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Effects of Single and Combined Water, Sanitation and Handwashing Interventions on Fecal Contamination in the Domestic Environment: A Cluster-Randomized Controlled Trial in Rural Bangladesh

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*Supporting Information

ABSTRACT: Water, sanitation, and hygiene interventions have varying effectiveness in reducing fecal contamination in the domestic environment; delivering them in combination could yield synergies. We conducted environmental assessments within a randomized controlled trial in Bangladesh that implemented single and combined water treatment, sanitation, handwashing (WSH) and nutrition interventions (WASH Benefits, NCT01590095). After one and two years of intervention, we quantified fecal indicator bacteria in samples of drinking water (from source or storage), child hands, children’s food and sentinel objects. In households receiving single water treatment interventions, Escherichia coli prevalence in stored drinking water was reduced by 50% and concentration by 1-log. E. coli prevalence in food was reduced by 30% and concentration by 0.5-log in households receiving single water treatment and handwashing interventions. Combined WSH did not reduce fecal contamination more effectively than its components. Interventions did not reduce E. coli in groundwater, on child hands and on objects. These findings suggest that WSH improvements reduced contamination along the direct transmission pathways of stored water and food but not along indirect upstream pathways. Our findings support implementing water treatment and handwashing to reduce fecal exposure through water and food but provide no evidence that combining interventions further reduces exposure.

BACKGROUND

Diarrheal disease transmission occurs through a complex web of environmentally mediated pathways including drinking water, food, hands, fomites, and vectors. The complexity arises from the multitude of transmission routes, broad diversity of pathogens, importance of environmental conditions, and interactions between the environment and human behavior. Treating drinking water before consumption and washing hands with water and soap lower fecal contamination of drinking water and hands, respectively, and reduce reported diarrhea.1–4 However, hands are rapidly recontaminated by contact with objects and surfaces5–7 and treated water can be recontaminated by hands and utensils during storage.8 Sanitation interventions target reducing contamination further upstream by isolating fecal matter from the ambient surroundings but sanitation improvements to date have generally not reduced fecal indicators in the environment and shown mixed impact on health.9–12 Combining water, sanitation, and hygiene improvements could yield synergistic benefits. Coupling water treatment and

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safe storage with a hygiene intervention could reduce recontamination of treated water through cleaner hands, whereas combining handwashing with sanitation improvements could reduce recontamination of washed hands through reduced exposure to fecal matter in the environment. Water, sanitation, and hygiene interventions could also work in concert to reduce food contamination by making treated water available for food preparation, facilitating handwashing before food handling and reducing breeding sites for flies transmitting pathogens to stored food.13,14 No studies have investigated whether combined water, sanitation, and hygiene interventions more effectively reduce contamination of drinking water, food, hands and fomites than individual interventions. We conducted an environmental assessment within a randomized controlled trial in Bangladesh (WASH Benefits, ClinicalTrials.gov NCT01590095) to assess whether (1) water, sanitation and hygiene interventions vs controls and (2) combined vs single water, sanitation, and hygiene interventions reduce fecal indicator bacteria and flies in the domestic environment.

**MATERIALS AND METHODS**

**Study Design.** WASH Benefits enrolled participants in four districts (Gazipur, Kishoreganj, Mymensingh, Tangail) in rural Bangladesh. These areas were selected because their groundwater chemistry was suitable for the trial’s chlorine-based water treatment intervention15 and because they had no other major ongoing or planned water, sanitation, and hygiene programs. The trial enrolled pregnant women identified by screening the study area. Using global positioning system (GPS) coordinates, eight adjacent eligible women were grouped into clusters, and each eight clusters formed a block. Clusters were block-randomized into study arms by an off-site investigator (BFA), providing pair-matched randomization. WASH Benefits followed the birth cohort born to the enrolled pregnant women (“index children”) for two years. Further details of the trial design have been previously described.16 The primary outcomes of WASH Benefits were child diarrhea and growth, and additional outcomes included protozoan and soil-transmitted helminth (STH) infections; these have been reported separately.17−19 Measures of environmental contamination were prespecified intermediate outcomes.16

The trial had six intervention arms including single and combined water, sanitation, handwashing, and nutrition interventions and a double-sized control arm receiving no intervention. The intervention arms included (1) water treatment: point-of-use water treatment with sodium dichloroisocyanurate (NaDCC, Aquatabs) (Medentech, Wexford, Ireland) and safe storage in a narrow-mouth, lidded container with spigot, (2) sanitation improvements: upgrades to concrete-lined double-pit latrines, and provision of child potties and scoops for feces disposal, (3) handwashing promotion: provision of handwashing stations in the kitchen and latrine areas with a water reservoir, a bottle of soapy water mixture and a basin for rinsewater (4) nutrition improvements: provision of lipid-based nutrient supplements for children aged 6−24 months, recommendations for exclusive breastfeeding for children aged up to 6 months and age-appropriate nutrition recommendations from pregnancy through 24 months of age, (5) combined water, sanitation and handwashing (WSH), and (6) nutrition plus combined WSH (N+WSH).

Community health promoters hired from among local women and trained specifically for the study visited intervention households six times per month on average to demonstrate and encourage the targeted behaviors (e.g., water treatment, handwashing at critical times, latrine use for defecation) and supply intervention products for free throughout the trial period. Control households did not receive any interventions or health promoter visits. User adherence to the interventions was measured through spot-check and structured observations in unannounced household visits (as an independent investigation from the study activities...
These demonstrated high adherence throughout the trial, including hardware availability observed by spot-checks and user practices assessed by structured observations. Spot-checks indicated >95% of households in arms receiving the sanitation intervention had a latrine with a water seal compared to 23% of controls, and >77% of households in arms receiving the handwashing intervention had water and soap present <6 steps from the kitchen or latrine compared to 21% of controls. Structured observations showed in >94% of households in sanitation arms adults used a hygienic latrine for defecation (vs 40% of controls), in >67% of households in handwashing arms participants washed hands with soap after using the latrine (vs 29% of controls) and in >65% of households in water treatment arms participants drank chlorine-treated water from a safe storage container (vs none of controls). Further details of user adherence have been described.

**Procedures.** A subset of trial participants in the control, nutrition, WSH and N+WSH arms was enrolled in an environmental enteric dysentery (EED) substudy. EED enrollment was based on proximity to the laboratory and therefore (unlike the parent trial) not geographically pair-matched. We conducted an environmental assessment among EED substudy participants. This allowed us to assess the environmental impact of the combined WSH intervention by comparing pooled data from the WSH and N+WSH arms (both receiving the WSH package, referred to as “WSH arm” hereinafter) to pooled data from the control and nutrition arms (neither one receiving the WSH intervention, referred to as “control arm” hereinafter). We also sampled a random subset of households in the single water and handwashing arms to assess the impact of combined vs individual interventions. The sanitation arm was not sampled as a previous assessment showed no early environmental impact in this arm and a separate longitudinal study is underway to assess the long-term impact of sanitation on environmental contamination (manuscript forthcoming). Sampling was conducted at two time points, approximately 1 and 2 years after intervention initiation, to match the timing of the trial’s health outcome measurements (see Supporting Information (SI) Text S1 and Figure S1 for details of all environmental assessments nested within WASH Benefits).

Trained field workers from the International Centre for Diarrhoeal Disease Research, Bangladesh (icddr,b) visited households enrolled in the environmental assessment to collect samples. In the control and WSH arms, we quantified fecal indicator bacteria in drinking water, on index child hands, on toy ball “sentinel objects” and (in year two only) in food served to young children (Figure 1). Sentinel objects are presterilized objects (e.g., toy ball) left in the household to take up contamination from the domestic environment while household members interact with it and then tested for fecal indicator bacteria. They serve as a measure of overall contamination of domestic surfaces and objects, and have been shown to distinguish households with vs without improved water, sanitation, and hygiene conditions in Bangladesh. In single intervention arms, we only collected a relevant subset of sample types expected to be directly affected by the interventions; we sampled drinking water and (in year two) children’s food in the water arm, and index child hands and (in year two) children’s food in the handwashing arm. Additionally, field staff examined caregiver and index child hands (fingernails, fingerpads and palms) in the control, handwashing and WSH arms for visible dirt; observed dirt on hands has been validated as a proxy for handwashing and shown to correlate with bacterial counts on hands. Field staff also enumerated and speciated synanthropic flies in the kitchen and latrine areas in the control and WSH arms.

**Sample Collection.** Samples were collected using sterile Whirlpak bags (Nasco Modesto, Salida, CA). Clean gloves were worn while collecting hand rinse, toy rinse, and food samples. To collect drinking water, field workers asked the respondent to provide a glass of water in the same manner they would give it to their children and pour approximately 150 mL into the Whirlpak; field workers recorded if the water was provided from the source (tubewell) or from a storage container. If the respondent reported using chlorine, sodium thiosulfate was used to neutralize residual chlorine, and an additional sample was collected to measure the free chlorine residual with a digital colorimeter (Hach Pocket Colorimeter II). To sample child hands, field workers asked the respondent to place the index child’s left-hand into a Whirlpak prefilled with 250 mL of distilled water. The hand was massaged from outside the bag for 15 s and shaken for 15 s. The procedure was repeated with the right-hand in the same bag, and the rinsewater was preserved in Whirlpak. To sample sentinel objects, field workers delivered presterilized nonporous plastic toy balls to participants and instructed them to let the index child as well as other children in the compound play freely with the ball. They returned 24 h later to rinse the ball in a Whirlpak prefilled with 250 mL of distilled water by massaging it from outside the bag and shaking the bag, for 15 s each. The rinsewater was preserved for analysis and the toy was left at the household; a new ball was delivered at the time of the year two sampling. To sample food, field workers identified stored food prepared to be served to children <5 years and asked the respondent to provide a small amount in the same manner they would serve it to their children. They prioritized sampling rice if available. Food was scooped from the dish that it was provided in into a 50 mL sterile tube using a sterile spoon. Samples were transported to the icddr,b field laboratory on ice at 2–8 °C and analyzed within 12 h of collection.

**Sample Processing.** We used membrane filtration to analyze water samples (undiluted) and hand and toy rinse samples (both undiluted and with 1:10 and 1:100 dilutions). 100 mL aliquots were filtered through a 0.45-μm cellulose filter and incubated for 24 h on 60 mm MI agar plates (BD Difco, Franklin Lakes, NJ). Water and hand rinse samples were incubated at 35 °C to enumerate *Escherichia coli* and total coliforms following standard protocols. We modified the protocol to incubate toy rinse samples at 44.5 °C to enumerate *E. coli* and fecal/thermotolerant coliforms as a previous study found that fecal (rather than total) coliforms on toy balls were a more sensitive indicator of environmental contamination.

We used the pour-plate technique to analyze food samples. 10 g food aliquots were homogenized with 40 mL of distilled water. The homogenate was analyzed both undiluted and with 1:100 dilution. A 2.5 mL aliquot of homogenate and 15 mL of TBX media (Sigma-Aldrich, St. Louis, MO) were added to a 100 mm Petri dish by pour-plate and incubated for 24 h at 44.5 °C to enumerate *E. coli* following standard protocols. An additional 5 g food aliquot was oven-dried overnight to determine moisture content.

Counts were expressed in colony forming units (CFU) (per 100 mL for water samples, per 2 hands for child hands, per 1 toy for sentinel toys and per 1 dry gram for food). Plates with
200 colonies for water, hand rinses and toy rinses and >500 colonies for food were classified as too numerous to count (TNTC). The higher detection limit for the food samples reflects the larger surface area of the 100 mm plates, allowing a higher number of colonies to be visually distinguished (see SI Table S1 for detection limits for each sample type).

Quality Control. One laboratory control per analyst per day and 5% replicates (repeat aliquots from same Whirlpak for every 20th sample) were processed. Field workers collected 10% field blanks (one blank for every 10 samples) by asking respondents to pour distilled water from a sterile bottle into a Whirlpak as if collecting a water sample, by opening and massaging a prefilled Whirlpak as if sampling a hand, and by rinsing a presterilized toy ball in prefilled Whirlpak as if sampling a toy.

Fly Counts. To enumerate flies, field workers identified a suitable location in the kitchen and latrine areas (away from the stove smoke, under a roof or protected from rain) and horizontally hung three 1.5-foot strips of nonbaited sticky fly tape. They returned to the household 24 h later to count the captured synanthropic flies and speciate them according to a visual identification chart.

Ethics. Participants provided written informed consent in the local language (Bengali). The study protocol was approved by human subjects committees at the icddr,b (PR-11063), University of California, Berkeley (2011−09−3652), and Stanford University (25863).

Statistical Methods. We prespecified and registered our analysis plan on Open Science Framework (https://osf.io/6u7cn/).

Sample Size. The EED substudy, within which the environmental sampling was nested, targeted 1500 households (375/arm) and enrolled approximately 2000 households in year one to allow for attrition by year two. In addition, we enrolled 180 water arm households and 360 handwashing arm households in the environmental assessment. We obtained measures of contamination and intraclass correlation coefficients from the literature and unpublished pilot data (see analysis plan). We used a one-sided α of 0.05, assuming that the interventions would decrease but not increase contamination. Our sample size yielded 80% power to detect a 0.20 log10 reduction in E. coli concentration in drinking water, hand rinses and toy rinses at each sampling time point, compared to controls.
**Parameters of Interest.** Our outcomes were (1) prevalence and concentration of *E. coli* and total/fecal coliforms, (2) prevalence and number of flies near the kitchen and latrine, and (3) prevalence of caregiver and child hands with visible dirt. Our parameters were prevalence ratios (PR) and differences (PD) for binary measures, log_{10} reductions for *E. coli* and coliform concentrations, and fly count ratios for the number of flies. We substituted bacterial counts with half the lower detection limit for nondetects, 200 CFU for TNTC water, hand rinse and toy rinse samples and 500 CFU for TNTC food samples.\textsuperscript{34} Means for multiple dilutions were obtained by dividing the sum of plate counts by the total sample volume filtered for all countable plates.

**Estimation Strategy.** We compared all intervention arms to controls and the combined WSH arm to the single water and handwashing arms. Analyses were intention-to-treat; this preserves the randomization and is appropriate given the high intervention adherence.\textsuperscript{20} Randomization balanced covariates across arms.\textsuperscript{17} Therefore, we relied on unadjusted estimates in our analysis. We estimated unadjusted parameters using generalized linear models with robust standard errors, and a log link for PRs, linear link for PDs and log_{10} reductions, and log link allowing for overdispersion (negative binomial regression) for fly count ratios. Secondary analyses adjusted for prespecified covariates using doubly robust targeted maximum likelihood estimation (TMLE) incorporating an ensemble machine learning method called Super Learner.\textsuperscript{35,36} We conducted separate comparisons at each of the two measurement rounds and performed a Bonferroni correction for multiple measurements by multiplying the p-values for effect estimates by two. We conducted separate analyses stratifying by whether the sampled drinking water came from the source or storage container. We assessed effect modification by season by including an interaction term for wet vs dry season; Bangladesh has a monsoon season (June–October), during which it receives >80% of its rain\textsuperscript{37} and environmental contamination levels typically increase.\textsuperscript{38}

**RESULTS AND DISCUSSION**

**Enrollment.** Of 2445 households selected for EED enrollment in year one, we enrolled 1980 (81%) in the environmental assessment between October 2013 and December 2014; we also enrolled 181 water arm households.
and 366 handwashing arm households. At year two, among 1515 households successfully enrolled in the EED substudy, we enrolled 1363 (90%) in the environmental assessment between May 2015 and May 2016; we also enrolled 184 water arm households and 356 handwashing arm households. Reasons for households not being enrolled were no live birth or index child death (10% in year one, 0.3% in year two), absence or relocation (7% in year one, 4% in year two) and refusal (2% in year one, 5% in year two). Covariates were balanced between arms among enrolled households (SI Table S2).

**Observed Hand Cleanliness.** Among caregivers in the control arm, dirt was observed on 80% of nails and 33–34% of fingertips and palms at year one, and 50% of nails and 15% of fingertips and palms at year two (Figures 2 and 3). Among children in the control arm, dirt was observed on 89% of nails and 62–66% of fingertips and palms at year one (mean child age: 14 months), and 66% of nails and 41–43% of fingertips and palms at year two (mean child age: 30 months); these ages roughly coincided with WHO windows for rolling, crawling and learning to walk (5–18 months) vs walking well (>18 months). Caregivers in the handwashing arm were substantially less likely to have dirt on fingertips and palms but not under fingernails at both time points (Figures 2 and 3). Similarly, children in the handwashing arm were less likely to have dirt on fingertips and palms and, to a smaller extent, under fingernails (Figures 2 and 3). Caregivers or children in the WSH arm were no less likely to have dirt on their hands than controls at either time point (Figures 2 and 3).

**Fecal Contamination.** Levels of contamination. We analyzed 6409 samples at year one and 6127 at year two. Of drinking water samples, 29% were provided directly from a tubewell and 71% from a storage container (in arms receiving the water intervention, this was predominantly the provided safe storage container). Among controls, we detected *E. coli* in 59–62% of tubewells, 90–93% of stored drinking water, 96–
98% of child hands and 76–81% of toys at the two time points and 55% of children’s food at year two (Figures 4 and 5).

**Intervention Effects.** Among households receiving water treatment interventions, 90% of stored drinking water samples were reported to be chlorinated, and of these, 80% had detectable (>0.1 mg/L) free chlorine residual at the two time points, while 1% of controls reported water treatment. Compared to controls, stored drinking water in the single water arm had approximately 50% reduction in *E. coli* prevalence and 1-log reduction in *E. coli* concentration at both time points (Yr1: prevalence ratio [PR] = 0.53 (0.44, 0.63), \( \Delta \log_{10} = 0.13 (-0.09, -0.23) \); Yr2: PR = 0.51 (0.43, 0.62), \( \Delta \log_{10} = 0.15 (-0.13, -0.27) \); all Bonferroni-corrected \( p < 0.001 \) (Figures 4 and 5, SI Tables S3 and S4). Compared to controls, stored drinking water in the WSH arm also had similar reductions in *E. coli* prevalence and concentration at both time points (Yr1: PR = 0.63 (0.58, 0.68), \( \Delta \log_{10} = -0.83 (-0.79, -0.86) \); Yr2: PR = 0.65 (0.59, 0.70), \( \Delta \log_{10} = -0.83 (-0.8, -0.77) \); all Bonferroni-corrected \( p < 0.001 \) (Figures 4 and 5, SI Tables S3 and S4). Compared to controls, food (measured only in year two) had approximately 30% reduction in *E. coli* prevalence in the water arm (PR = 0.70 (0.57, 0.86)) and handwashing arm (PR = 0.68 (0.56, 0.83)) and a borderline reduction in the WSH arm (PR = 0.89 (0.78, 1.01)). Similarly, food *E. coli* counts were reduced by about 0.5-log in the water arm (\( \Delta \log_{10} = -0.42 (-0.72, -0.12) \)) and handwashing arm (\( \Delta \log_{10} = -0.52 (-0.79, -0.26) \)) and borderline reduced in the WSH arm (\( \Delta \log_{10} = -0.18 (-0.40, 0.05) \)) compared to controls (Figure 5, SI Table S4). Tubewell water, child hands and sentinel toys had no reductions in *E. coli* prevalence or concentration in intervention arms vs controls at either time point (Figures 4 and 5, SI Tables S3 and S4). Compared to controls, *E. coli* counts were borderline increased on child hands in the handwashing arm and on toys in the WSH arm at year one but not at year two (Figures 4 and 5, SI Tables S3 and S4).

Comparing combined vs individual interventions, the WSH arm had higher stored water *E. coli* prevalence and concentration compared to controls, food (measured only in year two) had approximately 30% reduction in *E. coli* prevalence in the water arm (PR = 0.70 (0.57, 0.86)) and handwashing arm (PR = 0.68 (0.56, 0.83)) and a borderline reduction in the WSH arm (PR = 0.89 (0.78, 1.01)). Similarly, food *E. coli* counts were reduced by about 0.5-log in the water arm (\( \Delta \log_{10} = -0.42 (-0.72, -0.12) \)) and handwashing arm (\( \Delta \log_{10} = -0.52 (-0.79, -0.26) \)) and borderline reduced in the WSH arm (\( \Delta \log_{10} = -0.18 (-0.40, 0.05) \)) compared to controls (Figure 5, SI Table S4). Tubewell water, child hands and sentinel toys had no reductions in *E. coli* prevalence or concentration in intervention arms vs controls at either time point (Figures 4 and 5, SI Tables S3 and S4). Compared to controls, *E. coli* counts were borderline increased on child hands in the handwashing arm and on toys in the WSH arm at year one but not at year two (Figures 4 and 5, SI Tables S3 and S4).

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concentration than the water arm at year two (PR = 1.26 (1.05, 1.50), Δlog₁₀ = 0.23 (0.09, 0.36), both Bonferroni-corrected p < 0.05); there was a similar albeit borderline nonsignificant pattern at year one (SI Tables S3 and S4). The WSH arm also had higher food E. coli prevalence and concentration than the water arm (PR = 1.27 (1.02, 1.57), Δlog₁₀ = 0.24 (−0.03, 0.52)) and handwashing arm (PR = 1.30 (1.08, 1.55), Δlog₁₀ = 0.34 (0.12, 0.57)) (SI Tables S3 and S4). There were no differences in E. coli prevalence or concentration in the other sample types between combined and individual intervention arms.

Secondary analyses adjusting for confounders yielded similar results except that food E. coli reductions were slightly attenuated (SI Tables S3 and S4). Subgroup analyses suggested overall lower contamination and larger reductions in E. coli in stored drinking water and food from the interventions during the dry season; there were no other seasonal effects (SI Tables S5 and S6). Total/fecal coliforms showed patterns similar to E. coli (SI Tables S7 and S8).

**Quality Control.** Across sample types and sampling rounds, 23% of samples were nondetect and 6% exceeded upper detection limits. Intraclass correlation between replicates was >85% at both time points. E. coli was detected in 2.5% of blanks and the geometric mean E. coli count among positive blanks was 7 CFU. Repeating the analyses after removing the data from days with contaminated blanks did not change findings (SI Tables S9 and S10).

**Fly Presence.** At year one, at least 1 fly was captured near the kitchen in 57% and near the latrine in 56% of control households; the mean number of flies was 6.59 (range: 1−201) near the kitchen and 3.00 (range: 1−229) near the latrine. At year two, at least 1 fly was captured near the kitchen in 45% and near the latrine in 40% of control households; the mean number of flies was 3.32 (range: 1−86) near the kitchen and 1.60 (range: 1−98) near the latrine. The predominant fly species was the common housefly (Musca domestica). Compared to controls, fly prevalence was reduced significantly in the WSH arm at year one, both near the kitchen (PR = 0.83 (0.75, 0.94), Bonferroni-corrected p < 0.005) and the latrine (PR = 0.81 (0.74, 0.89), Bonferroni-corrected p < 0.001), but not at year two (Figures 4 and 5). There was no consistent impact on fly numbers at either time point (Figures 4 and 5).

**Implications.** Water, sanitation, and handwashing interventions reduced fecal exposure along the direct transmission pathways of stored drinking water and food but not along the indirect, more upstream pathways of child hands, household objects and groundwater sources. Chlorine and safe storage effectively reduced E. coli in stored water, consistent with prior evidence.

E. coli in food was reduced in the water and handwashing arms. This could indicate that participants used treated water to prepare food and rinse utensils in the water arm and improved their handwashing around food handling in the handwashing arm. In structured observations, 5% of caregivers in the handwashing arm washed hands specifically before preparing food vs 0.5% of controls, suggesting hand hygiene around food handling remained poor. However, overall handwashing practices improved; in structured observations, 67% of caregivers in the handwashing arm washed hands after using the latrine vs 29% of controls. This supports our observation that caregivers in the handwashing arm were significantly less likely to have visible dirt on fingertips and palms (but not nails) than controls. The hand observation method has the caveat that field staff could over perceive hand cleanliness in the handwashing arm as they were not blinded to the intervention assignment (blinding was not feasible since the interventions had distinct hardware components). However, the staff perceived no such effect in the WSH arm, suggesting no blanket bias from knowledge of treatment status.

Similarly, children in the handwashing arm had less dirt on fingertips and palms, and slightly less dirt under nails. As the children were too young to wash their own hands, this might indicate that improved hygiene among caregivers translated into washing/wiping child hands. However, the interventions did not reduce fecal bacteria on child hands. This could be because our sampling method of massaging and vigorously rinsing hands might have eluted dirt from under fingernails that was not removed by washing/wiping. Our previous work found that young children in our study population frequently touch soil, which is highly contaminated in this setting. Soil trapped under nails might have harbored sufficient E. coli to undermine the effect of cleaner fingers and palms; fingernails have been shown to harbor pathogens such as parasite eggs and larvae in Bangladesh.

The interventions did not reduce overall contamination in the ambient domestic environment, as measured by sentinel toys and groundwater samples. This is consistent with an earlier assessment among study households that found no difference in fecal contamination in the ambient environment (courtyard soil, ponds, groundwater) in the sanitation and combined WSH arms after approximately 4 months of intervention. Our findings differ from two studies in rural Bangladesh that found fewer fecal coliforms on sentinel toys in compounds with better water, sanitation and hygiene infrastructure. However, these studies used cross-sectional designs susceptible to confounding; compounds with improved sanitary conditions could have other characteristics leading to reduced contamination. Toy balls in our study were often coated in soil; the lack of E. coli reduction on toys is consistent with the lack of intervention impact on E. coli in soil in our previous assessment.

Our findings also suggest that single interventions achieved larger reductions in contamination than the combined WSH package. The water arm had less E. coli in stored water than the WSH arm, and the water and handwashing arms had less E. coli in food than the WSH arm. Similarly, children and caregivers in the handwashing but not the WSH arm were less likely to have visible dirt on hands. One explanation could be that the multicomponent package diffused the effectiveness of any one intervention; that is, targeting a specific behavior was more effective than attempting to change multiple behaviors. However, spot-checks and structured observations revealed only minor differences in uptake indicators between combined and single intervention arms.

**Environmental Findings in the Context of the Trial’S Health Outcomes.** WASH Benefits measured enteric infections in children including (1) caregiver-reported diarrhea, (2) protozoan infections with Cryptosporidium spp., Giardia duodenalis, E. histolytica measured by qPCR of fecal specimens, and (3) STH infections with Ascaris lumbricoides, Trichuris trichiura, and hookworm measured by Kato-Katz analysis of fecal specimens. These measurements showed reduced diarrhea in all arms except for the single water arm and reduced Giardia infection in all arms except for the single water and nutrition arms; Cryptosporidium and E. histolytica were too rare to assess impact. Hookworm was reduced in the
single water and sanitation arms as well as the combined WSH arm.19 The discrepancies between intervention impacts on *E. coli* vs health outcomes point to the complexity of environmentally mediated pathogen transmission, likely resulting from the divergence of fecal-oral pathogens, temporal fluctuations in transmission dynamics,44,45 differences in environmental survival,46–48 varying resistance to disinfection49,50 and imperfect correlations between fecal indicator bacteria and pathogens.51

Our *E. coli* measurements demonstrate a clear reduction in fecal exposure through stored drinking water in water intervention arms; this finding is consistent with our finding of reduced hookworm infection in the single water arm.19 The lack of diarrhea reduction in this arm may indicate that water was not a dominant transmission pathway for diarrheagenic pathogens in this population during this study. A previous study in a similar rural Bangladeshi population found 36% diarrhea reduction from the same water intervention,52 suggesting larger effect at a time of potentially more pronounced waterborne transmission. Another explanation is that measured reductions in chlorine-sensitive indicator bacteria do not indicate reductions in chlorine-resistant pathogens. Some viruses can withstand chlorine,49 and protozoan cysts are highly chlorine-resistant.50 If dominant diarrheagenic pathogens in the study population during the trial were chlorine-resistant, *E. coli* reductions in water would not translate to reductions in diarrhea. This is consistent with our finding that the single water arm had no reduction in infections with chlorine-resistant *Giardia*.18 *Cryptosporidium*, another chlorine-resistant protozoan pathogen,50 is a major cause of diarrhea in Bangladesh.53,54 *Cryptosporidium* infections (measured at the year two time point) were rare in our study;16 settings with high *Cryptosporidium* incidence would be another example where chlorination may reduce indicator bacteria in drinking water but not affect diarrhea. Our *E. coli* measurements also demonstrate a reduction in fecal exposure through food in the handwashing arm and, coupled with our observation of less dirt on caregiver hands in this arm, suggest reduced caregiver hand contamination. This is consistent with the reductions in diarrhea and *Giardia* infection in the handwashing arm.17,18 In contrast, STH infections were not reduced in this arm.19 This could be because the handwashing intervention did not remove dirt from fingernails; STH ova and larvae are detected under fingernails52 and nail clipping reduces parasite infection.55 Finally, the *E. coli* reductions in stored water and food in the combined WSH arm support the reductions in diarrheal, protozoan and STH infections in this arm, and the lack of synergistic impact on fecal contamination from combining WSH interventions is consistent with the lack of incremental health benefits in this arm.17–19

Our findings confirm that the use of *E. coli* as an indicator of water treatment effectiveness for chlorine-sensitive pathogens is appropriate but the alignment between *E. coli* measurements and infectious disease end points is heterogeneous and pathogen-specific. Our findings indicate that water, sanitation, and hygiene improvements reduced fecal contamination along the direct transmission pathways of stored drinking water and food but not in the overall ambient environment in this setting. These findings support implementing water treatment and handwashing to reduce fecal exposures through drinking water and food in low-income countries but provide no evidence that combining interventions further reduces exposure.

## ASSOCIATED CONTENT

### Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.est.8b05153.

Text S1. Environmental assessments nested within WASH Benefits Figure S1. Types of samples collected for environmental assessments approximately 4 months, one year and two years after the initiation of WASH Benefits interventions Table S1. *E. coli* detection units and limits Table S2. Enrollment characteristics by intervention group Table S3. *E. coli* prevalence and concentration measured in control, water treatment, handwashing and WSH arms after one year of intervention Table S4. *E. coli* prevalence and concentration measured in control, water treatment, handwashing and WSH arms after two years of intervention Table S5. Subgroup analysis by dry vs wet season on *E. coli* prevalence at year one Table S6. Subgroup analysis by dry vs wet season on *E. coli* prevalence at year two Table S7. Total/fecal coliform prevalence and concentration measured in control, water treatment, handwashing and WSH arms after one year of intervention Table S8. Total/fecal coliform prevalence and concentration measured in control, water treatment, handwashing and WSH arms after two years of intervention Table S9. *E. coli* prevalence and concentration at year one (after data from dates with contaminated blanks have been removed) Table S10. *E. coli* prevalence and concentration at year two (after data from dates with contaminated blanks have been removed) (PDF)

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### Notes

The authors declare no competing financial interest.

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