5,000 × g for 3 min. The supernatants were then analyzed for levels of DA, DOPAC, homovanillic acid (HVA), NE, 5-HT, and 5-HIAA. Levels were measured using HPLC with an eight-channel coulometric electrode array (ESA Coularray; ESA Laboratories, Chelmsford, MD). Quantification was made by reference to calibration curves made with individual standards.

Olfactory discrimination

Olfactory experiments were adapted from previous work from our laboratory (Tillerson et al., 2006). Briefly, wooden blocks (1.8 cm³) were placed individually in 50 mL conical tubes containing 1 g of animal bedding from test animals’ cages for 12 h. The animal was presented with a block scented with its own bedding and a block scented with another mouse’s bedding (of the same sex). The time spent sniffing (nose less than 1 cm away from the block) or in contact with each block was recorded for a 2-min trial (Tillerson et al., 2006). A single cohort of mice was tested at each time point; however, naïve mice were introduced each month to ensure that the original cohort of mice was not learning the task. Data from male and female mice was combined, since there were no detectable sex differences after testing 20 mice per genotype.

To measure non-social olfactory acuity, methods were modified from previous methods used in our laboratory (Tillerson et al., 2006). Glass plates with 25 μL of either a novel scent (lemon, peppermint, or vanilla) or water were presented simultaneously to the animals. Time spent sniffing each glass plate was recorded for a 3 minute session.

Non-olfactory sensory tests

To assess non-olfactory sensory function in VMAT2 WT and deficient mice, we examined the responses of these animals in several sensory tests using the paradigm of Tillerson and colleagues (Tillerson et al., 2006). Responsiveness to tactile stimulation was measured as latency to contact or remove a 113.1 mm² (1.3 cm in diameter) adhesive dot in a 2 minute session (Avery Office International) (Schallert et al., 2000; Tillerson et al., 2006). Animals were removed from their home cage, and the dot placed between the ears, on top of the head. Animals were then put back into their home cage and the cage returned to its normal position on the rack to reduce external distractions (Tillerson et al., 2006).

Quinine is often used in taste aversion paradigms, due to its unpalatable bitterness (Tillerson et al., 2006). The tip of a cotton swab was placed into either an aliquot of 2 mg/mL quinine or water and then into the animals’ mouths. Presentation of clean versus quinine stimuli was
counterbalanced between animals. Latency to groom and/or drag the jaw along the ground was recorded during a 1 minute test session (Grill and Norgren, 1978; Schallert and Whishaw, 1978; Tillerson et al., 2006).

The trigeminal nerve innervates areas of the olfactory epithelium and nasal mucosa, and is responsible for non-odor sensations such as mild irritation and burning (Tillerson et al., 2006). In this test, we assessed the function of the trigeminal nerve by exposing the mice to either ammonia, known to exert a trigeminal response, or water. A glass plate with water or ammonia (counterbalanced between animals) was placed in the animal’s cage. Time sniffing (as defined above) was recorded for a 2 minute session (Tillerson et al., 2006).

**Sleep latency**

VMAT2 WT and deficient animals were individually housed in large plexiglass cages and allowed to acclimate for 4 hours. Saline (0.9%) was then administered intraperitoneally and mice were observed by a trained experimenter for behavioral signs of sleep. Sleep was defined as 2 minutes of uninterrupted sleep behavior, and 75% of the next 10 minutes spent asleep (Mitchell et al., 2008). A single cohort of mice was tested at all time points.

**Gastric emptying**

Following a 12-hour fast, VMAT2 WT and deficient mice were allowed free access to food for 1 hour. The amount of food consumed was calculated based on food weight before and after access. Two hours after food removal, animals were killed and the stomach contents were weighed (wet and dry). The percentage of food remaining in the stomach was measured (Whited et al., 2006). Since this is a terminal procedure, separate cohorts of VMAT2 WT and deficient mice were aged and subjected to analysis for each time point.

**One hour stool collection**

Each mouse was placed in a separate clean cage and observed throughout the 60 min collection period. Fecal pellets were collected immediately after expulsion and placed in sealed (to avoid evaporation) 1.5 ml tubes. Tubes were weighed to obtain the wet weight of the stool, this was then dried overnight at 65°C and reweighed to obtain the dry weight (Li et al., 2006).

**Forced swim test**

These studies were conducted on mice using a modified method of Porsolt and coworkers (Porsolt et al., 1979). All mice were injected intraperitoneally with 100 μL of 5 mg/kg desipramine (Sigma-Aldrich, St. Louis, MO) or saline 30 minutes before testing. Mice were placed individually in glass cylinders (24 × 16 cm) with 15 cm of water maintained at 25°C. The mice were left in the cylinder and their behavior was videotaped from the side of the cylinder for 6 minutes. After the first 2 minutes, the total duration of time spent immobile was recorded during a 4 minute test. The mouse was deemed immobile when it was floating passively; subtle movement of feet or tail required to keep the head above the surface of the water were excluded as immobility. Immobility time refers to the time that the animal spent floating for at least 3 seconds (Porsolt et al., 1979; Xu et al., 2000; Fukui et al., 2007). A single cohort of mice was tested at each time point; however, naïve mice were introduced each month to ensure that the original cohort of mice was not learning the task.

**Tail suspension test**

These experiments were conducted using the methods of Cryan and colleagues (Cryan et al., 2004). All mice were injected intraperitoneally with 100 μL of 5 mg/kg desipramine (Sigma-Aldrich, St. Louis, MO) or saline 30 minutes before testing. Mice were individually suspended by the tail to a horizontal ring stand bar (distance from the floor = 30 cm) using adhesive tape.
A 6-minute test session was videotaped and scored by a trained observer for escape-oriented behavior and bouts of immobility. The time spent immobile was recorded for each mouse. Mice were excluded from the study if they were able to climb on top of the ring stand. A single cohort of mice was tested at each time point; however, naïve mice were introduced each month to ensure that the original cohort of mice was not learning the task.

Elevated Plus Maze

Male and female VMAT2 WT and deficient mice were individually housed in a reversed light cycle room (lights on at 1900 h, lights off at 700 h), and were allowed a minimum of two weeks to habituate to the new lighting conditions. Food and water were available ad libitum throughout the course of the study. Data from male and female mice was combined, since there were no detectable sex differences.

The elevated plus maze (EPM) paradigm was adapted from Schank and colleagues (Schank et al., 2007). The EPM apparatus consisted for two open arms and two enclosed arms arranged in a plus sign orientation. The arms were elevated 30 inches above the floor, with each arm projecting 12 inches from the center. Because rodents naturally prefer dark, enclosed compartments, a greater willingness to explore the open, well-lit arms is believed to represent a decrease in the animal’s anxiety (Pellow et al., 1985; File et al., 1988; Paine et al., 2002).

No drugs were administered prior to behavioral testing. To begin each test, mice were placed in the EPM, facing an open arm and allowed to freely explore the apparatus during a videotaped, 5 minute trial. Videotapes were later scored by an observer who was blind to genotype. The measure used for analysis is the percentage of time spent exploring the open arms, which was calculated by dividing the time spent in the open arms by the combined time spent in open and closed arms (Pellow et al., 1985). Entry into an arm of the EPM was defined as the animal placing all four paws in that particular part of the maze. All tests were run during the dark cycle, between 1400 and 1600 h. Mice were excluded from data analysis if they jumped or fell off the maze after the test began.

Grid test

The grid test was performed using the methods of Tillerson and colleagues (Tillerson and Miller, 2003). Mice were placed horizontally on the center of the grid and supported until they grasped the grid with their forepaws and hindpaws. The grid was then inverted and the mice were videotaped while hanging upside-down for latency to release their grip on the grid.

Electroretinalgraphy (ERG)

VMAT2 WT and deficient mice were dark-adapted for at least 12 hrs and anesthetized with a ketamine (60mg/kg) (Hospira, Inc., Lake Forest, IL)/xylazine (7.5mg/kg) (Sigma Aldrich, St. Louis, MO) mixture for recording the scotopic flash ERG. Pupils were dilated with 1% cyclopentolate/1% tropicamide solutions. A DTL-fiber working electrode was placed on the cornea of the eye, while the reference needle electrode was placed on the cheek just below the eye. A ground electrode was attached to the tail of the mouse and a rectal probe was inserted to monitor and control the temperature of the mouse. After recording a flat baseline, the mouse was exposed to multiple flashes of light with increasing intensities (.111, .228, .576, 1.16, 2.325, 5.85, 119.6 cd*s*m$^{-2}$). The responses to 3–10 flashes at each intensity were averaged to determine a-wave amplitude, b-wave amplitude, a-wave implicit time, and b-wave implicit time.

J Neurosci. Author manuscript; available in PMC 2010 January 28.
Statistical analysis

Data from male and female mice was combined, since there were no detectable sex differences. All data were analyzed using unpaired independent samples Student’s t-test (gastric emptying), completely randomized two-factor ANOVA followed by Bonferroni post hoc analysis (odor discrimination, olfactory acuity, sleep latency, gastric emptying, forced swim test, tail suspension test), or repeated measures two-factor ANOVA (ERGs). Analyses were completed using GraphPad Prism 5.0 for Windows, and for all tests all post hoc measures were error-corrected to keep the overall error rate per group at 0.05.

Results

VMAT2-deficient mice have diminished levels of monoamines at 12–15 months of age, accompanied by increased turnover

Previously, our laboratory has shown that VMAT2-deficient animals have age-dependent reductions in levels of striatal dopamine, accompanied by increased turnover (Caudle et al., 2007). To determine if the reduction in VMAT2 affect the other major monoamines in addition to DA, the striatum, cortex, and hippocampus were isolated from VMAT2 WT and deficient mice and analyzed by HPLC for neurochemical levels. At 12–15 months of age, the VMAT2-deficient mice had severely diminished levels of all three monoamines in all brain regions tested, although in general, DA and NE depletion were more severe than 5-HT depletions. In the striatum, DA and NE were decreased by 91%, while 5-HT was decreased by 81%. Similarly, in the cortex there was a 92% and 94% decrease in DA and NE, respectively, with only a 78% decrease in 5-HT. However, in the hippocampus, a 99% decrease in DA was seen, while NE and 5-HT were decreased by 87% and 86%, respectively (Figure 1A). VMAT2-deficient animals also had significantly increased DA and 5-HT turnover; a two-way ANOVA revealed a significant brain region by genotype interaction for both DA (F(2,38)=80.91; p<0.0001) and 5-HT turnover (F(2,44)=26.09; p<0.0001) (Figure 1B, C).

VMAT2-deficient mice display progressive olfactory discrimination deficits

One of the earliest manifestations of PD is the loss of the sense of smell. Olfactory abnormalities have been found in nearly all PD patients and can precede neurological deficits by decades (Braak et al., 2002; Braak et al., 2003; Langston, 2006). To determine whether VMAT2-deficient animals exhibit comparable olfactory dysfunction, both VMAT2 WT and deficient animals were subjected to a battery of olfactory discrimination tests. First, VMAT2 WT and deficient animals were subjected to odor discrimination tests, using wooden blocks scented with either home cage bedding or bedding from the cage of a foreign mouse of the same sex. No differences were seen between the behavior of VMAT2 WT or deficient animals at 2 and 4 months of age shown as the investigatory index (percentage of time spent investigating self – percentage of time spent investigating other; n=12, Figure 2). However, at 5, 6, and 12 months of age, VMAT2-deficient animals were unable to discriminate between the two blocks, and consequently displayed no preferential exploration of either block (2 months: % self=28.19, % other=71.81; 12 months: % self=44.63, % other=55.37; interaction between effects of scent and age: F(4,95)=2.41, p<0.05, n=12, Figure 2). In contrast, VMAT2 WT mice show marked preferential exploration of the block scented with foreign bedding (n=12–15, main effect of scent: F(1,110)=546.44, p<0.0001), with no change in behavior as they aged (2 months: % self=28.69, % other=71.31, 12 months: % self=13.14, % other=86.86; main effect of age: F(4,110)=0.36, p=0.8379). To ensure that these results were not due to differences in motivation, the total investigatory time was calculated for both genotypes of mice at all ages. Investigatory times were similar at 4 months (WT: 5.36±0.6505, n=11; LO: 4.50±1.167, n=10; t(19)=0.6627, p=0.5155), 5 months (WT: 6.17±0.5618, n=12; LO: 5.00±0.9309, n=10; t(20)=1.114, p=0.2785), and 6 months of age (WT: 4.36±0.8232, n=11; LO: 2.80±0.4667, n=10; t(19)=1.608, p=0.1244); only at 12 months of age did the VMAT2-deficient animals display a significant reduction in
total investigatory time relative to age-matched WT littermates (WT: 9.11±0.8219, n=15; LO: 5.52±1.418, n=11; t24=2.325, p=0.0288). The olfactory deficit is not corrected by L-DOPA treatment in human PD patients, nor is it effective in our mice. Acute administration of L-DOPA and the peripheral aromatic acid decarboxylase inhibitor, benserazide did not affect the performance of either genotype (data not shown) (Muller et al., 2002). Two-way ANOVA exposed a significant scent/genotype interaction; VMAT2-deficient mice also had deficits in non-social olfactory acuity using the scents of lemon (interaction between effects of genotype and age: F(1,32)=3.69, p=0.0637), vanilla (interaction between effects of genotype and age: F(1,34)=10.04, p=0.0032), and peppermint (interaction between effects of genotype and age: F(1,36)=4.91, p=0.0331), beginning at 3 months of age (data not shown) and persisting until 18 months of age (Figure 3).

Mice were tested for non-olfactory sensory deficits to ensure that there was not a problem in general sensory perception. VMAT2-deficient animals showed no deficits in response to tactile stimulation (interaction between effects of genotype and age: F(1,45)=0.03, p=0.8679), quinine-induced taste aversion (main effect of genotype: F(1,44)=0.00, p=0.9830), or to mild irritation/burning sensations stimulated by ammonia (main effect of genotype: F(1,38)=0.14, p=0.7136) as compared to age matched WT controls (Figure 4). Collectively, these results demonstrate that although the VMAT2-deficient animals have normal tactile and gustatory sensory perception, they do exhibit progressive deficits in olfactory discrimination, which is a common phenotype seen in PD.

**VMAT2-deficient animals display altered latency to behavioral signs of sleep**

Many PD patients experience sleep disturbances, including excessive sleepiness and insomnia (Comella, 2003; Langston, 2006; Ziemssen and Reichmann, 2007). In order to begin investigating behavioral sleep disturbances in the VMAT2-deficient mice, we conducted sleep latency tests in VMAT2 WT and deficient mice during their circadian nadir. Beginning at 2 months of age, VMAT2-deficient mice show a shorter latency to behavioral signs of sleep, which is most prevalent at 4–6 months of age, than age-matched WT controls (interaction effects of genotype and age: F(4,73)=2.82, p=0.0310). A Bonferroni post hoc analysis compared the two genotypes at each age; the difference in sleep latency between VMAT2-deficient and WT animals becomes increasingly less pronounced, and is absent at 24 months of age (Figure 5A). While VMAT2 WT animals display an age-dependent decrease in sleep latency, no age-related changes in sleep latency occur in VMAT2-deficient mice. The circadian activity of VMAT2-deficient animals is also significantly lower than that of age-matched WT controls at 4–6 months of age (main effect of genotype: F(1,156)=37.19, p<0.0001), but not at 12–15 (main effect of genotype: F(1,228)=2.62, p=0.1072) or 18 months of age (main effect of genotype: F(1,168)=1.16, p=0.2827), due to a decline in WT circadian activity at the latter ages (Figure 5B–E). These suggest that VMAT2-deficient mice develop premature changes in sleep latency and are displaying behavioral phenotypes reminiscent of older WT animals.

**VMAT2-deficient mice have delayed gastric emptying**

Gastrointestinal dysfunction in PD occurs in over 70% of PD patients and has been attributed to lack of activity, inadequate hydration, or autonomic and enteric neuronal dysfunction (Langston, 2006; Ziemssen and Reichmann, 2007). To study this non-motor symptom in VMAT2-deficient mice, WT and deficient animals were behaviorally examined for gastric emptying at 2, 6, 12, and 18 months of age. Student’s t-test revealed that solid gastric emptying was significantly delayed overall in VMAT2-deficient mice (WT: 21.98 ± 2.605 n=16, LO: 30.27 ± 2.101 n=16, t28=2.478, p=0.0195), and was most apparent at 2, 6, and 12 months of age, although not significant as shown by two-way ANOVA with Bonferroni post hoc analysis (n=16 per genotype, interaction effects of age and genotype: F(3,21)=0.63, p=0.6043) (Figure 6A). Two-way ANOVA revealed stool frequency was also found to be
altered in VMAT2-deficient mice, with a significant increase in frequency at 2 and 6 months of age, compared to VMAT2 WT animals (n=4 per genotype, interaction effects of age and genotype: F(3,24)=8.79, p=0.0004, Figure 6B).

**VMAT2-deficient mice display anxiety-like and depressive phenotypes**

One of the most prevalent non-motor symptoms of PD is depression, with signs of anxiety co-morbid in patients with major depression (Zimmerman et al., 2002; Fukui et al., 2007). Anxiety-like behavior was measured in VMAT2 WT and deficient animals using the elevated plus maze. Younger VMAT2-deficient mice (4–6 mo) spent significantly more time in the closed arms of the maze as compared to age-matched controls (n=11–12, interaction effects of age and genotype: F(1,38)=3.45, p<0.05, Figure 7A), while older VMAT2-deficient mice (12–15 months) did not display an anxiety-like phenotype compared to age-matched WT controls. Similar to what we observed in the sleep latency test, the behavior of VMAT2 WT animals in this test changed over time, while that of VMAT2-deficient mice did not, indicating possible premature development of symptoms normally associated with older animals.

The VMAT2-deficient animals were tested for altered behavior in the forced swim (FST) and tail suspension (TST) tests. At 4–6 months of age, there was no difference found between the immobility times of VMAT2 WT and deficient animals in both tests, and neither genotype responded to an acute low dose of desipramine (5 mg/kg administered 20 minutes prior to testing) (n=4–6, interaction effects of drug treatment and genotype: F(1,13)=0.53, p=0.4807 (FST); F(1,13)=0.08, p=0.7827 (TST); Figure 7B, C). However, at 12–15 months of age, immobility times were higher in VMAT2-deficient animals compared to age-matched WT controls (n=4–5, main effect of genotype: F(1,15)=9.72, p=0.0071 (FST); F(1,16)=11.12, p=0.0042 (TST), Figure 7D, E). The differences in immobility times across age groups in VMAT2-deficient animals suggests that there is an age-dependent development of depressive behavior, which correlates with the progressive neurochemical deficits previously characterized (Mooslehner et al., 2001; Caudle et al., 2007). Initially, both VMAT2 WT and deficient mice were exposed to 20 mg/kg desipramine, which was found to decrease immobility times in both genotypes (data not shown). However, when treated with an acute low dose of desipramine (5 mg/kg administered intraperitoneally 20 minutes prior to testing), the immobility times of VMAT2-deficient mice decreased to WT levels (n=4–5, interaction effects of treatment and genotype: F(1,15)=5.55, p=0.0325 (FST); F(1,16)=7.95, p=0.0123 (TST); Figure 7D, E); the immobility times of VMAT2 WT animals were not affected by desipramine at 12–15 months of age.

**VMAT2-deficient mice display normal vision and muscle strength**

While the phenotypic behaviors help define the disease, the lack of deficits in other systems are also important in showing the relative selectivity. As in PD, other sensory systems appear to be functioning normally in the VMAT2-deficient mice (Figure 3, 4); moreover, vision and muscle strength have also been found to be normal in patients. Although VMAT2-deficient mice had significantly decreased DA (64%) and DOPAC (45%) (interaction effects of neurotransmitter and genotype: F(1,20)=16.28, p=0.0006; Figure 8A), the magnitude of the decrease was substantially less than that found in the brain regions examined and was coupled with increased DA turnover (WT: 0.2434 ± 0.039 n=6, LO: 0.4017 ± 0.056 n=6, t10=2.317, p=0.0430). Consequently, visual function appeared to be normal. Visual function was quantified by use of electroretinography (ERG). The ERG can detect abnormalities in electrical responses of the photoreceptor cells (rods and cones) and inner retinal cells (bipolar, amacrine, and ganglion cells). Compared to wild type littermates, VMAT2-deficient mice displayed no differences in A-wave amplitude (main effect of genotype: F(1,4)=0.04, p=0.8569) or implicit time (main effect of genotype: F(1,4)=0.04, p=0.8480) or B-wave amplitude (main effect of genotype: F(1,4)=0.28, p=0.6237) or implicit time (main effect of genotype: F(1,4)=0.02,
p=0.8895) when their retinas were stimulated with light flashes as revealed by repeated measures two-way ANOVA with Bonferroni post hoc analyses (Figure 8B–E). In addition, Acdy1 mRNA expression, which is regulated in retina by DA D4 receptors (Jackson et al., 2009), was not different between VMAT-deficient (2.450±0.4204, n=11) and wild type mice (2.112±0.4775, n=10; t10=0.5324, p=0.8019). General muscle strength was assessed by subjecting VMAT2 WT and deficient animals to the grid test. VMAT2-deficient mice showed no difference in latency to fall on the grid test apparatus compared to age-matched WT animals (data not shown).

Discussion

Although Parkinson’s disease (PD) has been traditionally viewed as a neurodegenerative motor disorder, the increasing recognition of non-motor symptoms, including hyposmia, sleep abnormalities, anxiety, depression, and gastric dysfunction, suggests the disease is more multifaceted than commonly thought. Moreover, these symptoms imply that more general monoaminergic dysfunction is occurring in concert with dopamine (DA) degeneration. Evidence for degeneration of the locus coeruleus (LC) and dorsal raphe (DR) in PD patients highlights the importance of looking beyond the nigrostriatal system in order to illuminate the deficits in other neurotransmitter systems (Braak et al., 2002; Braak et al., 2003; Dauer and Przedborski, 2003; Fahn and Sulzer, 2004; Lemke et al., 2004; Rommelfanger and Weinshenker, 2007). In this study, we hypothesized that general monoamine dysfunction may recapitulate many of the non-motor symptoms of PD. We observed a reduction in monoamines that was associated with behavioral dysfunction.

Previous neurochemical analysis of VMAT2-deficient animals shows general age-dependent reductions in brain tissue levels of DA, norepinephrine (NE), epinephrine, and serotonin (5-HT) by approximately 80–90% (Mooselehner et al., 2001; Colebrooke et al., 2006; Caudle et al., 2007), which is accompanied by increased dopamine and serotonin turnover in multiple regions of the brain (Figure 1) (Colebrooke et al., 2006; Caudle et al., 2007). Loss of VMAT2 function causes gradual neurodegeneration in the SNpc, LC, and DR of aged VMAT2-deficient mice (Taylor and Miller, unpublished observations). In contrast, other pharmacological in vivo models of PD, such as 6-OHDA lesions, cause sudden and profound loss of cell bodies. Although the precise mechanisms of cell death remain unclear, this progressive degeneration may be caused by oxidant species produced from improper monoamine storage within the neuron (Caudle et al., 2008). Catecholamines have the intrinsic ability to auto-oxidize, yielding reactive oxygen species and cysteiny1 adducts; likewise, serotonin has an oxidizable indole ring (Graham, 1978; Jenner, 1998, 2003; Guillot and Miller, 2009). Presumably, improper storage of the monoamines contributes to progressive cellular injury.

Braak and colleagues have proposed that PD pathology actually begins in the lower brainstem and olfactory bulb, revealed by the presence of α-synuclein positive Lewy bodies (Braak et al., 2002; Langston, 2006). Olfaction abnormalities are present in 100% of PD patients, many noticing a decline in the sense of smell long before the onset of motor symptoms (Muller et al., 2002). Alterations in olfactory function are typically stable over time, non-responsive to traditional PD therapeutics, and unrelated to disease stage or duration (Doty et al., 1992; Ziemssen and Reichmann, 2007), and have been suggested to be an excellent pre-clinical marker of PD. VMAT2-deficient mice demonstrate a progressive loss in olfactory discrimination beginning at 5 months of age, stabilizing at 6 months, and remaining unchanged at 12 months that appears to correlate to the loss of striatal DAT (Caudle et al., 2007), similar to that seen in patients (Siderowf et al., 2005; Ross et al., 2008).

Sleep disturbances have been shown to occur prior to the clinical presentation of motor deficits (Braak et al., 2003; Langston, 2006). Nocturnal sleep disruptions occur in 60–98% of PD
patients, and are correlated with disease severity (Comella, 2003; Ziemssen and Reichmann, 2007). Sleep disturbances are manifested through a variety of syndromes including REM sleep behavioral disorder (RBD), excessive daytime sleepiness, sleep fragmentation, and deficits in sleep latency (Friedman, 1980; Comella, 2007; Ziemssen and Reichmann, 2007). VMAT2-deficient mice demonstrate a shorter latency to behavioral signs of sleep; however, more in depth studies using EEGs must be performed to better understand the underlying sleep architecture in these mice.

Gastrointestinal dysfunction is certainly one of the most common and possibly one of the earliest symptoms of PD (Edwards et al., 1993; Abbott et al., 2001; Pfeiffer, 2003). Dopamine depletion in the colon and Lewy bodies throughout the enteric nervous system suggest GI symptoms are a manifestation of the primary disease process, rather than an epiphenomenon related to motor dysfunction (Qualman et al., 1984; Kupsky et al., 1987; Wakabayashi et al., 1988; Edwards et al., 1993; Singaram et al., 1995; Abbott et al., 2001; Langston, 2006). Unfortunately, treatment of PD with L-DOPA may exacerbate GI dysfunction by slowing gastrointestinal motility, even though increased colonic transport time may derive from the same processes that cause motor abnormalities (Abbott et al., 2001; Ziemssen and Reichmann, 2007). The VMAT2-deficient mice exhibit some disturbances in GI dysfunction, including delayed gastric emptying and altered stool frequency that declines with age (Figure 6), indicating that reduced vesicular monoamine storage impacts GI function.

Disruptions in DA, NE, and 5-HT neurotransmission have been found in PD patients with anxiety and/or depression, including degeneration of the LC and DR (Lemke et al., 2004; Remy et al., 2005; Ziemssen and Reichmann, 2007). Depression in PD has been difficult to study since it differs from major depression alone; only about 6% of PD patients suffer from major depression (Tandberg et al., 1996; Ziemssen and Reichmann, 2007). Moreover, a reliable rating scale does not currently exist to measure depressive symptoms in PD patients (Tandberg et al., 1996; Ziemssen and Reichmann, 2007). However, antidepressants such as nortriptyline and paroxetine have proven effective in depressed PD patients without worsening motor symptoms (Andersen et al., 1980; Cervavolo et al., 2000). The role of VMAT2 in anxiety and depression was discovered decades ago and is well characterized; reserpine, a VMAT2 inhibitor, has been shown to precipitate depressive-like symptoms in humans (Freis, 1954). Recently, Wetsel and colleagues have shown that mice with a 50% reduction in VMAT2 display depressive behavior in the forced swim and tail suspension test that is ameliorated by acute antidepressant therapy (Fukui et al., 2007). However, these VMAT2 heterozygous knockouts do not display anxiety-like behavior (Fukui et al., 2007). We found that the severe reduction of VMAT2 expression in the VMAT2-deficient mice does trigger both anxiety and depressive behavior. VMAT2-deficient mice showed a significant increase in percent open arm time in the elevated plus maze at 4–6 months of age, while the increased immobility time in the forced swim and tail suspension tests did not occur until 12 months of age; suggesting that anxiety precedes depressive symptoms in VMAT2-deficient animals. Additionally, a low dose of desipramine that had no effect in wild-type animals was able to normalize immobility times in VMAT2-deficient mice. It is also important to note that many of the behaviors that are normal in PD patients (sensory, vision, strength) are also normal in the VMAT2-deficient mice, demonstrating a selective deficit in monoamine-related behaviors.

Data from the VMAT2-deficient mice suggest that all monoamine transmitter systems, not just DA, play a role in the clinical manifestations of PD. Classical descriptions/hypotheses the pathogenesis of PD suggest the degeneration or loss of DA neurons is the major contributor to the development of the disease. However, it is not implausible to couple NE and 5-HT with DA in these classically established hypotheses for PD genesis. NE and 5-HT share structural similarities with DA; NE shares a catechol ring, and all three species have the ability to oxidize within the cell yielding deleterious effects (Guillot and Miller, 2009), the qualities that
predispose the nigrostriatal system to oxidative damage may not be unique (Ahlskog, 2007); rather, these factors may be present in all monoaminergic neurons to varying degrees. Although the cause of the non-motor symptoms associated with PD remains unknown, it is clear that they are not caused purely by dopaminergic deficits; our data suggest that NE and 5-HT may contribute as well. When combined with the previously reported nigrostriatal degeneration in this model, the observed non-motor symptoms suggest that the VMAT2-deficient mice represent an excellent model of the monoaminergic deficits that manifest in human PD.

Acknowledgments

This work was supported by NIH grants P01 ES016731 (Miller), 1F31ES017247-01 (Taylor), R01 EY004864 (Iuvone), T32 GM007092 (Jackson), NSF 0450303 (Jackson), and the Michael J. Fox Foundation (Greene). The authors would like to thank Heather Mitchell and Dr. Jesse Schank for assistance with behavioral testing, Dr. Min Wang for assistance with the mouse colony, and Dr. Nikita Pozdeyev for technical assistance with ERG recordings. The authors are indebted to Dr. Piers Emson for providing the original VMAT2 hypomorph mouse strain.

References


Li ZS, Schmauss C, Cuenca A, Ratcliffie E, Gershon MD. Physiological modulation of intestinal motility by enteric dopaminergic neurons and the D2 receptor: analysis of dopamine receptor expression, location, development, and function in wild-type and knock-out mice. J Neurosci 2006;26:2798–2807. [PubMed: 16525059]


Figure 1.
VMAT2 LO mice display widespread reductions in monoamines at 12–15 months of age. A, All monoamine levels are significantly lower than age-matched wildtype controls at 12–15 months of age in the striatum, cortex, and hippocampus. Results represent the mean ratios of raw values (ng/mg tissue) ± SEM for 8 animals per genotype. B, C, Ratio of DOPAC/DA and 5-HIAA/5-HT in the striatum, cortex, and hippocampus of VMAT2 WT and VMAT2 LO mice. Results represent the mean ratios of raw values (ng/mg tissue) ± SEM for 4 animals per genotype. ***p<0.001
Figure 2.
VMAT2 LO animals display progressive deficits in olfactory discrimination. When given the choice between a wooden block scented with a foreign animal’s bedding and a block scented with the animal’s own bedding, VMAT2 WT animals show preferential exploration of the block scented with the foreign animal’s bedding at all ages, as shown by the investigatory index. The investigatory index represents the percentage of time the animal spent sniffing the block scented with a foreign animal’s bedding minus the percentage of time the animals spent sniffing the block scented with their own bedding. Conversely, VMAT2 LO animals exhibit a preferential exploration at 2 and 4 months of age, with a significant decrease at 5 months of age, worsening at 6 and 12 months of age. Results represent the investigatory index ± SEM for 10–15 animals per genotype. ***p<0.001, **p<0.01, *p<0.05
Figure 3.
VMAT2 LO mice display age-dependent deficits in non-social olfactory acuity. A, When given the choice between a novel odor (lemon) and water, both genotypes show a preferential exploration of the novel scent at 2 months of age, but the VMAT2 LO mice lose the ability to discriminate between lemon and water by 18 months of age. B, C, VMAT2 LO animals display similar behaviors at 18 months of age when presented with the novel odors of peppermint (B) and vanilla (C). Results represent the time spent investigating each scent ± SEM for 10 animals per genotype. ***p<0.001. **p<0.01
Figure 4.
VMAT2 LO mice do not display deficits in general sensory behavioral tests. A. At 6 months and 12 months of age VMAT2 LO mice display similar response latencies to tactile stimulation in the dot test. C, D. Trigeminal nerve function in VMAT2 wild-type and LO animals was tested by response to ammonia. Like the VMAT2 WT mice, VMAT2 LO mice displayed preferential exploration of water compared to ammonia at both 6 (B) and 12 (C) months of age. Gustatory function was also found to be normal in VMAT2 LO mice at 6 (D) and 12 (E) months of age as measured by the quinine taste aversion test.
Figure 5.
VMAT2 LO animals display normal circadian activity but a premature shortened latency to behavioral signs of sleep. A, VMAT2 LO mice at 4–6 months of age display shorter latency to behavioral signs of sleep as compared to WT controls. VMAT2 WT and LO animals have similar latencies to behavioral signs of sleep at 12–15 and 24 months of age. Results represent the time (min) passed until the animal achieved 2 minutes of uninterrupted sleep ± SEM for 8 animals per genotype. *p<0.05 B-E, VMAT2 LO animals display normal circadian activity levels at 2, 4–6, 12–15, and 18 months of age, as compared to age-matched WT controls.
Figure 6.
VMAT2 LO animals have delayed gastric emptying. A, Data broken down by age reveals a potential age-dependence of the effect. B, VMAT2 LO mice have increased stool frequency compared to age-matched VMAT2 WT animals at 2 and 6 months of age. Results represent average stool frequency for two trials ± SEM for 4 animals per genotype. **p<0.01, ***p<0.001
Figure 7.
VMAT2 LO animals display an anxiety-like and a progressive depressive-like phenotype. A, Four to six-month-old VMAT2 LO animals spend less time in the open arms and more time in the closed arms of the elevated plus maze as compared to age-matched WT animals over the 5 min test period. At 12–15 months of age, VMAT2 WT and LO animals display similar amounts of time in both the closed and open arms of the EPM. Results represent the mean time (seconds) ± SEM for 7 mice per genotype. *p<0.05 B and C, Four to six-month-old VMAT2 LO animals have similar immobility times in the forced swim test (B) and tail suspension test (C) as compared to age-matched WT animals; the immobility times of both genotypes is not affected by 100 μL of 5 mg/kg desipramine 30 minutes prior to testing given intraperitoneally. D, At 12–15 months of age, VMAT2 WT animals remain non-responsive to desipramine administration. However, VMAT2 LO mice have an increased immobility time, which is
decreased to WT levels when dosed intraperitoneally with 100 μL of 5 mg/kg desipramine 30 minutes prior to testing. Results represent the mean time (seconds) ± SEM for 4–5 mice per genotype. *p<0.05, **p<0.01. At 12–15 months of age, VMAT2 LO mice display a significant increase in immobility time compared to WT control animals, which is ameliorated by 5 mg/kg desipramine. **p<0.01, n=4–5 mice per genotype.
VMAT2 LO mice have decreased retinal DA but normal vision. A, VMAT2 LO animals have a 64% decrease in retinal DA, concurrent 45% decrease in retinal DOPAC, and increased DA turnover. B–E, No changes were observed in A-wave amplitude (B) and implicit time (C) or in B-wave amplitude (D) and implicit time (E) in VMAT2 LO mice compared to age-matched VMAT2 WT mice.

Figure 8.