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**Introduction**

An elevated serum uric acid is commonly associated with the metabolic syndrome [1–3], hypertension [4], and cardiovascular disease [5, 6]. While the increase in serum uric acid was often considered secondary to these conditions [7–9], recent studies suggest it may have a contributory role [10]. First, serum uric acid commonly precedes and independently predicts hypertension [11, 12], insulin resistance/diabetes [13, 14], and obesity [15], which is inconsistent with it solely being a secondary factor. Second, studies in cell culture and animal models have suggested a causal role of uric acid in models of the metabolic syndrome and hypertension [16, 17]. Third, recent clinical trials, primarily pilot in nature, found potential benefits of lowering uric acid on blood pressure and insulin resistance [18–20]. These studies have renewed interest in uric acid as a potential mediator in the current epidemic of obesity and cardiovascular disease.

Few studies have examined whether polymorphisms affecting urate transport can predict features of the metabolic syndrome. Polymorphisms in the uric acid transporter SLC2A9 are associated with elevated serum uric acid and risk for gout [21, 22], but SLC2A9 polymorphisms...
are not associated with obesity and metabolic phenotypes [22]. However, whereas gout is a disease driven by extracellular concentrations of uric acid, the metabolic effects of uric acid are driven by its intracellular concentrations [23, 24], and SLC2A9 exports uric acid out of cells [25, 26].

In contrast to SLC2A9, the transporter URAT1 mediates entry of uric acid into cells [27]. Sautin et al. [24] reported that the URAT1 transporter is located on adipocytes where it drives inflammation and oxidative stress in response to uric acid. Adipocytes may also produce uric acid via intracellular xanthine oxidoreductase (XOR) [28]. A key role for XOR has been shown in adipogenesis, and XOR knockout mice fail to get fat [28].

These studies raise the possibility that urate transport into adipocytes may have a role in obesity and the metabolic syndrome. As such, we hypothesized that polymorphisms in the URAT1 transporter gene SLC22A12 may be associated with obesity and the metabolic syndrome.

Methods

Study Population

The Pharmacogenomic Evaluation of Antihypertensive Responses (PEAR) study (NCT002446519) is an ongoing prospective randomized parallel group study undertaken primarily to investigate genetic determinants to the antihypertensive and adverse metabolic effects of a thiazide diuretic and atenolol in mild-to-moderate primary hypertensive patients aged between 17 and 65 years without a history of diabetes mellitus or heart disease [29]. Subjects with elevated creatinine (males with creatinine >1.5 and females with creatinine >1.4), secondary hypertension, history of gout, alcohol/drug abuse or elevated liver function tests (elevated AST or ALT) were excluded. Randomized patients had all antihypertensives discontinued for an average of 1 month after which baseline studies were conducted. Patients were then randomized to treatment with either atenolol or hydrochlorothiazide. The initial 414 randomized patients were selected for this study after approval from the PEAR steering committee. Institutional review boards at the sites where PEAR was conducted, including the University of Florida, Gainesville, Fla., the Mayo Clinic, Rochester, Minn., and Emory University School of Medicine, Atlanta, Ga., all approved the PEAR study protocol, and all study participants provided informed written consent.

Phenotypes Studied

PEAR involved the collection of baseline data in all patients in the untreated state. Those who were treated with antihypertensives on entry to the study had them discontinued, with a washout period that was a minimum of 18 days, and typically 4–6 weeks in duration. Baseline data were collected at the end of this washout period and prior to initiation of the study drug. All data for this analysis arise from the baseline untreated study period in the PEAR study before addition of atenolol or hydrochlorothiazide. Waist circumference was measured to the nearest 0.5 cm. Weight and height were measured to the nearest 0.1 kg and 0.5 cm, respectively. Baseline laboratory studies including blood urea, serum creatinine, serum uric acid, and fasting lipids were measured on a Hitachi 911 Chemistry analyzer at the Mayo Clinic.

Genotyping

We selected promoter SNPs rs11602903 and rs505802 primarily to evaluate a likely functional SNP of the SLC22A12 gene that may be in linkage disequilibrium (LD) with an SNP reported in association studies with serum uric acid and gout (rs11231825) [30, 31]. The SNP rs11602903 in the promoter region was previously reported to be associated with reduced fractional excretion of uric acid (FEUA), which is defined as \( \frac{U_{UA} \times S_{Cr}}{S_{UA} \times U_{Cr}} \times 100 \), where \( U_{UA} \) and \( U_{Cr} \) are the urinary concentrations of uric acid and creatinine and \( S_{UA} \) and \( S_{Cr} \) reflect the respective serum concentrations [30]. SNP rs505802 is in complete LD \((\phi^2 = 1)\) in Caucasians with SNP rs11231825. Based on the lowest minor allele frequency (MAF) of 0.15, we estimated that about 250 study subjects would provide a power of 80% to detect the baseline difference in uric acid of 0.4 SD (0.42–0.5 mg/dl of uric acid) with an \( \alpha \) value of 0.05. We presumed that a significant difference in uric acid by genotype may be enough to detect a genotype association with obesity and the metabolic syndrome. Genomic DNA isolated from peripheral lymphocytes was used for genotyping. Genotype determination was carried out by the ABI TaqMan Prism 7900HT sequence detection system (Applied Biosystems) for rs505802 and rs11231825. The SNP rs11602903 was genotyped by pyrosequencing. The following PCR and sequencing primers were used for pyrosequencing: forward primer 5’-ATTGGGCACACCGAACCGAC-CTG-3’, reverse primer 5’-TGCTGTTGGCCTCCT-3’, and sequencing primer 5’-GAAGCTCCAGCGAGG-3’. For quality control, 10–15% of samples were genotyped twice. All genotypes were in concordance on repeat analysis.

Statistical Analysis

The Hardy-Weinberg equilibrium (HWE) was tested by using the \( \chi^2 \) test. To reduce kurtosis of the distribution, we square root transformed serum uric acid. Due to usual statistical genetic procedures, and particularly because of the differences in uric acid and MAF by race, all analyses were conducted separately in Caucasians and African Americans. The association of baseline serum uric acid with each of the three SNPs (rs505802, rs11602903, and rs11231825) was done by using analysis of covariance (ANCOVA) tested separately by race. An additive genetic model for the SNPs was used by assigning codes of 0, 1, and 2 for the wild-type homozygotes, heterozygotes, and variant homozygotes, respectively, for each SNP. We used age, gender, and body mass index (BMI) as covariates in association analysis with uric acid. ANCOVA was used for the association analysis of BMI, waist circumference, HDL cholesterol, and serum triglycerides after adjustment for age and gender. The criteria used to diagnose the metabolic syndrome were from the revised Adult Treatment Panel of the National Cholesterol Education Program [32]. We used logistic regression to evaluate the association of SLC22A12 SNPs with the metabolic syndrome after adjusting for age and gender in the model. All analyses were performed using SAS 9.1. Although three SNPs were tested, it turned out they were all in nearly perfect LD, thus the analyses were really essentially the same in Caucasians. Thus no correction was made for testing multiple SNPs. However, because we tested 5 different phenotypes, we defined \( p < 0.01 \) as statistically significant.
Results

Out of 414 subjects from PEAR, 234 Caucasians and 166 African Americans were included in the analysis. We excluded 14 other subjects who were of different ethnicities. Table 1 shows the MAF of SLC22A12 SNPs rs11602903, rs505802, and rs11231825 in Caucasians and African Americans. Each of the minor alleles of the three SLC22A12 SNPs in Caucasians were observed to be the major alleles in African Americans. All analyzed SNPs in each race were in HWE (p ≥ 0.05). LD between the three SNPs is shown in figure 1. This reveals that in Caucasians, the three SNPs were in nearly perfect LD, with lower levels of LD among African Americans. Because of the nearly complete LD in Caucasians, we report only the data for promoter SNP rs11602903 since the data for the other two are nearly identical.

Mean serum uric acid (±SD) was 5.66 ± 1.35 mg/dl in Caucasians and 5.38 ± 1.45 mg/dl in African Americans. In Caucasians, rs11602903 was not significantly associated with serum uric acid level (ANCOVA p = 0.15; table 2). The difference in serum uric acid between A/A versus T/T showed a trend toward significance with lower uric acid with increasing copies of the T allele (p = 0.0625 after adjustment for age and gender). For African Americans, the relationship of serum uric acid with SNP rs11602903 genotypes showed no differences between groups (A/A, 5.81 ± 0.12; A/T, 5.60 ± 0.22, and T/T, 5.37 ± 0.47 mg/dl, p = 0.85).

In Caucasians, rs11602903 was highly significantly associated with BMI (p < 0.0001, table 2) and waist circumference (p = 0.003, table 2). SNP rs11602903 explained 7% of the variation in BMI in Caucasians. In addition, SNP rs11602903 was significantly associated with HDL cholesterol (p = 0.018, table 2) but not with serum triglycerides although there was a trend towards significance (A/A vs. T/T, p = 0.09, table 2). As a replication cohort, we selected the Hypertension Genes Study involving 189 Caucasians with essential hypertension. There was no significant association between SLC22A12 SNP rs11602903 and BMI in this cohort. In African Americans, SLC22A12 SNP rs11602903 was not associated with BMI, waist circumference, HDL cholesterol or triglycerides, nor were the other two SNPs analyzed in this study.

SNP rs11602903 was significantly associated with the metabolic syndrome in Caucasians by logistic regression analysis (table 3). Patients with the AA genotype had 2.5 times odds of having the metabolic syndrome compared to those with the TT genotypes (odds ratio, OR: 2.53, 95% confidence interval, CI: 1.03–6.22, p = 0.033). In contrast, no association was observed in African Americans, with an OR of 1.27 (0.23–7.07), p = 0.81, for AA, and an OR of 1.28 (0.21–7.72), p = 0.82, for AT, compared to TT individuals.

Discussion

Our primary finding is that the URAT1 gene SLC22A12 SNP rs11602903 in the promoter region was highly associated with BMI (p < 0.0001) and explained 7% of the variation in BMI in hypertensive Caucasians from the PEAR study. rs11602903 was also associated with increased waist circumference, lower HDL and the metabolic syndrome in Caucasians. The observation of the association of SNPs in URAT1 with obesity, increased waist circumference, low HDL cholesterol, and

Table 1. MAF and HWE for SLC22A12 polymorphisms in Caucasians and African Americans

<table>
<thead>
<tr>
<th>SLC22A12</th>
<th>Caucasians</th>
<th></th>
<th>African Americans</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>minor allele</td>
<td>MAF</td>
<td>HWE p value</td>
<td>minor allele</td>
<td>MAF</td>
</tr>
<tr>
<td>rs505802</td>
<td>G</td>
<td>0.318</td>
<td>0.197</td>
<td>A</td>
</tr>
<tr>
<td>rs11602903</td>
<td>A</td>
<td>0.322</td>
<td>0.276</td>
<td>T</td>
</tr>
<tr>
<td>rs11231825</td>
<td>T</td>
<td>0.325</td>
<td>0.129</td>
<td>C</td>
</tr>
</tbody>
</table>

Fig. 1. LD in Caucasians (a) and African Americans (b) for the three SLC22A12 SNPs rs11602903, rs505802, and rs11231825 are shown as $r^2$. 

Table 1. MAF and HWE for SLC22A12 polymorphisms in Caucasians and African Americans
the metabolic syndrome is consistent with epidemiological [13, 33] and experimental studies [17, 34] suggesting a contributory role of uric acid to the pathogenesis of the metabolic syndrome. An elevated uric acid was shown to be an independent risk factor for obesity in a study in Japan [15], and a meta-analysis also found uric acid to independently predict insulin resistance and diabetes [13].

Interestingly, the SLC22A12 SNP rs11602903 correlated with serum uric acid levels in Caucasians in this study, although it did not reach significance. In a case-control study involving Caucasians, Graessler et al. [30] reported that the A/A genotype of SLC22A12 SNP rs11602903 was associated with reduced renal uric acid excretion (OR: 1.8, 95% CI: 1.129–2.993, p = 0.0139 for FEUA ≤ 6.5%). A decreased renal uric acid excretion implies increased URAT1 activity which should correlate with higher intracellular uric acid levels.

In contrast to the findings in Caucasians, no relationship of rs11602903 with obesity and the metabolic syndrome was observed in African Americans. Since rs11602903 is not considered a functional SNP, this could be explained by LD differences between both groups. Other explanations are possible. For example, African Americans have a diet that is much higher in sugar [35], and the fructose component in sugar is known to raise intracellular uric acid [36], which could theoretically reduce the impact of a polymorphism involved in urate transport into the cell.

Limitations in this study include our inability to reproduce these findings in a different cohort (The Hypertension Genes Study) involving 189 Caucasians with a similar hypertensive phenotype. This could have been due to the small sample size, sampling error, dietary differences, or differences in genetic factors affecting obesity in that cohort. The weight and height in the hypertension cohort were self-reported so there may be errors in the reported weight by individuals. Despite the limitations, this study raises important questions as to whether transporters regulating intracellular uric acid are involved in the human obesity epidemic.

In conclusion, our data suggest that urate transport SLC22A12 gene polymorphisms may influence obesity.

<table>
<thead>
<tr>
<th>Variable</th>
<th>OR</th>
<th>95% CI</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age</td>
<td>0.96</td>
<td>0.93–0.99</td>
<td>0.01</td>
</tr>
<tr>
<td>Female gender</td>
<td>1.57</td>
<td>0.92–2.67</td>
<td>0.097</td>
</tr>
<tr>
<td>rs11602903 AA genotype</td>
<td>2.53</td>
<td>1.03–6.22</td>
<td>0.033</td>
</tr>
<tr>
<td>rs11602903 AT genotype</td>
<td>0.99</td>
<td>0.56–1.74</td>
<td>0.12</td>
</tr>
</tbody>
</table>

Table 3. Association of SLC22A12 SNP rs11602903 with BMI, waist circumference and HDL cholesterol

<table>
<thead>
<tr>
<th>Variable</th>
<th>A/A</th>
<th>A/T</th>
<th>T/T</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Caucasians (n = 234)</td>
<td>n = 28</td>
<td>n = 95</td>
<td>n = 111</td>
<td></td>
</tr>
<tr>
<td>Age, years</td>
<td>52.60 ± 1.67</td>
<td>52.04 ± 0.98</td>
<td>50.68 ± 0.82</td>
<td>0.44</td>
</tr>
<tr>
<td>Women, %</td>
<td>53.57</td>
<td>52.17</td>
<td>41.44</td>
<td>0.26</td>
</tr>
<tr>
<td>Serum uric acid, mg/dl</td>
<td>5.91 ± 0.21</td>
<td>5.64 ± 0.11</td>
<td>5.46 ± 0.10</td>
<td>0.15</td>
</tr>
<tr>
<td>Triglycerides, mg/dl</td>
<td>168.54 ± 18.16</td>
<td>152.04 ± 10.02</td>
<td>134.12 ± 9.19</td>
<td>0.17</td>
</tr>
<tr>
<td>HDL cholesterol, mg/dl</td>
<td>41.97 ± 2.44</td>
<td>48.83 ± 1.34</td>
<td>49.78 ± 1.23</td>
<td>0.018</td>
</tr>
<tr>
<td>BMI</td>
<td>34.93 ± 1.01</td>
<td>30.27 ± 0.55</td>
<td>30.51 ± 0.51</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Waist circumference, cm</td>
<td>105.63 ± 2.28</td>
<td>97.22 ± 1.24</td>
<td>98.39 ± 1.14</td>
<td>0.0033</td>
</tr>
</tbody>
</table>

African Americans (n = 166)

<table>
<thead>
<tr>
<th>Variable</th>
<th>A/A</th>
<th>A/T</th>
<th>T/T</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, years</td>
<td>48.55 ± 7.89</td>
<td>48.0 ± 8.65</td>
<td>49.0 ± 11.46</td>
<td>0.92</td>
</tr>
<tr>
<td>Women, %</td>
<td>66.94</td>
<td>84.21</td>
<td>85.71</td>
<td>0.084</td>
</tr>
<tr>
<td>Serum uric acid, mg/dl</td>
<td>5.8 ± 0.12</td>
<td>5.6 ± 0.22</td>
<td>5.37 ± 0.47</td>
<td>0.85</td>
</tr>
<tr>
<td>Triglycerides, mg/dl</td>
<td>108.75 ± 8.16</td>
<td>107.83 ± 14.75</td>
<td>100.7 ± 32.2</td>
<td>0.97</td>
</tr>
<tr>
<td>HDL cholesterol, mg/dl</td>
<td>49.44 ± 1.36</td>
<td>50.86 ± 2.46</td>
<td>51.25 ± 5.43</td>
<td>0.84</td>
</tr>
<tr>
<td>BMI</td>
<td>31.23 ± 0.55</td>
<td>31.41 ± 0.98</td>
<td>30.82 ± 0.51</td>
<td>0.97</td>
</tr>
<tr>
<td>Waist circumference, cm</td>
<td>96.10 ± 1.29</td>
<td>99.10 ± 2.32</td>
<td>99.07 ± 5.35</td>
<td>0.4888</td>
</tr>
</tbody>
</table>

Data are expressed as means ± SD.
and the metabolic syndrome in essential hypertension. We hope these studies will stimulate additional research studies investigating the role of genes involved in uric acid metabolism and transport in the pathogenesis of obesity and the metabolic syndrome.

Acknowledgments

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Disclosure Statement

Dr. Richard Johnson has patent applications related to lowering uric acid as a means for reducing the metabolic syndrome via the University of Florida. All other authors have no conflicts of interest.

References


