Nucleotide Substrate Specificity of Anti-Hepatitis C Virus Nucleoside Analogs for Human Mitochondrial RNA Polymerase


Center for AIDS Research, Laboratory of Biochemical Pharmacology, Department of Pediatrics, Emory University School of Medicine, Atlanta, Georgia, USA; Cocrystal Pharma, Inc., Tucker, Georgia, USA

ABSTRACT Nucleoside analog inhibitors (NAIs) are an important class of antiviral agents. Although highly effective, some NAIs with activity against hepatitis C virus (HCV) can cause toxicity, presumably due to off-target inhibition of host mitochondrial RNA polymerase (POLRMT). The in vitro nucleotide substrate specificity of POLRMT was studied in order to explore structure-activity relationships that can facilitate the identification of nontoxic NAIs. These findings have important implications for the development of all anti-RNA virus NAIs.

KEYWORDS antiviral agents, hepatitis C virus, toxicity

Received 7 March 2017 Returned for modification 20 March 2017 Accepted 20 May 2017 Accepted manuscript posted online 30 May 2017


Copyright © 2017 American Society for Microbiology. All Rights Reserved.
Address correspondence to Raymond F. Schinazi, rschina@emory.edu.
high levels of 2'-C-methyl-GTP intracellularly, inhibited mitochondrial RNA transcription (17). Interestingly, treatment with a 2,6-diaminopurine nucleotide (DAPN) prodrug, an investigational anti-HCV agent that also generates 2'-C-methyl-GTP (18), did not have similar deleterious effects. Lower levels of intracellular NAI-TP accumulation were proposed to account for the distinct cytotoxicity profiles of these compounds (17).

Considering the importance of safety in developing novel ribonucleoside analogs as inhibitors of HCV and other RNA viruses, we explored the nucleotide substrate specificity of POLRMT. Here we report on the in vitro POLRMT incorporation profiles of over 40 rNTP analogs, in order to shed light on structure-activity relationships that may lead to the identification of nontoxic NAIs. NAI-TPs that were not substrates for this enzyme were further examined for antiviral activity. Knowledge gained from this study has important implications for the development of antiviral NAIs not only for HCV but for all RNA viruses.

**Incorporation of nucleoside analogs by POLRMT.** In order to examine the nucleoside substrate specificity of POLRMT, we evaluated the incorporation profiles of 41 nucleosides with various chemical modifications on the ribose or base moiety (the chemical structures are summarized in Fig. 1). As described previously (14, 17), 125 nM purified POLRMT enzyme was incubated with 500 nM 5'-radiolabeled RNA/DNA primer/template that allowed the incorporation of A, C, G, or U nucleotide analogs at position +1. Each nucleoside triphosphate (NTP) analog (100 μM) was incubated for 2 h at 30°C with the POLRMT-RNA/DNA complex containing the appropriate DNA template. The amount of NAI-TP incorporation was normalized to that observed with the corresponding natural nucleotide substrate (Fig. 2). We determined that most modifications on the ribose ring of nucleotide analogs did not completely neutralize incorporation by POLRMT. As depicted in Fig. 2A, the addition of 2'-O-methyl, 2'-fluoro, or 2'-amino modifications did not consistently affect POLRMT incorporation for all nucleotides tested. Consistent with previous findings (14, 15), the addition of a C-methyl group at the 2' position generally led to notable reductions in incorporation for C, G, and U analogs. Nucleotides harboring this modification have been examined for their antiviral activities against HCV (19, 20) and dengue virus (21). Interestingly, the effect of the 2'-C-methyl modification was especially pronounced for 2'-C-methyl-UTP, whose incorporation was reduced to 10% of that of natural UTP, suggesting that U analogs may be particularly vulnerable to chemical modifications with regard to POLRMT incorporation. This is in agreement with the observation that 2'-C-methyl-2'-fluoro-UTP, the active metabolite of sofosbuvir, is an exceedingly poor substrate for POLRMT (Fig. 2A).

Under the conditions tested, we found that 3'- or 2'-deoxynucleoside triphosphates (dNTPs) were generally good substrates for POLRMT (with the exception of 3'-dCTP) (Fig. 2B). However, the simultaneous removal of both hydroxyl groups (yielding 2',3'-dideoxynucleoside triphosphates [ddNTPs]) completely abrogated incorporation, suggesting that the presence of at least one hydroxyl group is essential for NAI-TP incorporation. Our data are consistent with previous observations that little discrimination exists against sugar-modified nucleotides (22). The observed promiscuity of POLRMT has implications for the fidelity of this enzyme during mitochondrial RNA transcription.

Considering that 2'-dNTP analogs were substrates for POLRMT, we next asked whether deoxynucleoside analog inhibitor triphosphates (dNAI-TPs) active against DNA viruses or retroviruses might also be substrates for POLRMT (see Fig. 1B for chemical structures). As expected, diaminopurine dioxolane triphosphate (DAPD-TP), guanosine dioxolane triphosphate (DXG-TP), AZT-TP, and entecavir triphosphate (ETV-TP) were not incorporated by POLRMT. Surprisingly, we found 30% and 36% incorporation for lamivudine triphosphate (3TC-TP) and ganciclovir triphosphate (GCV-TP), respectively, at 100 μM NTP (Fig. 2B). Although extensive literature on the impact of dNAI-TPs on mitochondrial DNA synthesis exists, it is worth noting that little information on the role of these compounds with regard to interference with mitochondrial RNA transcription in cells is available.
FIG 1 Chemical structures of NAI-TPs. (A) Structures of NAI-TPs with modifications on the ribose moiety. Compounds are grouped according to the base moiety. (B) Structures of dNAI-TPs with anti-HIV, anti-HBV, or anti-HSV activity. (C) Structures of rNTP analogs with modifications on the base moiety.
In our search for nucleoside analogs that are not substrates for POLRMT but are active against the viral RNA polymerase of HCV, we next investigated the incorporation profiles of NTP analogs harboring various base moiety modifications (Fig. 1C). Of the nine compounds tested, we found that the majority of modifications at positions 4, 5, 6, and 8 of purines and pyrimidines did not significantly reduce incorporation by POLRMT.
Similarly, NTP analogs containing bulky modifications (such as 8-azido-ATP, 4-thio-UTP, and 5-bromo-UTP) were readily incorporated by POLRMT. The combined presence of 2’-C-methyl and 2-fluoro modifications resulted in poor POLRMT incorporation (12% of that of the natural ATP substrate). We reported previously on the anti-HCV activity of this compound (23); the phosphoramidate prodrug of 2’-C-methyl-2-fluoro-ATP was found to inhibit HCV replicons with submicromolar activity.

The addition of the NHOH chemical group at position 4 of the pyrimidine ring (yielding 4-N-OH-CTP) was observed to reduce incorporation by POLRMT to 50% of that of CTP (Fig. 2C). The addition of the 2’-C-methyl group further reduced incorporation, to 20% of that of CTP. Both 4-N-OH-CTP and 2’-C-methyl-4-N-OH-CTP demonstrated anti-HCV activity in cell culture (24, 25).

Incorporation of N1-methyl-GTP by POLRMT. We identified N1-methyl-GTP as a NTP analog with minimal incorporation by POLRMT. Considering that the addition of the N1-methyl group on the base moiety reduced POLRMT incorporation to 10% of that of the natural GTP substrate (Fig. 2C), N1-methyl-GTP was selected for further analysis with regard to incorporation by viral nonstructural protein 5B (NS5B) RNA polymerase. NS5B incorporation assays were performed as described previously (17). NS5B-RNA/RNA complexes were incubated with increasing concentrations of GTP or N1-methyl-GTP. Single-nucleotide incorporation was observed over time and visualized on a denaturing polyacrylamide gel (Fig. 3), with the 5’-radiolabeled 9-mer primer extended to a 10-mer product. As expected, the natural GTP substrate was rapidly incorporated, with an apparent dissociation constant ($K_{d,app}$) value of 4.1 ± 1.6 µM for GTP (17). Increasing concentrations of N1-methyl-GTP (15.6 µM to 500 µM) were incubated with the preformed NS5B-RNA/RNA complex as described above. The $K_{d,app}$ value for N1-methyl-GTP was estimated to be >100 µM.

We next asked whether RNA extension could occur after N1-methyl-GTP incorporation by the NS5B enzyme. NAI-TP incorporation was assessed in the presence of 10 µM CTP, ATP, and UTP and increasing amounts of N1-methyl-GTP (Fig. 4A). We found that, as N1-methyl-GTP levels increased, increases in full-length 20-mer product levels were observed, suggesting that nucleotide extension can occur following N1-methyl-GTP incorporation (Fig. 4B, left). As expected, full-length RNA synthesis was more

**FIG 3** Incorporation profile for N1-methyl-GTP. (A) Increasing concentrations of GTP (0.7 µM to 180 µM) were incubated with a preformed NS5B-RNA/RNA complex, and nucleotide extension was measured over time. The extended 10-mer RNA product was visualized on a 20% denaturing acrylamide gel. Rates of incorporation at various nucleotide concentrations were plotted as described previously, in order to obtain an apparent dissociation constant ($K_{d,app}$) value of 4.1 ± 1.6 µM for GTP (17). (B) Increasing concentrations of N1-methyl-GTP (15.6 µM to 500 µM) were incubated with the preformed NS5B-RNA/RNA complex as described above. The $K_{d,app}$ value for N1-methyl-GTP was estimated to be >100 µM.
readily observed with low levels of GTP (Fig. 4B, right). Overall, these data suggested that, although N1-methyl-GTP was a poor substrate for the NS5B enzyme, its incorporation did not result in RNA chain termination. We next asked whether N1-methyl-GTP could inhibit RNA polymerization in the presence of competing GTP. We found that, when 1 μM GTP was present in addition to 10 μM CTP, ATP, and UTP, N1-methyl-GTP had no effect on full-length RNA synthesis (Fig. 4C). This is in contrast to the control compound 2'-C-methyl-GTP, which inhibited RNA synthesis with a 50% inhibitory concentration (IC50) of 3.3 ± 0.5 μM (average ± SD of two separate experiments).

In conclusion, in this study we examined the in vitro nucleotide substrate specificity of POLRMT. To our surprise, we found that the POLRMT active site was relatively tolerant in incorporating most of the NTP analogs tested. Several anti-HCV NAI-TPs were identified as poor substrates for POLRMT. In conclusion, the information on the NAI-TP incorporation profile of POLRMT presented herein sheds light on the biochemical properties of this enzyme active site and can inform future ribonucleotide analog drug design for all RNA viruses.
ACKNOWLEDGMENTS

This work was supported in part by NIH grant 5P30-AI-50409 (Center for AIDS Research) (to R.F.S.). M.E. is the recipient of a CIHR-NCRTP postdoctoral fellowship award and an American Liver Foundation postdoctoral fellowship award.

We thank Sijia Tao for performing the cellular pharmacology studies and Louise McCormick for conducting the influenza virus and respiratory syncytial virus assays.

REFERENCES


atatis C virus (HCV) and are bioconverted intracellularly to bioactive 2,6-diaminopurine and guanosine 5′-triphosphate forms. J Med Chem 58:3445–3458. https://doi.org/10.1021/jm501874e.