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Abstract

Excessive alcohol use is extremely prevalent in the United States, particularly among trauma-exposed individuals. While several studies have examined genetic influences on alcohol use and related problems, this has not been studied in the context of trauma-exposed populations. We report results from a genome-wide association study of alcohol consumption and associated
problems as measured by the alcohol use disorders identification test (AUDIT) in a traumaexposed cohort. Results indicate a genome-wide significant association between total AUDIT score and rs1433375 \( [N = 1036, P = 2.61 \times 10^{-8} \text{ (dominant model)}, P = 7.76 \times 10^{-8} \text{ (additive model)}] \), an intergenic single-nucleotide polymorphism located 323 kb upstream of the sodium channel and clathrin linker 1 (SCLT1) at 4q28. rs1433375 was also significant in a meta-analysis of two similar, but independent, cohorts \( (N = 1394, P = 0.0004) \), the Marine Resiliency Study and Systems Biology PTSD Biomarkers Consortium. Functional analysis indicated that rs1433375 was associated with SCLT1 gene expression and cortical-cerebellar functional connectivity measured via resting state functional magnetic resonance imaging. Together, findings suggest a role for sodium channel regulation and cerebellar functioning in alcohol use behavior. Identifying mechanisms underlying risk for problematic alcohol use in traumaexposed populations is critical for future treatment and prevention efforts.

**Keywords**

alcohol consumption; alcohol use disorder; AUDIT; expression QTL; fMRI; genome-wide association study

**INTRODUCTION**

Excessive alcohol use is the third leading cause of preventable deaths in the United States (Jonas et al. 2012; Centers for Disease Control and Prevention 2012). A large literature has demonstrated that excessive alcohol use and related disorders are particularly prevalent among traumaexposed individuals, such as those living in neighborhoods with high rates of assaultive and interpersonal violence (Stockdale et al. 2007) and among military personnel (Allen, Crawford, & Kudler 2016). While several studies have examined genetic influences on alcohol involvement, there is a limited understanding of genetic risk for drinking in populations with high levels of trauma exposure.

While several large-scale genetic studies of alcohol dependence (AD) have been conducted over the past two decades, few replicable genetic markers have been identified. Notable exceptions include findings from the Study of Addiction: Genetics and Environment and the Collaborative Study on the Genetics of Alcoholism, which report risk variants for AD in novel loci including KIAA0040 (Zuo et al. 2012), the PHF3-PTP4A1 gene complex (Zuo et al. 2011, 2014), rs1437396 at chromosomal location 2p16.1 and other variants in the alcohol dehydrogenase (ADH) gene cluster (Gelernter et al. 2014). However, with the exception of the ADH gene cluster, genetic variants (and corresponding biological pathways) identified in previous genetic studies of AD have not yet pointed to tractable mechanisms underlying genotype-phenotype associations that intersect with current neurobiological understanding of the effects of alcohol on behavior.

Previous studies have indicated that genetic analyses of quantitative phenotypes that precede AD, such as frequency and quantity of alcohol consumption, may improve power to detect variants with small effects (Agrawal et al. 2009). For example, genome-wide association studies (GWASs) of alcohol consumption have implicated several genome-wide significant variants (e.g. Schumann et al. 2011; Clarke et al. 2017; Webb et al. 2017). Reviews of the
current state of GWAS findings for alcohol use behaviors indicate that larger sample sizes and quantitative measures of alcohol use increase the likelihood of identifying genome-wide significant findings via an increase in statistical power. For this reason, conducting GWAS on quantitative measures of alcohol use has gained popularity (Agrawal et al. 2009; Baik et al. 2011; Heath et al. 2011; Schumann et al. 2011; Takeuchi et al. 2011; Agrawal et al. 2013; Kapoor et al. 2013; Wang et al. 2013; Xu et al. 2015; Clarke et al. 2017; Webb et al. 2017).

In this study, we aimed to increase understanding of genetic risk for alcohol use behavior by conducting a GWAS in the Grady Trauma Project (GTP), a large, primarily African-American community cohort study initially designed to assess post-traumatic stress disorder (PTSD; Binder et al. 2008; Gillespie et al. 2009; Ressler et al. 2011; Almli et al. 2013). The GTP sample was recruited from the general medical clinics of the Grady Memorial Hospital, a publicly funded hospital that serves economically disadvantaged individuals in Atlanta, Georgia. To validate GWAS findings from the GTP, we also analyzed the findings in a similar sample consisting of 1394 men and women from two independent cohorts with high rates of trauma exposure. Finally, we characterized GWAS findings in the discovery (GTP) sample using both gene expression and brain imaging data available on a subset of the population. Given that the executive control network (ECN) has been repeatedly associated with alcohol use behavior, and in particular, fronto-cerebellar connectivity (Chanraud et al. 2010; Herting, Fair, & Nagel 2011; Sutherland et al. 2012; Fryer et al. 2013; Krmpotich et al. 2013; Moulton et al. 2014), we examined the influence of genetic risk variants implicated in alcohol use behavior on this neural network.

METHODS AND MATERIALS

Discovery sample and assessment

The GTP cohort was collected to investigate the genetic and environmental predictors of psychiatric disorders in a population of urban men (28.2 percent) and women (71.8 percent). The mean (SD) age in the sample was 39.5 (13.4) years, ranging from 18 to 65 years (Table S1). Screening interviews were completed on participants approached while in the waiting rooms of primary care or obstetrical-gynecological clinics of the Grady Memorial Hospital in Atlanta, Georgia. Participant’s demographic information (e.g. self-identified race, sex and age) and substance use behaviors were assessed by verbal interview ($15 for participation in this phase of the study). Further details regarding the GTP dataset can be found in Gillespie et al. (2009). Written and verbal informed consent was obtained for all participants, and all procedures in this study were approved by the institutional review boards of Emory University School of Medicine and Grady Memorial Hospital, Atlanta, Georgia.

Alcohol consumption and associated behaviors were measured using alcohol use disorders identification test (AUDIT), which measures both frequency of use and consequences accompanying excessive use (Saunders et al. 1993; Babor et al. 2001); the total score was obtained by summing the responses from the 10 questions on the AUDIT survey. Each item of the AUDIT is rated on a scale of 0–4, with higher scores reflecting more problematic alcohol drinking. Thus, our composite score is a quantitative measure of alcohol consumption and related hazardous behaviors (Babor et al. 2001; Iacono et al. 1999; Reinert & Allen 2002, 2007). Of the individuals included in the GTP genetic sample ($N = 3340,
Table S1), 55 percent had AUDIT data ($N = 1829$). Of those with AUDIT data, 57 percent reported drinking in the last year ($N = 1036, 373$ men and $663$ women). To avoid confounding our measure of problematic drinking behavior with the decision to initiate alcohol use (Sartor et al. 2013), only individuals who had at least one drink during the last year (57 percent) were included in the GWAS ($N = 1036$). While the AUDIT is typically used as a screening tool in clinical settings, two recent studies have validated the use of AUDIT scores as a useful tool to examine genetic risk for alcohol use behavior in samples with lower levels of overall alcohol consumption and alcohol use problems as is observed in the GTP (Mbarek et al. 2015; Xu et al. 2015).

**Genetic data: quality control and analyses**

**Genotyping**—Grady Trauma Project participants provided a saliva sample and/or blood sample. With Oragene saliva kits (DNA Genotek Inc., Ottawa, Ontario, Canada), DNA was extracted from saliva in Oragene collection vials (DNA Genotek Inc.) using the DNAdvance kit (Beckman Coulter Genomics, Danvers, MA), while DNA from blood was extracted using either the E.Z.N.A. Mag-Bind Blood DNA Kit (Omega Bio-Tek, Inc., Norcross, GA) or ArchivePure DNA Blood Kit (5 Prime, Inc., Gaithersburg, MD). Genome-wide single-nucleotide polymorphism (SNP) genotyping was conducted on 4622 subjects using Illumina’s (San Diego, CA) HumanOmniExpress ($N = 280$) and Omnil-Quad BeadChip ($N = 4342$). The HumanOmniExpress interrogates 730 525 individual SNPs per sample, while the Omnil-Quad BeadChip interrogates 1 011 219 individual SNPs. Genotypes were called using Illumina’s GENOMESTUDIO software. PLINK (Purcell et al. 2007) was utilized to perform quality control (QC) analyses on the genetic data from each chip separately and then after the data from each chip was combined (Supporting Information Fig. S1). For each chip, we first removed individuals with greater than 5 percent missing data. We further identified and removed related individuals by using PLINK to estimate the proportion of identity by descent (IBD) for each pair of individuals. Among pairs of individuals with an IBD proportion $>0.12$ (indicating cousins or a closer relation), we removed the individual in each pair with the higher rate of missing genotype data. We restricted SNPs to autosomes, those with greater than a 95 percent call rate, a frequency of less than 0.01 and Hardy-Weinberg equilibrium (HWE) $P$-value $< 1 \times 10^{-5}$. We next performed principal component analysis (PCA) to infer axes of genetic ancestry. Prior to PCA, we used PLINK to prune the data in windows of 50 base pairs, removing one SNP from each pair of SNPs with $r^2 > 0.05$ to obtain a set of roughly independent SNPs. Based on PCA, we retained those individuals who fell within three standard deviations of the medians of the first and second principal components of self-reported African Americans in order to minimize genetic differences due to genetic ancestry. Supporting Information Fig. S2 shows that these individuals cluster with Africans in a plot of the first and second principal components anchored by three HapMap populations [CEU, YRI and Asian (CHB and JPT combined)]. After completion of QC and PCA, we merged the cleaned data from the two chips and performed more stringent QC filters; SNPs with call rates less than 98 percent, a frequency of less than 0.1 and a HWE $< 1 \times 10^{-5}$ were removed. After conducting a PCA on the combined chip data, we removed
another 99 subjects who fell outside of our criteria (i.e. three standard deviations from the medians of the first and second principal components of self-reported African Americans); thus, the genetic sample consisted of 3340 African-American individuals genotyped for 501,278 SNPs. When comparing the genotyped individuals to the non-genotyped individuals, the proportion of men to women was different with a greater proportion of men in the analytic sample (Table S1). We note that to address this potential confound, we used sex as a covariate throughout the discovery and follow-up analyses.

**Gene expression**—A subset of the participants following the screening interview returned for more detailed interviews and a blood draw. The methodology for assessment of gene expression has been previously described by Mehta et al. (2011, 2013). Briefly, whole blood was collected between 8 and 9 A.M. in Tempus RNA tubes. Whole-genome expression profiles were generated as follows: RNA was isolated using the Versagene kit (Gentra Systems, Big Lake, MN), quantified using the NanoPhotometer and checked for quality on the Agilent (Santa Clara, CA) Bioanalyzer. Total RNA (250 ng) were reverse transcribed to cDNA, converted to biotin-labeled cRNA using the Ambion kit (AMIL1791, Applied Biosystems, Foster City, CA). cRNAs (750 ng) were hybridized to Illumina HT-12 v3.0 or v4.0 arrays (Illumina) and incubated overnight for 16 hours at 55°C. Arrays were washed, stained with Cy3-labeled streptavidin, dried and scanned on the Illumina BeadScan confocal laser scanner. A total of 21,394 transcripts were on both the v3.0 and v4.0 arrays and were significantly expressed above background levels (detection $P < 0.05$) in at least 10 percent of subjects and were thus eligible for further analysis. Briefly, raw microarray scan files from Illumina HT-12 v3.0 arrays were exported using the Illumina BEADSTUDIO program and loaded into R for downstream analysis (www.R-project.org). Evaluation of the different microarray steps was performed using the Illumina internal controls. The data were transformed and normalized using variance-stabilizing normalization (Huber et al. 2002). A total of 15,877 probes passing the filter criteria of Illumina probe detection $P$-value of $<0.01$ in at least 5 percent of the individuals were used for subsequent analysis. To correct for confounding due to batch effects, data were normalized using an empirical Bayes method for batch correction (Johnson, Li, & Rabinovic 2007). Reproducibility of gene expression data was assessed using six pairs of technical replicates, yielding average Pearson correlations of 0.996. Only the nearest gene to the most significant GWAS variant was investigated.

**Statistical analyses for genetic data in discovery sample**—Using the statistical package PLINK (Purcell et al. 2007), we regressed the quantitative outcome, total AUDIT score (among drinkers only, $N=1036$), on allele count assuming an additive model (zero, one or two copies of risk allele), including sex, chip type (Illumina OmniExpress or Omni-Quad) and 10 principal components (Price et al. 2006; Lin & Zhoea 2009) as covariates. Each marker was run separately; thus, to account for the multiple testing, a threshold of $9.97 \times 10^{-8}$ (Bonferroni correction = 0.05/501,278 markers analyzed; note that this differs from the standard GWAS threshold of $5 \times 10^{-8}$) was required to meet genome-wide significance. Quantile-quantile (QQ) plots and Manhattan plots were generated using R (R development team, 2011). Regional association plots were generated using Locus Zoom (Pruim et al. 2010). To ensure that our test results did not depend on linearity or distributional assumptions (Supporting Information Fig. S3), we verified key results with PLINK’s max(T)
permutation ($1 \times 10^{-8}$). To test the model of best fit for the top SNP (e.g. additive or dominant), we used Clarke’s method, which tests the hypothesis that both models are equally distant from the true model (Clarke 2007). To determine if any individual AUDIT item was driving the overall association effect, the significant variant was subsequently tested for association with the individual AUDIT items. To examine the specificity of effects on alcohol use problems, the significant variant was also tested for association with co-morbid conditions of drug abuse, PTSD, depression and anxiety. Gene expression analyses were conducted in R by regressing SNP genotype on transformed and normalized gene signal intensities (Mehta et al. 2013) using both additive and dominant models. Gene-based tests of association were conducted using Multi-marker Analysis of GenoMic Annotation (de Leeuw et al. 2015), using sex, chip type and 10 principal components as covariates. To account for the number of genes tested, a $P$-value threshold of $3.31 \times 10^{-5}$ was considered genome-wide significant (0.05/1508 genes tested).

**Confirmation analyses in similar cohorts**—Parallel SNP-based association analyses were conducted in the Marine Resiliency Study (MRS) and Systems Biology PTSD Biomarkers Consortium (SBPBC) military samples for all variants meeting genome-wide significance criteria. These military cohorts have higher than average trauma levels (Supporting Information, Table 2). The all-male MRS (Baker et al. 2012; Mustapic et al. 2014; Nievergelt et al. 2014) is a prospective study of Marines recruited from four infantry battalions of the First Marine Division stationed at the Marine Corps Air-Ground Combat Center, 29 Palms, or Camp Pendleton, both in California ($N = 1275$). The SBPBC military cohort was recruited as part of a larger study that was designed as a longitudinal examination of male and female combat veterans (Yan et al. 2013; Yehuda et al. 2014), and participants were recruited at two sites in New York City ($N = 119$). Analyses were run parallel to those described in the GTP. Briefly, using the statistical package PLINK, we regressed total AUDIT score on allele count (zero, one or two risk alleles) as above, with sex (for SBPBC), study information (for MRS: MRSI/MRSII) and principal components (five for MRS and three for SBPBC) as covariates, thus controlling for ancestry across the analyses. Using inverse-variance meta-analysis, in R (cite), we assessed the significance of the association between SNP and total AUDIT score when combined across two independent cohorts (the SBPBC cohort plus MRS, which includes four separate self-identified racial/ethnic groups), using the R package Metafor (Viechtbauer 2010).

Post hoc, we examined whether there were associations between total AUDIT score and published SNPs previously associated with AD or alcohol consumption (Bierut et al. 2010; Gelernter et al. 2014; Xu et al. 2015); linear regressions were conducted as described above (Supporting Information).

**Imaging data**

**Data acquisition and preprocessing**—Imaging data were obtained on a subset of the GTP sample including 67 African-American women between 27 and 45 years of age (mean $= 40.6$, $SD = 12.8$; 52 women had GG genotype ‘resilient’ and 15 had AA/AG ‘risk’ genotype of rs14333 75, Supporting Information). Exclusion criteria for the GTP imaging sample included current psychotropic medication use; medical or physical conditions that
preclude magnetic resonance imaging scanning; a history of bipolar disorder, schizophrenia or other primary psychotic disorder; current alcohol or substance dependence; medical conditions that contribute significantly to psychiatric symptoms; history of head injury or loss of consciousness for >5 minutes; or history of neurological disease. Scanning took place on a research-dedicated Siemens 3-T TIM-Trio scanner (Siemens Medical Solutions, Malvern, PA) using a 12-channel head coil at Emory University Hospital. Structural images were acquired using a three-dimensional T1-weighted magnetization-prepared rapid gradient echo imaging sequence (176 slices, 1 × 1 × 1 mm³, repetition time (TR) = 2600 milliseconds, echo time (TE) = 3.02 milliseconds, flip angle = 8°). Resting state functional images were acquired using the Z-SAGA pulse sequence (Heberlein & Hu 2004) to recover signal loss due to susceptibility artifacts. Participants were instructed to focus on the cross in the middle of the screen while staying very still and awake during the resting state scan. Volumes were acquired axially, parallel to the anterior-posterior commissure line (scan time = 7 minutes 23 seconds, 150 volumes, 30 slices, 3.44 × 3.44 × 4 mm³, TR = 2950 milliseconds, TE1 = 30 milliseconds, TE2 = 67 milliseconds, flip angle = 90°).

Data preprocessing was accomplished using a combination of tools from AFNI (Cox 1996) and FSL (Smith et al. 2004) packages. Structural images were skull stripped, segmented and then registered to the standard Montreal Neurological Institute (MNI) space (AFNI’s 3dQwarp). Functional image preprocessing begins with despiking, slice timing correction, motion correction and spatial smoothing (full width at half maximum = 6 mm). The time series were additionally processed to minimize artifacts from head motion, respiration, cardiac pulsation and hardware using the ANATICOR method (Jo et al. 2013) by regressing out motion parameters and averaged signal from eroded local white matter mask as well as band-pass filtering between 0.01 and 0.1 Hz. Finally, functional images were registered to the MNI space via co-registration to the structure image using boundary-based registration (Greve & Fischl 2009) within the FSL package.

Functional connectivity analyses—The ECN includes the ventrolateral prefrontal cortex (VLPFC), the superior parietal cortex, the angular gyrus and cerebellum crus regions (Seeley et al. 2007; Krienen & Buckner 2009). To specifically derive fronto-cerebellar connectivity, we conducted functional connectivity analyses to examine the contralateral fronto-cerebellar circuit using previously defined right and left cerebellum crus regions (Krienen & Buckner 2009) (MNI = −36, −70, −46 and 36, −68, −44 with 6-mm-diameter spheres). First, the preprocessed time series were averaged across these seed regions. Pearson’s correlation coefficient was calculated between the seed time series and other time series within the cerebellum ROI. Then, these correlation coefficients were converted to z-values using Fisher’s transformation. Two-sample t-tests were used to examine quantitative group difference in functional connectivity. To correct for multiple comparisons, Monte Carlo simulations were performed using AFNI’s 3DCLUSTSIM program with whole-brain mask.

RESULTS

The demographic characteristics of the GTP subjects are shown in Table S1, including a comparison between the full genetic sample and the analytic sample. Note that there was a
greater proportion of men in the analytic sample compared to the full genetic sample. Sixty-seven percent of those who consumed alcohol reported drinking less than two alcoholic drinks per day, with male drinkers consuming more alcohol per day than female drinkers. Among alcohol consumers, the total AUDIT score had an overall mean of 7.0 (SD = 7.3) and was higher among men (male: mean = 9.6, SD = 8.0; female: mean = 5.6, SD = 6.5).

**Genome-wide association study in discovery sample**

Analyses included 501,278 SNPs genotyped on Illumina’s Omni1-Quad and OmniExpress arrays. There was no evidence to suggest inflation of the association test statistics in the discovery (GTP) sample (genomic inflation factor $\lambda = 0.98$; Supporting Information Fig. S4). Following a Bonferroni multiple-test correction, rs1433375, an intergenic SNP residing upstream of SCLT1 at chromosomal position 4q28, was significantly associated with total AUDIT score [minor allele frequency of 0.17; assuming an additive model: $N = 1036$, $\beta (SE) = 2.21 (0.41)$; $P = 7.76 \times 10^{-8}$; Fig. 1a, Table 1; permuted $P = 1.2 \times 10^{-7}$]. The means of total AUDIT score by genotype are as follows: AA 8.6, AG 8.6 and GG 6.3. Further, five SNPs in linkage disequilibrium with rs1433375 also associated with total AUDIT score (Fig. 1b). Of note are two sub-threshold SNPs in UDP-Gal:betaGlcNAc beta 1,4-galactosyltransferase, polypeptide 6 (B4GALT6) (Table 1).

We conducted several *post hoc* analyses. Clarke’s method indicated that the dominant model was the preferred model ($P = 2.1 \times 10^{-14}$). Under a dominant model, rs1433375 was genome-wide significant [$\beta (SE) = 2.69 (0.48)$; $P = 2.61 \times 10^{-8}$, Table 1]. In addition to the total AUDIT score, rs1433375 was associated with individual AUDIT items and related measures of substance and alcohol use behavior (e.g. ‘Have you had a problem with alcohol or drugs within the last year’; Supporting Information Table S3) but was not associated with co-morbid conditions of drug abuse, PTSD, depression and anxiety (Supporting Information). Total AUDIT score was not associated with published SNPs previously associated with AD or alcohol consumption (Supporting Information Table S4). No gene-based test of association withstood a genome-wide significance threshold; however, genes meeting a nominal level of significance are shown in Supporting Information Table S5.

**Additional genetic analyses in similar samples**

Next, rs1433375 was tested for association in two similar cohorts assessed with the total AUDIT score. Using an inverse-variance meta-analysis of two independent cohorts ($N = 1394$), the MRS and SBPBC military cohorts, additional evidence for the association of the A allele of rs1433375 and total AUDIT score was observed [$\beta (SE) = 0.60 (0.17)$, $P = 0.0004$, Fig. 2, Table 2]. Thus, in both the discovery sample and the extension sample, the A allele of rs1433375 is associated with increased total AUDIT score. Note that results were driven by the SBPBC military cohort and by the MRS participants who identified as ‘Hispanic and Native American’.

**Functional analyses: SCLT1 gene expression**

To determine the functional basis of the GWAS risk variant, we examined whether rs1433375 influences expression levels of SCLT1, which was the closest gene passing QC. Furthermore, rs1433375 is reportedly a suggestive expression quantitative trait locus for
SCLT1 in the frontal cortex (BRAINEAC, Trabzuni et al. 2011). In a subset of the GTP discovery sample, with peripheral blood RNA available, the normalized signal intensities of SCLT1 (Illumina probe ILMN_1797107; accession NM_144643.2) were used for expression analysis (N = 330). Findings indicated that rs1433375 genotype significantly predicted messenger RNA (mRNA) expression [N = 330, β (SE) = 0.047 (0.02), P = 0.047]. Under the dominant model [N = 330, β (SE) = 0.059 (0.03), P = 0.026], we show that A-allele carriers of rs1433375 exhibited higher mRNA expression (i.e. higher signal intensities; Fig. 3a).

Functional analyses: neuroimaging

We hypothesized that rs1433375 would influence fronto-cerebellar connectivity, an intermediate phenotype previously associated with alcohol use disorders. Functional connectivity analyses were performed using seed regions for the left and right cerebellum crus regions, the areas of highest SCLT1 gene expression (Fig 3b). We found that the right-seeded contralateral fronto-cerebellar connectivity was significantly decreased in the risk group (P < 0.05, whole-brain correction), relative to the resilient group, in the left VLPFC (Brodmann area 44; Fig. 3c). Although there was no cortical-cerebellar difference when using the left seed, we found that the risk group showed decreased cerebellar connectivity relative to the resilient group between the left cerebellum crus regions (Supporting Information Fig. S5).

DISCUSSION

This study reports a novel association between intergenic variant rs1433375 (upstream of SCLT1) and alcohol use behavior in a community cohort of African Americans with high levels of trauma exposure. Two independent populations of mixed ancestry provide additional support for this association. In addition, rs1433375 associates with SCLT1, which is expressed most highly in the cerebellum, a brain area that has been repeatedly associated with alcohol involvement (Baker et al. 1999; Andersen 2004; Manzardo et al. 2005; Hill et al. 2011; Fitzpatrick & Crowe 2013). Finally, we examined resting state brain activity (functional magnetic resonance imaging) in a subset of the GTP to determine whether human cerebellar function was differentially associated with the rs1433375 polymorphism and found altered connectivity in the ECN in carriers of the ‘A’ risk allele. Converging evidence from genotypic, expression-based and functional neuroimaging data presented in this study provides support for the role of SCLT1 in alcohol use and related problems, particularly among highly trauma-exposed populations.

Findings from this study point to novel genetic pathways and mechanisms, in addition to providing support for previously identified pathways. Results from both the discovery (GTP) and additional (SBPBC and MRS) samples indicate that rs1433375, an SNP residing upstream of SCLT1 at chromosomal position 4q28, and five correlated (in linkage disequilibrium) SNPs significantly associated with total AUDIT score. Further, we found that rs1433375 was associated with related measures of substance use and behavior in the full GTP cohort, suggesting that this association is robust to different measures of alcohol use behavior. In addition, the finding of rs1433375 in two independent cohorts of mixed ancestry suggests that this variant may not be unique to populations of African ancestry. We
also note that two SNPs located within B4GALT6 (Table 1) associate with total AUDIT score but do not meet genome-wide significance. However, B4GALT6 has previously been associated with alcohol-related phenotypes, including alcohol consumption during pregnancy (Guasch, Renau-Piqueras, & Guerri 1992; Segui et al. 1996). This converging evidence suggests that variants in both SCLT1 and B4GALT6 should be investigated in future genetic association studies of alcohol use disorders.

While no gene-based test of association withstood a genome-wide significance threshold, several important genes met nominal levels of significance below genome-wide threshold (detailed in Supporting Information Table S5), including genes that have previously been associated with alcohol-related phenotypes. For example, HTR1A and HTR1B have been implicated in several genetic association studies of alcohol and drug dependence, including studies of co-morbid AD and depression (Huang et al. 2003; Sinha, Cloninger, & Parsian 2003; Wrzosek et al. 2011; Contini et al. 2012; Cao, LaRocque, & Li 2013; Zuo et al. 2013). In addition, PER2 has been shown to influence alcohol consumption and related problems in human and animal studies (Spanagel et al. 2005; Yuferov, Bart, & Kreek 2005; Partonen 2015).

The AUDIT was designed for the World Health Organization to screen for hazardous as well as harmful drinking in various cultural settings. There have been a number of studies reflecting potential benefits of using different cutoff points for identification of at-risk drinking (Reinert & Allen 2007). Prior studies have evaluated the test-retest reliability of the AUDIT when it is scored dichotomously to classify patients as at risk. Three prior studies (Selin 2003; Dybek et al. 2006; Rubin et al. 2006) were conducted with general population samples and reported ks of 0.70, 0.86, and 0.89, respectively, at the standard AUDIT cut-point of 8. Together, such data suggest that while a variety of cut-points may be used, an AUDIT score of 8 is generally considered a good marker of risk (Reinert & Allen 2007). Given that we found means of total AUDIT score by genotype of AA/AG = 8.6, whereas GG = 6.3, we believe that these are likely clinically significant differences representing differential at-risk alcohol drinking behavior.

Heritability estimates based on twin data suggest moderately high heritability at 60 percent for AUDIT (Mbarek et al. 2015). AUDIT score represents the composite effects of alcohol consumption and related behaviors; this variable is quite distinct from maximum number of drinks (as reported in Gelernter et al. 2014; Xu et al. 2015) and the case-control AD (as reported in Bierut et al. 2010). Previously published variants conferring risk for heavy alcohol consumption and/or AD were not associated with AUDIT scores in the GTP cohort. We hypothesize that this lack of replication is due to several aspects of the GTP population and study design. Given the known heterogeneity of genetic influences on typical alcohol use, risk drinking and AD (Kendler et al. 2011), it is likely that different risk variants may influence maximum drinks consumed in a 24-hour period, AD diagnoses and AUDIT scores for example. In addition, the characteristics of the GTP study population differ in several ways from previous studies of alcohol use behavior and problems including genetic ancestry, patterns of substance use, demographic characteristics and relatively high levels of trauma exposure, PTSD and depression co-morbidity. For example, the majority of previous studies...
have been conducted in samples of European ancestry, in which general patterns of alcohol use are greater, levels of SES are higher and exposure to assaultive trauma is lower.

rs1433375 is located within a distal upstream enhancer region for SCLT1 transcriptional regulation. Interestingly, SCLT1 produces a linker-like protein between the voltage-gated sodium channel Na(v)1.8 (SCN10A) and clathrin (CLTC), a ubiquitous protein involved in receptor endocytosis and recycling (Liu et al. 2005). Sodium channel activity, particularly Na(v)1.8, has been associated with the neuronal effects of ethanol exposure (Mullin & Hunt 1987; Harris & Allan 1989; Horishita & Harris 2008) and linked to alcohol use behavior (Botta et al. 2010). Moreover, the interaction with sodium channels has been hypothesized as a potentially robust target of novel drugs for the treatment of alcohol use disorders (Rezvani et al. 2012). We note that risk allele carriers of rs1433375 exhibited higher SCLT1 mRNA expression levels in this study. The mechanism for which ‘overexpression’ of SCLT1 can lead to pathological conditions is beyond the scope of this study; however, examples of this relationship can be found in Prelich (2012).

Because SCLT1 is highly expressed in the cerebellum, we used cerebellar seed regions for connectivity analyses. We hypothesized that variation in rs1433375 would influence connectivity between the cerebellum and prefrontal cortex, components of the ECN. Significant differences in connectivity emerged between participants with and without the risk genotype for rs1433375 (‘A’ carriers); risk allele carriers demonstrated lower fronto-cerebellar connectivity compared to those without the risk allele (Figure 3c).

These patterns of neural connectivity suggest decreased functional interactions between prefrontal brain regions controlling complex decision making and the cerebellum, which is thought to be associated with motoric and habitual behavior. Both the prefrontal cortex and cerebellum are thought to play a role in executive control functions, particularly in the presence of emotionally salient stimuli (Konarski et al. 2005). One particularly salient control function is the ability to refrain from maladaptive, but inherently rewarding behaviors, such as excessive alcohol consumption. Earlier studies have shown that individuals with alcohol use problems have demonstrated deficits in executive control (Chanraud et al. 2010; Maurage et al. 2014) as well as diminished activation in prefrontal brain regions (Forbes et al. 2014) and lower frontal-cerebellar connectivity (Rogers et al. 2012) during performance of reward-related tasks.

Given that the genotype groups were well matched in terms of alcohol use, the present findings suggest that those individuals who carry the rs1433375 risk allele may be more susceptible to problematic alcohol use. This may be secondary to alterations in connectivity in the ECN, including cerebellar regulation of responses mediated by the frontal cortex. Thus, altered fronto-cerebellar connectivity may represent an intermediate phenotype for risky alcohol use. Because SCLT1 is expressed highly in the cerebellum (Fig. 3b), we hypothesized that variation in rs1433375 would influence fronto-cerebellar connectivity. Functional connectivity analyses for the cerebellum (i.e. an area of high SCLT1 gene expression) indicated that there were significant differences in connectivity between the cerebellar crus and VLPFC for those with and without the risk genotype (rs1433375 ‘A’ allele carriers). Given that the VLPFC has an established role in impulse control (Sebastian

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et al. 2014), this may suggest a potential neural substrate for problematic alcohol use patterns for carriers of this risk genotype.

Results from this study should be interpreted in light of several key limitations. First, sample sizes in both the discovery (GTP) and additional (MRS and SBPBC) samples were limited; however, the use of a quantitative measure of alcohol use problems, the cross-methodology convergence, as well as the extension of the findings in an independent sample lessens this concern. Second, we note the lack of replication in an independent sample of African ancestry, owing to limitations in availability of samples with the same phenotype. Such replications represent an important next step in understanding these findings. Third, the present analysis uses data available at a single timepoint, using retrospective reports of alcohol use behavior. Previous studies have found that individuals typically underreport their drinking consumption (e.g. Ekholm, Strandberg-Larsen, & Gronbaek 2011), which would likely diminish the association effects observed in this study. However, recent studies directly comparing retrospective reports to observational data conclude that this bias is significantly less pronounced in regular drinkers (Ekholm et al. 2011). Further, the assessment of alcohol use behavior used in this study, total AUDIT score, has excellent psychometric properties and has been used widely in prior health research (Saunders et al. 1993; Babor et al. 2001; Reinert & Allen 2002, 2007; Carey, Carey, & Chandra 2003; Cassidy, Schmitz, & Malla 2008).

This study has provided evidence of association between alcohol involvement and a novel genetic variant located upstream of SCLT1, a gene that has been associated with the regulation of sodium channel functioning, a putative target of ethanol action. Importantly, rs1433375 was also associated with differential cortical–cerebellar functioning, a neural circuit that has been hypothesized to be mechanistically associated with alcohol use disorders. Converging data for the role of a sodium channel regulator from the genotypic, expression-based and functional neuroimaging findings presented in this study provide support for these mechanistic pathways in alcohol use and related behaviors.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgements

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References


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Figure 1.
Genome-wide association study (GWAS) for problematic alcohol use and behavior in the GTP discovery sample. (a) Manhattan plot of the GWAS for total alcohol use disorders identification test score ($N = 1036$, rs1433375 $P = 7.76 \times 10^{-8}$, black line is Bonferroni-adjusted threshold for genome-wide significance). (b) Regional plot of single-nucleotide polymorphism (SNP) associations within 500 kb of rs1433375 (purple triangle) on chromosome 4. Several SNPs are in linkage disequilibrium with the top SNP.
Figure 2.
Meta-analysis of two independent cohorts extends the findings of association between single-nucleotide polymorphism (SNP) rs1433375 and problematic alcohol use and behavior. Forest plot showing the meta-analysis of SNP rs1433375 with total AUDIT score in the additional samples (N = 1394, P = 0.0004). Abbreviations: EA, European Americans; AA, African Americans; HNA, Hispanic/Native Americans; SBPBC, Systems Biology PTSD Biomarkers Consortium
Figure 3.
Functional analyses of rs1433375 in Grady Trauma Project discovery sample. (a) rs1433375 associates with gene $SCLT1$ as shown by significant differences in $SCLT1$ messenger RNA expression levels in blood (probe signal intensities, mean + standard error of the mean) between genotypes ($N = 330$, $P = 0.026$). (b) Allen Brain Atlas image shows high $SCLT1$ expression (red dots) in the cerebellum. (c) Effect of genotype on resting state functional connectivity in executive control network using the right seed: the risk group (A-allele carriers, $N = 15$) showed decreased fronto-cerebellar connectivity relative to the resilient
group (GG genotype, N = 52), between the left cerebellum crus region and a cluster in the left ventrolateral prefrontal cortex (Brodmann area 44, BA 44)
### Table 1

Top 10 variants from SNP-based analysis of alcohol consumption and behavior (alcohol use disorders identification test score)

<table>
<thead>
<tr>
<th>Chr</th>
<th>Gene</th>
<th>Approximate distance to downstream gene</th>
<th>SNP</th>
<th>BP location</th>
<th>MAF</th>
<th>$\beta (SE)$</th>
<th>$P$-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>4</td>
<td>Intergenic</td>
<td>323 kb, SCLT1</td>
<td>rs1433375</td>
<td>130338059</td>
<td>A (0.17)</td>
<td>2.21 (0.41)</td>
<td>$7.76 \times 10^{-8}$a</td>
</tr>
<tr>
<td>4</td>
<td>Intergenic</td>
<td>350 kb, SCLT1</td>
<td>rs995637</td>
<td>130364383</td>
<td>T (0.16)</td>
<td>2.22 (0.42)</td>
<td>$1.96 \times 10^{-7}$</td>
</tr>
<tr>
<td>3</td>
<td>Intergenic</td>
<td>149 kb, CHL1</td>
<td>rs991639</td>
<td>89511</td>
<td>A (0.13)</td>
<td>2.24 (0.46)</td>
<td>$1.22 \times 10^{-6}$</td>
</tr>
<tr>
<td>4</td>
<td>Intergenic</td>
<td>318 kb, SCLT1</td>
<td>rs6837783</td>
<td>130332644</td>
<td>A (0.13)</td>
<td>2.32 (0.48)</td>
<td>$1.60 \times 10^{-6}$</td>
</tr>
<tr>
<td>4</td>
<td>Intergenic</td>
<td>332 kb, SCLT1</td>
<td>rs1816429</td>
<td>130347005</td>
<td>G (0.21)</td>
<td>1.80 (0.38)</td>
<td>$2.58 \times 10^{-6}$</td>
</tr>
<tr>
<td>18</td>
<td>B4GALT6</td>
<td>Intragenic</td>
<td>rs1667284</td>
<td>29226208</td>
<td>A (0.14)</td>
<td>2.20 (0.47)</td>
<td>$3.63 \times 10^{-6}$</td>
</tr>
<tr>
<td>8</td>
<td>Intergenic</td>
<td>151 kb, ST3GAL1</td>
<td>rs7843718</td>
<td>134735220</td>
<td>G (0.18)</td>
<td>1.86 (0.41)</td>
<td>$5.05 \times 10^{-6}$</td>
</tr>
<tr>
<td>20</td>
<td>PIPRT</td>
<td>Intragenic</td>
<td>rs11086862</td>
<td>41754601</td>
<td>C (0.15)</td>
<td>2.00 (0.44)</td>
<td>$5.14 \times 10^{-6}$</td>
</tr>
<tr>
<td>18</td>
<td>Intergenic</td>
<td>15 kb, TIR</td>
<td>rs1375445</td>
<td>29156999</td>
<td>C (0.26)</td>
<td>1.64 (0.36)</td>
<td>$6.27 \times 10^{-6}$</td>
</tr>
<tr>
<td>18</td>
<td>B4GALT6</td>
<td>Intragenic</td>
<td>rs1791161</td>
<td>29204210</td>
<td>G (0.15)</td>
<td>2.04 (0.45)</td>
<td>$6.42 \times 10^{-6}$</td>
</tr>
</tbody>
</table>

SNPs on chromosome 4 are in linkage disequilibrium (shown in Fig. 1). Abbreviations: Chr, chromosome; BP, base-pair position; SE, standard error; MAF, minor allele frequency; SNP, single-nucleotide polymorphism.

*Bonferroni-adjusted significance in the additive model, genome-wide significant in the dominant model [$\beta (SE) = 2.69 (0.48); P = 2.61 \times 10^{-8}$]*
### Table 2

Sample characteristics for discovery and extension samples for association between rs1433375 and total alcohol use disorders identification test (AUDIT) score.

<table>
<thead>
<tr>
<th>Discovery cohort</th>
<th>Gender</th>
<th>Total sample (N)</th>
<th>MAF</th>
<th>β</th>
<th>SE</th>
<th>P</th>
</tr>
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<tbody>
<tr>
<td>Grady Trauma Project</td>
<td>Male: 373</td>
<td>Female: 663</td>
<td>1036</td>
<td>A (0.17)</td>
<td>2.21(^a)</td>
<td>0.41</td>
</tr>
</tbody>
</table>

**Additional Cohorts**

**Marine Resiliency Study**

<table>
<thead>
<tr>
<th></th>
<th>Gender</th>
<th>Total sample (N)</th>
<th>MAF</th>
<th>β</th>
<th>SE</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>European American</td>
<td>790</td>
<td>—</td>
<td>790</td>
<td>G (0.42)</td>
<td>-0.40(^b)</td>
<td>0.24</td>
</tr>
<tr>
<td>African American</td>
<td>82</td>
<td>—</td>
<td>82</td>
<td>G (0.70)</td>
<td>-0.98(^b)</td>
<td>1.08</td>
</tr>
<tr>
<td>Hispanic, Native American</td>
<td>236</td>
<td>—</td>
<td>236</td>
<td>G (0.39)</td>
<td>-1.04(^b)</td>
<td>0.38</td>
</tr>
<tr>
<td>Other</td>
<td>167</td>
<td>—</td>
<td>167</td>
<td>G (0.46)</td>
<td>-0.27(^b)</td>
<td>0.58</td>
</tr>
</tbody>
</table>

**Systems Biology PTSD Biomarkers Consortium**

<table>
<thead>
<tr>
<th></th>
<th>Gender</th>
<th>Total sample (N)</th>
<th>MAF</th>
<th>β</th>
<th>SE</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hispanic</td>
<td>39</td>
<td>7</td>
<td>46</td>
<td>A (0.5)</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>Non-Hispanic Asian</td>
<td>6</td>
<td>0</td>
<td>6</td>
<td>A (0.42)</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>Non-Hispanic Black</td>
<td>23</td>
<td>3</td>
<td>26</td>
<td>A (0.19)</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>Non-Hispanic Other</td>
<td>3</td>
<td>0</td>
<td>3</td>
<td>A (0.13)</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>Non-Hispanic White</td>
<td>34</td>
<td>4</td>
<td>38</td>
<td>G (0.46)</td>
<td>—</td>
<td>—</td>
</tr>
</tbody>
</table>

Note that in all cohorts the A allele is positively associated with the AUDIT score and the ‘G’ allele is negatively associated with the AUDIT score.

\(^a\) Reference allele A.

\(^b\) Reference allele ‘G’.