Glutamatergic Tuning of Hyperactive Striatal Projection Neurons Controls the Motor Response to Dopamine Replacement in Parkinsonian Primates

Highlights

- Selectively reducing AMPAR or NMDAR transmission stabilizes SPN responses to DA
- Reduced glutamate signaling stabilizes DA-induced activity increases and decreases
- Reduced glutamate signaling across SPNs suppresses dyskinesias in primates
- Data support the mechanistic role of SPN hyperactivity in responses to DA in PD

Authors

Arun Singh, Meagan A. Jenkins, Kenneth J. Burke, Jr., ..., Annalisa Scimemi, Stephen F. Traynelis, Stella M. Papa

Correspondence
spapa@emory.edu

In Brief

Singh et al. analyze the role of SPN hyperactivity in abnormal responses to dopamine replacement in parkinsonian primates with selective local blockade of glutamate signaling. Lowering basal firing stabilizes the SPN response to dopamine and normalizes motor responses.

Singh et al., 2018, Cell Reports 22, 941–952
January 23, 2018 © 2017 The Author(s).
https://doi.org/10.1016/j.celrep.2017.12.095
Glutamatergic Tuning of Hyperactive Striatal Projection Neurons Controls the Motor Response to Dopamine Replacement in Parkinsonian Primates

Arun Singh, Meagan A. Jenkins, Kenneth J. Burke, Jr., Goichi Beck, Andrew Jenkins, Annalisa Scimemi, Stephen F. Traynelis, and Stella M. Papa

1Department of Neurology, Emory University School of Medicine, Atlanta, GA 30329, USA
2Department of Pharmacology, Emory University School of Medicine, Atlanta, GA 30322, USA
3Department of Anesthesiology, Emory University School of Medicine, Atlanta, GA 30322, USA
4Department of Biology, State University of New York, Albany, NY 12222, USA
5Department of Neurology, Emory University School of Medicine, Atlanta, GA 30329, USA
6Lead Contact
Correspondence: spapa@emory.edu
https://doi.org/10.1016/j.celrep.2017.12.095

SUMMARY

Dopamine (DA) loss in Parkinson’s disease (PD) alters the function of striatal projection neurons (SPNs) and causes motor deficits, but DA replacement can induce further abnormalities. A key pathological change in animal models and patients is SPN hyperactivity; however, the role of glutamate in altered DA responses remains elusive. We tested the effect of locally applied AMPAR or NMDAR antagonists on glutamatergic signaling in SPNs of parkinsonian primates. Following a reduction in basal hyperactivity by antagonists at either receptor, DA inputs induced SPN firing changes that were stable during the entire motor response, in clear contrast with the typically unstable effects. The SPN activity reduction over an extended putamenal area controlled the release of involuntary movements in the “on” state and therefore improved motor responses to DA replacement. These results demonstrate the pathophysiological role of upregulated SPN activity and support strategies to reduce striatal glutamate signaling for PD therapy.

INTRODUCTION

Motor failure in Parkinson’s disease (PD) is caused primarily by progressive neurodegeneration of the substantia nigra pars compacta. The loss of nigral dopamine (DA) cells has usually reached a considerable level by the time motor deficits develop (Lang and Lozano, 1998). The central role of DA is also demonstrated by the effectiveness of DA replacement to improve motor symptoms in all stages of the disease. However, our understanding of the pathophysiology of motor control in PD is far from clear, particularly with respect to the response to DA replacement. Adding DA to the system does not restore normal movement but rather induces a partial and short recovery that is further complicated by involuntary movements called dyskinesias (Obeso et al., 2000). Indeed, in experiments that are controlled for pharmacological variables, the effective DA stimulation is not yet followed by the expected restitution of normal function (Bravi et al., 1994; Nutt et al., 2000).

DA modulates the excitability of striatal projection neurons (SPNs), which express DA D1 receptors (D1R) or DA D2 receptors (D2R), forming the direct and indirect striatal output pathways, respectively (Gerfen and Surmeier, 2011). Direct SPNs (dSPNs) and indirect SPNs (iSPNs) undergo multiple functional and morphological changes following nigrostriatal denervation that may be involved in altered responses to dopaminergic stimulation (Surmeier et al., 2014). One of the salient changes is the increased spontaneous SPN activity that has been found across animal models and patients. From activity levels usually below 2 Hz in the normal condition, the average firing frequency increases variably in rodent models to 5–12 Hz under anesthesia (Taeng et al., 2001) and to more than 20 Hz in alert, advanced parkinsonian primates and patients with PD (Liang et al., 2008; Singh et al., 2016). These large SPN activity increases in primates and humans were not yet identified in cells segregated into specific output pathways. In line with classic views of the functional model of PD, the use of optogenetics in transgenic mouse models has suggested that iSPNs are most likely the upregulated units after DA denervation (Kravitz et al., 2010). However, further studies disputed the classic views of the model, demonstrating the cooperative activity of both striatal pathways for basal ganglia outputs and movement initiation (Cui et al., 2013; Freeze et al., 2013). In addition, the primate studies show few low-activity units and opposite responses to DA among the recorded SPNs. These observations are difficult to reconcile with the idea of recordings limited to one SPN subgroup in the primate and thereby call into question previous assumptions on the distribution of hyperactive SPNs (Beck et al., 2017). Yet crude single-cell recordings in primates and patients critically show that there are large firing increases in the active SPNs in the absence of DA. Such a state of high basal activity likely may interfere with the strength of DA signaling to modulate SPN excitability. Congruent with this premise, dopaminergic stimulation induces unstable changes in SPN firing frequency that are associated with dyskinesias in primates with advanced Parkinsonism (Liang...
et al., 2008; Singh et al., 2015). Thus, SPN hyperactivity may play a primary role in the altered responses to DA replacement.

Glutamate inputs from cortical and thalamic terminals provide the excitatory drive of the SPN and likely contribute to the hyperactivity developed in PD. The cumulative evidence supports up-regulation of corticostriatal signals (Guabellini et al., 2002; Ingham et al., 1998), but recent data also show changes in the strength of thalamostriatal synapses after DA loss (Parker et al., 2016). Glutamatergic synaptic contacts undergo significant reorganization due to morphological changes of the SPN dendritic arborization (Day et al., 2006; Villalba and Smith, 2017). Notably, spine loss and dendrite changes are differentially developed in dSPNs and iSPNs, indicating that various adaptations may remodel glutamatergic synapses. In the same line, ex vivo recordings following DA depletion demonstrate increased excitability and loss of long-term potentiation and depression at corticostriatal synapses (Bagetta et al., 2011; Shen et al., 2008). Although some of these changes can be reversed by DA replacement, corticostriatal synaptic connectivity, strength, and plasticity remain altered (Fieblinger et al., 2014; Picconi et al., 2003). At the level of glutamate receptors, both DA loss and replacement are associated with changes in the expression, subunit composition, and ratio of z-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptor (AMPAR) to N-methyl-D-aspartate receptor (NMDAR) (Bagetta et al., 2012; Mellone et al., 2015). Altogether these data suggest that an intervention to regulate the glutamate signals tuning SPN activity may enable more effective DA modulation.

Here, we used the primate model of advanced PD to study the SPN firing changes related to the motor response induced by dopaminergic stimulation under the control of effective glutamate inputs on the recorded SPN by applying selective NMDAR or AMPAR antagonists. Prior to these tests in the primate, we determined (1) the selectivity of the drug at the concentration applied locally using patch-clamp recordings in cultured cells, (2) the expected concentration of the drug reached in vivo using in silico prediction of diffusion following intraparenchymal injection, and (3) the corresponding doses of the NMDAR and AMPAR antagonists for an equivalent effect reducing high SPN activity in vivo. We also tested the motor effects of glutamate-controlled SPN activity over the primate putamen. Our data show that a decrease of either AMPAR or NMDAR signaling that substantially reduced the basal SPN firing rate results in effective stabilization of the neuronal response to DA during the whole period of reversal of parkinsonian symptoms, referred to as the “on” state. This effect at the single-cell level translated into motor behavioral effects following the NMDAR antagonist infusion over an extended striatal area. The drug-induced reduction of activity across SPNs markedly decreased the dyskinesias caused by L-DOPA. These data support a pivotal role of striatal glutamatergic excitation and SPN hyperactivity in the response to DA replacement in PD.

RESULTS

Selectivity of NMDAR and AMPAR Antagonists for In Vivo Microinjection Assays
The impact of reduced glutamatergic signaling on the dysregulated SPN activity in parkinsonian monkeys was determined with single-cell recordings after intrastriatal injection of high concentrations of selective NMDAR and AMPAR antagonists. Because the rapid dilution and diffusion of antagonists in vivo can be determined quantitatively, we experimentally confirmed the antagonist selectivity in vitro by establishing the concentration-effect relationship for inhibition by each antagonist on currents mediated by recombinant isofoms of NMDAR and AMPAR found in the SPN. We evoked these currents using rapid glutamate applications in order to mimic the time course of the glutamate concentration profile in the synaptic cleft during synaptic transmission (Clements et al., 1992). Pre-application of the NMDAR-selective antagonist LY235959 for 1 s antagonized AMPAR currents only at concentrations much higher than those predicted to be reached with intraparenchymal injection following diffusion and dilution (see below). Specifically, the NMDAR-selective antagonist LY235959 inhibited recombinant AMPAR (GluA1/stargazin) currents by 80% ± 2% at 9 mM and 50% ± 6% at 900 μM but only 13% ± 7% at 90 μM; the half maximal inhibitory concentration (IC50) of LY235959 was 1.1 mM. By contrast, the IC50 of NMDAR antagonist LY235959 at GluN1/GluN2A NMDARs activated by 1 mM glutamate was 12 μM (Figure 1A), indicating that LY235959 is at least 100-fold selective for NMDARs over AMPARs activated by concentrations of glutamate reached in the synaptic cleft. We also evaluated the selectivity of the AMPAR competitive antagonist NBOX by determining non-selective inhibition of recombinant GluN1/GluN2A NMDAR currents. NBOX inhibited NMDAR current responses to 1 mM glutamate by 50% ± 0.3% at 1 mM and 36% ± 0.7% at 300 μM but only 14% ± 0.6% at 100 μM; the fitted IC50 for NBOX non-selective inhibition of GluN1/GluN2A NMDARs was 0.9 mM. By contrast, the IC50 for NBOX inhibition of GluA1/stargazin AMPARs activated by rapid application of 1 mM glutamate was 2.7 μM (Figure 1B). Thus, NBOX is 300-fold selective for AMPAR over NMDAR activated by synaptic-like rapid exposure to glutamate. We expect both competitive antagonists to be even more potent at blocking their target receptors when these are activated by brief glutamate pulses in the synaptic cleft, which are less likely to displace any pre-bound antagonist molecules.

The striatum of a rhesus macaque is approximately 1,000 mm3 (Yin et al., 2009), which is equivalent to 1 mL in volume. Thus, injection of 1 μL of antagonist into this brain area, at steady state, would lead to an approximate 1,000-fold dilution. The time dependence of diffusion is governed by Fick’s laws and depends on the complex geometry of cell processes forming barriers and hindering diffusion in the brain tissue. We estimated the peak concentration of AMPAR and NMDAR antagonists at different distances and at different time intervals after being injected into brain parenchyma (Figure 1C). Briefly, we described their space-time concentration profile from an instantaneous point source using the classic solution of diffusion from a point source in an isotropic medium, with parameters that describe the diffusion properties of small molecules in the brain neuropil (Savtchenko and Rusakov, 2005; Zheng et al., 2008). The diffusion analysis showed that a more than 1,000-fold dilution of LY235959 or NBOX occurs within less than 1 min from the injection (Figures 1D and 1E). These data suggest that injection of millimolar concentrations of antagonists will rapidly produce sub-micromolar
concentrations in the brain tissue surrounding the injection site. These simulations, together with voltage-clamp recordings of the non-equilibrium receptor response to the rapid application of glutamate at concentrations that approach those in the synapse, suggest that our in vitro estimation of diffusion and potency can be used to predict in vivo properties and that the antagonists reached concentrations in brain parenchyma that were selective following microinjection.

Dose Selection of NMDAR and AMPAR Antagonists

On the basis of the results of antagonists’ specificity in vitro and on available estimates of drug dilution in brain tissue, we performed repeated striatal microinjections of 1–9 mM LY235959 and 0.5–3 mM NBQX in the parkinsonian primate to select doses that are equally effective in reducing cell activity for application in subsequent experiments of L-DOPA administration. A dose-dependent decrease in spontaneous SPN firing frequency was observed with each antagonist
Figure 2. NMDAR Antagonist Injection at the Site of SPN Recording Reduced the Basal Activity and Stabilized DA Responses in the Parkinsonian Primate

(A–D) Control tests. Firing frequency changes of each SPN at baseline (pre), after local injection of aCSF (post), and after transitioning to “on” (on) and dyskinesias states (dys) following s.c. injection of L-DOPA. In each panel, each colored curve (top graph) represents an individual SPN grouped according to the type of DA response (increase, A and B, or decrease, C and D, in the “on” state followed by stable, A and C, or unstable, B and D, response in the dyskinesia state), and the averaged firing frequency change for the group is shown as percentage (bottom graph). In each SPN, differences between post and on are significant at p < 0.01 (A–D). Responses were classified as unstable by significant changes in the dyskinesia state (p < 0.01). Total SPNs, 29; units with activity increase, 14; units with activity decrease, 15.

(legend continued on next page)
In the "on" state, 24% units with activity decrease, 15. In each group analysis, differences between pre and post and between post and on are significant at p < 0.01. Differences between on and dys were non-significant (ANOVAs for repeated-measure followed by Bonferroni correction). In (F) and (H), *p < 0.05 between LY235959 and aCSF unstable (E–H) NMDAR antagonist tests. The firing frequency changes of each SPN as described above for control tests are shown in (E) and (G).

### Effect of Reduced NMDAR and AMPAR Signaling on the SPN Response to DA in Parkinsonian Primates

The SPN activity in parkinsonian primates (n = 5; Table S1) was recorded continuously throughout the local application of artificial cerebrospinal fluid (aCSF) (control tests), LY235959, or NBQX, and s.c. L-DOPA injection that induced motor changes (Figures 1F and 1G). Recordings started in the "off" state from before to after local injection and continued through the L-DOPA-induced "on" and dyskinesia states. Thus, neuronal activity data were stored for offline analysis at the following time points: (1) before local injection ("pre"), (2) after local injection ("post"), (3) "on" state (onset of reversal of parkinsonism induced by L-DOPA; "on"), and (4) dyskinesia (L-DOPA-induced dyskinesia at the peak-dose effect; "dys"). Following local aCSF application (control test) that had no effect or produced very brief changes of SPN activity (pre to post), L-DOPA induced the typical responses of increased or decreased firing frequency (post to on; Figures 2A–2D) (Singh et al., 2015). In these control tests, SPN activity increased in 14 units and decreased in the other 15 units, and these firing changes correlated with the behavioral changes indicating that the animal had transitioned to the "on" state. Also, aCSF application had no effect on the magnitude of responses. The firing frequency in the "on" state increased by 106% and decreased by 36% (average changes in SPNs grouped by their response to DA), comparable with previously reported data. At the peak of L-DOPA effect, the firing rate changes were maintained or continued to develop (stable responses) in 41% of SPNs (on to dys; Figures 2A and 2C). In contrast, at the peak effect responses reversed in 59% of SPNs producing the inverted firing rate changes during the "on" state that correlate with the appearance of peak-dose dyskinesias in advanced parkinsonian primates (Figures 2B, 2D, 2I, and 2J) (Liang et al., 2008).

Following reduction of the baseline ("off" state) SPN activity by LY235959 application, L-DOPA administration produced changes in SPN firing associated with the "on" state that were similar to those found in control experiments (i.e., activity increase or decrease from post to on). However, the firing frequency changes during the "on" state were stable in the large majority of SPNs (Figures 2E–2H, 2K, and 2L). As dyskinesias remain the same with the limited effect of local microinjection of LY235959, the stabilized SPN activity in response to DA was caused by the NMDAR antagonist. Similar effects were obtained with NBQX application (Figure 3). Thus, both NMDAR and AMPAR antagonists at selective doses that induced nearly 50% reduction of SPN activity equally prevented the inversion of frequency changes at the peak of the L-DOPA response (concomitant with dyskinesias), contrasting with the control experiments with aCSF alone that resulted in a large number of inverted (unstable) responses. The stability of changes during the "on" state in experiments of local application of NMDAR or AMPAR antagonist was found regardless of the type of response to L-DOPA (i.e., increase or decrease of the firing rate during the "on" state). Following LY235959 application, 39 of 43 SPNs had stable responses to L-DOPA, and following NBQX application, 43 of 45 SPNs also had stable responses, reaching a total of 93% of the recorded SPNs with stable responses (Figures 4 and S3). Therefore, the blockade of NMDAR or AMPAR transmission that effectively reduced the SPN hyperactivity of the parkinsonian state restored full, stable responsiveness to DA signaling.

To further determine the relationship between the DA response and the activity reduction induced by the NMDAR or AMPAR antagonist, we analyzed the correlation of firing changes in each SPN. The lowered firing frequency after LY235959 or NBQX application at the recording site (post/pre) was a predictor of the amount of increased activity in response to L-DOPA at the initial behavioral change (on/post), or at the peak-dose effect (dys/post), accounting for 35%–50% of the change (see $R^2$ values in Figures 5A and 5B; p < 0.01). In this subset of SPNs (units with DA-induced activity increase), the response to DA was thus highly dependent on the level of reduced basal activity. In contrast, in the SPN subset with DA-induced activity decrease, the amount of activity reduction in the "on" or dyskinesia state was not predicted by the level of reduced basal activity (Figures 5C and 5D; p > 0.05). In this subset of SPNs, DA-induced changes stabilized, but the strength of the response was independent of the reduced baseline activity. Therefore, data indicate different DA regulation across the distinguished SPN subsets.

### Effect of Reduced Striatal NMDAR Signaling on Motor Responses to L-DOPA in Parkinsonian Primates

Motor responses to L-DOPA in primates with advanced parkinsonism are consistently complicated by dyskinesias that are
Figure 3. AMPAR Antagonist Injection at the Site of SPN Recording Reduced the Basal Activity and Stabilized DA Responses in the Parkinsonian Primate

(A–D) The same control data as presented in Figures 2A–2D, respectively, for comparison with results obtained with the AMPAR antagonist. See Figure 2 for details.

(E–H) AMPAR antagonist tests. Firing frequency changes of each SPN as described for control tests are shown in (E) and (G) after local injection of NBQX. In each SPN, differences between Pre and Post, and between Post and On are significant at p < 0.01. Differences between On and Dys were non-significant (see also Figure S3). SPN stable responses after NBQX are compared to control tests in (F) and (H). Total SPNs, 43; units with activity increase in the “on” state, 26; units with activity decrease, 17.

(legend continued on next page)
associated with inversion of the SPN firing rate changes during the “on” state. To test whether the reduced glutamatergic transmission that stabilized the SPN response to L-DOPA had the expected behavioral correlate, LY235959 (9 mM) or aCSF was infused unilaterally into the putamen of primates (n = 3; Table S1) before the s.c. injection of L-DOPA. The infusion volume (10 μL) was intended to cover only the most posterolateral region corresponding to the sensorimotor territory (Sanftner et al., 2005). The preselected individual dose of L-DOPA induced reproducible monophasic peak-dose dyskinesias in the tested animals. Striatal infusion of the NMDAR antagonist reduced the global dyskinesia scores (Figures 6A and 6C) because of 71% lower scores in the hemibody contralateral to the infusion side after L-DOPA injection (p < 0.05; Figures 6B and 6D; Movies S1 and S2). LY235959 infusion did not affect the antiparkinsonian action of L-DOPA (unchanged motor disability scores [MDS], p > 0.05; Figure 6E). Therefore, LY235959 infusion into the putamen showed that the extended stabilization of DA responses across SPNs translates into motor changes with significantly reduced dyskinesias. This demonstrates that decreasing glutamatergic excitation in the striatum induces beneficial motor effects in the parkinsonian primate.

The motor effects of systemic administration of LY235959 and NBQX were also tested in a group of parkinsonian primates (n = 5; Table S1) that included three of the animals used in striatal application of the antagonists. Both LY235959 (3 mg/kg s.c.) and NBQX (2 mg/kg s.c.) significantly reduced dyskinesias (p < 0.05; Figures S4 and S5) without compromising the antiparkinsonian action of L-DOPA (Löschmann et al., 1991; Papa and Chase, 1996). The similarity of these effects (including animals used for SPN recordings) to those obtained with striatal infusion suggests that the reduction of glutamate signals on SPNs may be responsible for the effects induced by systemic administration.

**DISCUSSION**

The selective reduction of either NMDAR or AMPAR signals in SPNs tested here in parkinsonian primates supports our hypothesis that altered SPN responses to DA and their associated abnormal movements can be reversed by controlling the dysregulated glutamatergic drive. Because the animals used in these single-cell recordings had developed significant SPN hyperactivity, the impact of antagonist application on the spontaneous firing of these neurons was fully assessed. The most important finding from this study was that the reduced excitatory glutamate signals resulted in stable responses to DA in 93% of the recorded SPNs, and these effects were found across single recordings of SPNs regardless of their response to DA (i.e., units with activity increase or decrease). In addition, both the NMDAR and AMPAR antagonists equally stabilized DA responses in the large majority of neurons, and effects of the NMDAR antagonist on an extended putamenal area reduced dyskinesias improving the L-DOPA response.

The interpretation of the recorded SPN activity is limited by lack of specific cell-type identification with optogenetics, because transgenic modeling could not be applied to these primate studies. More important to this end, DA stimulation evokes pathological “unstable” responses that are widely distributed across neurons, with an increase or decrease of activity in the “on” state. Indeed, as L-DOPA reaches its peak effect, a high proportion of units do not maintain the DA-induced firing increase or decrease, which contrasts markedly with stable changes in other units and thereby creates a large imbalance of discharges. On the basis of evidence that the cooperative activity of both striatal pathways mediates normal movement execution (Cui et al., 2013), imbalance of discharges between and within the output pathways may lead to abnormal motor responses, releasing involuntary movements or causing motor fluctuations (Singh et al., 2015; Tecuapetla et al., 2014). In support of this view, the instability of DA-induced firing changes that cause discharge imbalance is associated with dyskinesias in the primate. Here, DA-induced responses were consistently stabilized across SPNs after glutamate inputs were blocked and the high baseline firing frequency was decreased, indicating that hyperactivity across SPNs is a key mechanism in the pathophysiology of PD “off” and “on” states. These data are
Of note, gene regulation after DA loss leads to changes in not fully develop in commonly used rodent and primate models. Thus, it is plausible that extensive SPN hyperactivity does advanced with long-standing chronicity (Potts et al., 2014). and “slowly” induced parkinsonism that was classified as lateral and acute lesions, the primates used here had a bilateral such as spine remodeling (Villalba and Smith, 2017). Unlike uni-

important are the lesion type and its time course, which may in-

changes in models of DA loss are the hyperphosphorylation of GluR1 and the expression of Ca²⁺-permeable GluR2-lacking AMPAR (Bagetta et al., 2012).
Changes in Ca^{2+} kinetics are consistent with the pathological basal activity of SPNs and potentially their "unstable" response to DA. The NMDAR- and AMPAR-selective competitive antagonists (LY235959 and NBQX) used here were not subunit selective, and thus each effectively blocked most receptor subtypes within the targeted families, reducing the "off"-state hyperactivity to a similar extent as intended for comparison. Therefore, the ability of both NMDAR and AMPAR blocks to induce the same stabilization of DA responses supports the contribution of presynaptic (increased input) and postsynaptic (hyperexcitability) mechanisms in the SPN dysfunction.

The consistent glutamate antagonist effect on single neurons suggested that extended effects across SPNs would prevent the imbalance of discharges, and possibly improve the behavioral response to DA. Our tests support this idea. The infusion of the same concentration of the NMDAR antagonist over an extended area of the putamen reduced dyskinesias in the parkinsonian primate. Because the infusion into the putamen was unilateral and centered on the posterolateral motor territory using a discrete volume to avoid overflow to surrounding extrastriatal areas, most likely the covered area did not extend to the whole target region, but nevertheless, dyskinesias were reduced by 71%. In line with our results, the only drug in clinical use to treat dyskinesias is amantadine, an agent with actions at multiple sites, including the NMDAR (Oertel et al., 2017). On the basis of the effects of selective agents, the antidyskinetic effect of amantadine is likely mediated by NMDAR block. Regarding further effects of NMDAR antagonists on L-DOPA responses, it is possible that chronically reduced SPN activity also results in reduction of motor fluctuations and improved "on" state. In addition to chronic block, the antagonist intrinsic effect on parkinsonism remains to be tested, because in the present tests the infusion timeline was designed to assess L-DOPA-induced dyskinesias. Importantly, the present results provide proof of concept for the significance of reduced glutamatergic tone and stabilization of DA-induced firing changes in SPNs. Because the current therapy for patients with PD remains symptomatic and based on dopaminergic stimulation for relief of motor disability, the present results have significant clinical implications. Treatment efficacy depends on consistently recovering mobility with responses free of dyskinesias, but most patients between mid- and late-stage disease suffer from disabling motor complications. Critically, the primates used in these experiments reproduced the phenotype of patients in
that category, and SPN recordings in these patients have shown the same level of SPN hyperactivity as in the parkinsonian primate. Likewise, glutamate-driven SPN hyperactivity plays a central role in the abnormal DA responses developed in patients with PD. Strategies targeting specifically the striatal glutamate overactivity may thus help improve the efficacy of DA replacement. Of particular interest are the mechanisms regulating protein composition and posttranslational changes in AMPARs/NMDARs, including molecular changes that increase AMPAR activity or channel conductance (Hosokawa et al., 2015; Jenkins et al., 2014; Kristensen et al., 2011) and thereby upregulate SPN activity.

**EXPERIMENTAL PROCEDURES**

**In Vitro NMDAR and AMPAR Blockade and Simulated Antagonist Diffusion**

The experiments in vitro and simulations of drug diffusion in brain tissue were designed to determine whether LY235959 and NBQX would yield concentrations that selectively blocked NMDAR or AMPAR, respectively, using the specific and time parameters of the microinjection used for recording experiments. Details are provided in Supplemental Experimental Procedures.

**Non-human Primate Model of PD**

Nine adult male and female macaques (Macaca mulatta and Macaca fascicularis; 5–11 kg body weight) were used in the studies (see Table S1). All procedures followed guidelines of the NIH Guide for the Care and Use of Laboratory Animals and were approved by the Institutional Animal Care and Use Committee of Emory University. A model of “chronic and advanced” parkinsonism was produced in all animals by systemic administration of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP), and behavioral assessment using a standardized motor disability scale for primates. In these parkinsonian primates, the daily maintenance L-DOPA treatment led to the development of L-DOPA-induced dyskinesias. In each animal, the effective subcutaneous (s.c.) dose of L-DOPA methyl ester plus benseradine (Sigma-Aldrich) for use in recording experiments was determined as the dose eliciting a clear “on” state with peak-dose dyskinesias. A complete description is provided in the Supplemental Information.

**Continuous Single-Cell Recording with Striatal NMDAR or AMPAR Blockade and L-DOPA-Induced Motor States**

Five animals were surgically implanted with recording chambers and head-restraining devices, and striatal regions were identified with electrophysiological mapping of basal ganglia. Standard techniques were used for single-cell recordings with withdrawal of the oral maintenance L-DOPA treatment the day of recording. NMDAR or AMPAR antagonist was delivered at the site of recording, using “injectrodes” connected to a microinjection pump. The antagonist solution (or the same volume of aCSF alone as control) was injected in a volume of 200 nL at a rate of 1 μL/min. LY235959 was dissolved in aCSF and NBQX in aCSF/water. The tip of the electrode was placed at a fixed distance from the tip of the cannula (400 μm; Figure 1C). The antagonist concentration (within the limits set from in vitro tests and simulations to maintain selective doses at the synapses) that effectively reduced activity of the recorded cell by ∼50% was assessed using different drug concentrations and analyzing a total of 69 SPNs. After dose selection, complete experiments started by local application of antagonist followed by s.c. L-DOPA injection at the predetermined dose with data storage (≥3 min) according to the timeline (Figure 1G). If the baseline activity was held throughout the total duration of the experiment, the offline analysis yielded one or occasionally two units per experiment. A total of 117 SPNs were analyzed in complete experiments of DA responses, 88 SPNs for antagonists, and 29 SPNs for vehicle control tests. The animal’s behavior was monitored using a video camera. Details of experiments, including online unit isolation, behavioral changes during recordings indicative of motor states and histological verification of recording sites are included in the Supplemental Information.

**Electrophysiology Data Analysis and Statistics**

All data were analyzed offline using spike sorting and standard methods for differentiation and classification of SPNs (Figure S6). Significant activity changes in the “on” state (p < 0.05, ANOVAs for repeated-measures followed by post hoc Bonferroni test) separated units with increase or decrease response. Also, significant frequency changes in dyskinesia state determined whether the increased or decreased firing rate in the “on” state was stable or not. Activity changes in the “on” and dyskinesia states were analyzed for their relationship to the reduced firing frequency after the local application of LY235959 or NBQX using the regression equation

\[ F_{\text{Dopamine}} = \beta_{\text{Antagonist}} F_{\text{Baseline}} + \epsilon, \]

where \(F_{\text{Dopamine}}\) is frequency after L-DOPA injection in either “on” or dyskinesia state, \(F_{\text{Antagonist}}\) is frequency after the local antagonist injection, and \(F_{\text{Baseline}}\) is frequency before antagonist injection. All analyses are described in details in the Supplemental Information.

**Striatal Infusion of Antagonist**

LY235959 (0 mM) was dissolved in aCSF, and 10 μL of the solution distributed among five sites (2 μL/site) was infused unilaterally into the posterolateral putamen in three animals (see the criteria for selection of infusion parameters in the Supplemental Information). Ten minutes following the completion of infusion, the animal received s.c. injection of L-DOPA at the selected suboptimal dose. Because of the invasiveness of the procedure, each animal could receive only two infusions, one infusion of LY235959 and another infusion of vehicle alone (aCSF10 μL) as control. Movies S1 and S2 are accompanied by movie legends provided in the Supplemental Information. All details of the procedure and the behavioral assessment after brain infusion or systemic administration of antagonist are provided in the Supplemental Information.

**Statistical Analysis of Behavioral Data**

Behavioral responses were graded within wide ranges, and values included no integers, so data formed quantitative variables, which were analyzed using ANOVAs for repeated-measures (p < 0.05) followed by post hoc Fisher’s protected least significant difference (PLSD) tests. In all analyses, data distribution and variance homogeneity were examined, and appropriate corrections applied. Further details are provided in the Supplemental Information.

**SUPPLEMENTAL INFORMATION**

Supplemental Information includes Supplemental Experimental Procedures, six figures, one table, and two movies and can be found with this article online at https://doi.org/10.1016/j.celrep.2017.12.095.

**ACKNOWLEDGMENTS**

This work was supported by NIH grants NS045962 and NS073994 (S.M.P.) and NS036854 and NS065371 (S.F.T.); SUNY Albany Research Foundation, and National Science Foundation (NSF) grant IOS1653865 (A.S.); and National Center for Research Resources (NCRR) grant RR001066 and Office of Research Infrastructure Programs (ORIP/OD) grant OD011132 (Yerkes National Primate Research Center). We thank Dr. Yoland Smith for advice on histological parameters and Bhagya Dyavar Shetty for technical assistance in the production and care of the primate model with advanced PD.

**AUTHOR CONTRIBUTIONS**

REFERENCES


Cell Reports 22, 941–952, January 23, 2018