Mycobacterium abscessus subspecies massiliense infection after skin graft and cholecystectomy in a burn patient

Nathan A. Summers, Emory University
Russell Ryan Kempker, Emory University
Federico Palacio Bedoya, Emory University

Journal Title: International Journal of Infectious Diseases
Volume: Volume 76
Publisher: Elsevier: Creative Commons Attribution Non-Commercial No-Derivatives License | 2018-11-01, Pages 29-31
Type of Work: Article | Final Publisher PDF
Publisher DOI: 10.1016/j.ijid.2018.08.016
Permanent URL: https://pid.emory.edu/ark:/25593/tn4zd

Final published version: http://dx.doi.org/10.1016/j.ijid.2018.08.016

Copyright information:
© 2018 The Authors
This is an Open Access work distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Accessed August 1, 2019 11:44 PM EDT
Short Communication

**Mycobacterium abscessus** subspecies *massiliense* infection after skin graft and cholecystectomy in a burn patient

Nathan A. Summers*, Russell Kempker, Federico Palacio

Division of Infectious Diseases, Department of Medicine, Emory University School of Medicine, Atlanta, GA, USA

**A R T I C L E   I N F O**

Article history:
Received 31 July 2018
Received in revised form 20 August 2018
Accepted 21 August 2018
Corresponding Editor: Eskild Petersen, Aarhus, Denmark

**Keywords:**
Rapidly growing mycobacteria
Burn
Postoperative infection
*Mycobacterium massiliense*

**A B S T R A C T**

Diagnosing skin and soft tissue infections due to rapidly growing mycobacteria (RGM) can often prove difficult, leading to delays in treatment. Postoperative infections caused by RGM are increasingly recognized both within and outside the USA, but are rarely encountered in burn units. We report a case of postoperative skin and soft tissue infection along a cholecystectomy incision in a burn patient caused by *Mycobacterium abscessus* subspp. *massiliense*. Postoperative infections caused by RGM require a high index of suspicion, often necessitating biopsy for definitive diagnosis. Physicians should consider this diagnosis when postoperative infections arise later than typically seen for routine bacterial infections and fail to respond to first-line therapy.

© 2018 The Authors. Published by Elsevier Ltd on behalf of International Society for Infectious Diseases. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

**Introduction**

*Mycobacterium abscessus* subspp. *massiliense* is a rapidly growing Mycobacterium (RGM) that was first described when *M. abscessus* was divided into three different species in 2006: *M. abscessus* sensu stricto, *Mycobacterium massiliense*, and *Mycobacterium bolletii*. Multilocus sequence typing in 2011 then reclassified these three organisms as subspecies within the *M. abscessus* species, which was later supported by whole-genome sequencing in 2013 and matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF MS) in 2014 (Mougari et al., 2016).

The most commonly encountered RGM isolated within the USA include *M. abscessus*, *Mycobacterium fortuitum*, and *Mycobacterium chelonae*, which account for up to 90% of the infections caused by RGM (Mougari et al., 2017). By definition, these mycobacteria grow on solid media within 7 days, confirmed by repeating the culture on subculture to avoid falsely rapid growth because of a large inoculum size. These organisms are ubiquitous and are readily isolated in the environment and from non-sterile water sources, and healthcare-associated infections due to RGM are increasingly recognized (Brown-Elliott and Wallace, 2015; Petrini, 2006).

**Case report**

The case patient was a 55-year-old previously healthy woman who had sustained partial and full-thickness burns involving 45% of her body surface area 6 months prior, when her grill had exploded. After a prolonged hospitalization requiring several excisional skin and soft tissue debridement procedures with cadaveric and epithelial autologous skin grafts, she underwent an open cholecystectomy 1 month before transfer to our hospital.

Shortly after, the patient developed a rash with pink-to-red papules and nodules coalescing into plaques in a linear distribution along her incision and involving her autologous skin grafts (Figure 1). A biopsy revealed granulomatous inflammation with giant cells; staining failed to identify any pathogens (Figures 2 and 3). Cultures grew on Middlebrook 7H11 agar in 3 days, suggesting RGM.

The patient was started on cefoxitin, clarithromycin, and moxifloxacin empirically. Species identification was performed with molecular assays (including *rpoB* gene and 16S rRNA sequencing) and antimicrobial susceptibility testing was performed by microdilution assays. The isolate was susceptible to azithromycin, clarithromycin, cluzafamize, kanamycin, and tigecycline, intermediate to cefoxitin, and resistant to all other drugs tested. The patient improved and was discharged on clarithromycin and moxifloxacin for a 6-month course.

**Discussion**

Skin and soft tissue infections due to the *M. abscessus* group typically occur after accidental trauma or surgery and can occur in

https://doi.org/10.1016/j.ijid.2018.08.016
1201-9712/© 2018 The Authors. Published by Elsevier Ltd on behalf of International Society for Infectious Diseases. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/)
bacterial infections but fail to respond to first-line therapy (Griffith et al., 2007).

Clusters of postoperative infections have arisen after a myriad of surgical procedures. A total of 21 cases of postoperative wound infections caused by RGM were described following cosmetic surgeries performed in the Dominican Republic in 2013–2014, with the majority growing M. abscessus. These infections were considered to have occurred due to the use of a contaminated water supply (Schnabel et al., 2016). Outbreaks have also been seen in the USA with post-injection joint infections secondary to failure of the disinfectant benzalkonium chloride (Tiwari et al., 2003), as well as following laser-assisted in situ keratomileusis due to the use of non-sterile water in a misting humidifier (Edens et al., 2015). Recently a series of cases of RGM port-site infections were described following laparoscopic surgery in India. Of the 32 patients identified with chronic port-site infections, 20 had acid-fast bacilli (AFB) identified on staining and 15 of these grew RGM (13 were M. abscessus, two were M. fortuitum); the most commonly performed procedure was a cholecystectomy (Ghosh et al., 2017).

RGM are infrequently encountered among patients in burn units. One case report describes a case of M. abscessus bacteremia in a patient in a burn unit (Vaghaiwalla et al., 2014). A recent case series reviewed over 2428 patients with thermal burns admitted to the United States Army Institute burn center and found only two patients who had RGM isolated: M. abscessus in a patient’s tracheal aspirate and blood cultures and M. fortuitum in a patient’s sputum (Boyer et al., 2010). In a case series describing RGM infections after a tsunami struck Thailand in 2004, RGM were isolated from 15 patients, nine of whom had skin grafts (Appelgren et al., 2008). These cases were not of clonal descent so were felt to be environmental contamination acquired during initial injury from the tsunami itself, rather than from contaminated healthcare exposure. The case presented herein appears to be the first published report of a burn patient developing a RGM skin and soft tissue infection at the site of a skin graft and cholecystectomy incision.

M. abscessus species possess inherent and acquired resistance to many commonly used antimicrobials (Nessar et al., 2012). Although macrolides are the mainstay of therapy, acquired resistance to this drug class can rapidly occur due to the presence of the inducible erm(41) gene, encoding a 23S methylase (Mougari et al., 2017; Nessar et al., 2012). M. massiliense is unique among the M. abscessus group in that it lacks an effective erm(41) due to deletions within the gene. However, resistance to macrolides can

...
still develop through other mutations, most commonly due to mutations in the rrl gene, which results in a mutant 23S rRNA (Maurer et al., 2012; Mougari et al., 2017). Therefore, current American Thoracic Society (ATS)/Infectious Diseases Society of America (IDSA) guidelines recommend combination therapy consisting of a macrolide with two additional agents, generally amikacin, cefoxitin, or imipenem. Other agents with potential efficacy include linezolid, tigecycline, and telithromycin, but these have not been studied extensively. It is recommended that combination antimicrobial therapy is continued for a minimum of 4 months to avoid the development of drug resistance, with surgical debridement recommended if abscesses or foreign bodies are present, or if there is extensive disease (Griffith et al., 2007).

In summary, RGM are increasingly recognized as causes of postoperative infections, and outbreaks are being seen both within and outside the USA (Edens et al., 2015; Ghosh et al., 2017; Schnabel et al., 2016; Tiwari et al., 2003). The presence of a chronic draining abscess or plaque, onset of symptoms later than what is typically seen for routine bacterial postoperative infections, and failure to respond to first-line antibiotics for bacterial infections should raise concern that a RGM may be present. Clinicians should keep a high index of suspicion in these cases and pursue skin biopsy early when skin and soft tissue infections fail to improve on empirical therapy. It is also important to have close communication with the microbiology laboratory to ensure a proper workup is done, including identification to the species level to guide appropriate antimicrobial therapy.

Acknowledgements

The authors would like to thank Dr Emily Cole from the Department of Dermatology, Emory University, for providing the images used in this publication.

Financial support

N.A.S. is supported in part by the National Center for Advancing Translational Sciences of the National Institutes of Health (grant numbers UL1 TR002378, TL1 TR002382).

Ethical approval

None required.

Conflict of interest

None declared.

References