Langerhans cell sarcoma arising from chronic lymphocytic lymphoma/small lymphocytic leukemia: Lineage analysis and BRAF V600E mutation study

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Introduction

Both interdigitating dendritic sarcoma (IDCS) and Langerhans cell sarcoma (LCS) are derived from myeloid progenitor cells, which are different from precursors for lymphoid neoplasms. However, recent studies have shown that histiocytic/dendritic cell sarcomas and some lymphoproliferative diseases, such as follicular lymphoma, chronic lymphocytic leukemia/small lymphocytic lymphoma (CLL/SLL) and hairy cell leukemia, may occur in the same individual and interestingly, these neoplasms are clonally related. This phenomenon is dubbed “transdifferentiation”. [1-8] There have been several reports showing synchronous IDCS and CLL/SLL; however, only one case of concurrent LCS and CLL/SLL has been documented in the English literature.[9] Here we present a case in which LCS arose from CLL/SLL and the two diseases appeared to share the same clonality.

The BRAF V600E mutation (T1799A) has been shown to be a molecular change underlying the pathogenesis
of many malignancies, the most notable one being melanoma. A recently developed monoclonal antibody vemurafenib that targets BRAF V600E mutant has been approved to treat late-stage or unresectable melanomas and the outcome is very encouraging. Considering the effectiveness of this drug in melanoma treatment, it would be interesting to look into its potential effect on other cancers bearing the same mutation. Recent studies have demonstrated that 38-57% of Langerhans cell histiocytosis (LCH) harbor the BRAF V600E mutation. However, whether LCS shares the same molecular defect remains to be addressed. Here we present the first LCS case that carries BRAF V600E mutation.

Materials, Methods, and Results
The patient was a 68-year-old female with a past medical history significant for papillary thyroid carcinoma status post-thyroidectomy. Six years ago, she was diagnosed with CLL and followed with watchful waiting. Four years ago the patient received eight cycles of Rituxan and fludarabine due to disease progression. The treatment brought down the WBC count to 5.9 × 10⁹/L with 9% lymphocytes. However, 1 year ago her CLL relapsed and three doses of Rituxan were administered. Unfortunately she developed deep vein thrombosis and pulmonary embolism after chemotherapy and was put on anticoagulation therapy since. On the most recent admission, she complained about general malaise, poor appetite, shortness of breath, and progressively enlarging right inguinal lymph nodes. Blood workup showed a WBC count of 12.6 × 10⁹/L with 1% of lymphocytes. One lymph node from her right groin was biopsied. The flow cytometry results (data not shown), histologic examination, and immunohistochemistry confirmed CLL/SLL in this lymph node with atypical lymphocytes positive for CD20, CD5, and CD23 [Figure 1d, e and f]. Surprisingly, population of large epithelioid cells was positive for CD20, CD5, and CD23 [Figure 1d, e and f].

Once the two diagnoses were confirmed, the next question was whether the LCS and the CLL/SLL were two independent diseases that happened to occur at the same anatomic site or they were related. To address this question, we performed a fluorescence in situ hybridization (FISH) analysis using a CLL panel probes (centromere6, 6q23, 11q14, 13q34, centromere12, IGH, and 17p13; Vysis CLL FISH probe kit, Abbott Laboratories. Abbott Park, Illinois, USA). The results showed that both the CLL/SLL [Figure 1j] and the LCS cells [Figure 1k] lost the 6q23 signal, suggestive of same clonality of these two populations.

To further understand the genetic changes in LCS cells, we investigated whether the LCS cells carry a BRAF V600E mutation. This was prompted by recent studies showing this mutation in up to 38–57% in LCH. DNA was extracted from the formalin fixed paraffin embedded tissue. The BRAF gene was amplified by PCR with forward primer 5’- TGA AGA CCT CAC AGT AAA AAT CTG ATC AC-3’ and reverse primer 5’- /5Biosg/TCC AGA CAA CTG TTC AAA CTG AT -3’ (Integrated DNA Technologies, Inc, Coralville, Iowa). The PCR product was sequenced with primer 5’- TGA TTT TGG TCT AGC TAC A -3’ on PyroMark Q96 (Qiagen) according to manufacturer’s instructions. Nucleotides were dispensed with the following sequence: ACGTACGATC. The V600E mutation was identified by a peak at the fifth adenosine position and the mutation was harbored in 25% of total DNA (T to A point mutation, 25%, Figure 2d), suggestive of a heterozygous mutation (LCS was about 50% of the total lymph node). The result, for the first time, confirmed the BRAF V600E mutation in LCS. Furthermore, although the gold standard for detection of BRAF V600E mutation is PCR, a recently developed monoclonal antibody VE1 shows high sensitivity and specificity for this mutation, and has been widely used in place of PCR for research purpose. Therefore, we also performed an immunostain with the VE1 antibody (Spring Bioscience, Pleasanton, CA 94566) in this lymph node. Again, the LCS cells, but not in the CLL/SLL cells, showed positivity for VE1 [Figure 2c], suggesting BRAF V600E mutation in LCS. A negative control [Figure 2a] and a PCR confirmed positive melanoma control [Figure 2b] for this antibody are also shown. This result was consistent with the
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In the patient's case, a molecular study, in which only 25% of DNA carried mutation (heterozygous mutation for LCS, negative for CLL/SLL). Following the diagnosis, the patient received one cycle of salvage chemotherapy with DHAC (Dexamethasone, Doxorubicin, ARA-C, and Carboplatin) but failed to respond. She complained increasing abdominal pressure and girth and a diagnostic laparoscopy revealed multiple nodules (presumed to be CLL/SLL) scattered throughout the small bowel causing adhesions and obstruction. The large mass in the right inguinal region kept growing. The patient decided to check into hospice and received palliative care since then.

Discussion

Feldman et al. provided the first evidence of clonal evolution or transdifferentiation of B-cell lymphoma into histiocytic/dendritic cell sarcoma, in which both neoplasms shared a common IGH gene rearrangement and carried the same BCL2/IGH translocation. In the following years, the list of B-cell malignancies capable of transdifferentiation had been expanded to extranodal marginal zone lymphoma (MALT lymphoma), splenic marginal zone lymphoma, hairy cell leukemia and CLL/SLL, etc. This phenomenon is most likely attributed to the lineage plasticity of the B-cell neoplasms. Although the exact mechanisms are not fully elucidated, several hypotheses have been proposed. The first possibility is of the direct transdifferentiation of neoplastic B-cells into malignant dendritic cells. This is supported by the experimental data showing that enforced expression of some transcription factors such as C/EBPα and C/EBPβ can lead to direct transdifferentiation of mature B-cells into macrophages. A second hypothesis is that this transformation is achieved through multiple steps, which mainly involve de-differentiation of neoplastic lymphocytes to early progenitors, followed by re-differentiation of progenitor cells to dendritic cells. The potential for de-differentiation and re-differentiation of B-cells has been well evidenced. For example, in vitro deletion of PAX-5, an important regulator for B-cell development, can lead to de-differentiation of mature B-cells into progenitor cells, which subsequently differentiate to T-cells. Furthermore, it has been found that, in the case of LCS arising from B lymphoblastic leukemia, the LCS cells seem to express ID2 (a key factor in dendritic cell development) much more strongly than PAX-5. The ebbing of PAX-5 and the concurrent rising of ID2 may underlie this de-differentiation.

Although the role of BRAF mutation in LCS oncogenesis has never been studied, it is well known that BRAF mutation is associated with activation of the mitogen-activated protein kinase (MAPK) pathway in melanoma. As discussed earlier, enhanced expression of transcription factors C/EBPα and C/EBPβ may lead to transdifferentiation of lymphocytes to dendritic/Langerhans cells. A recent study by Ignatova et al. suggests that there may be crosstalk between C/EBPα and C/EBPβ and the MAPK pathway. These authors demonstrated that C/EBPα and C/EBPβ control basal transcription through the

Figure 1: Synchronous and clonally related LCS and CLL/SLL in a lymph node. (a) Sheets of small SLL/CLL cells admixed with large LCS cells in clusters (100×). (b) The LCS cells show high mitotic activity (400×). (c) The LCS cells demonstrate a much higher proliferation index (MIB-1, 40%) than that of CLL/SLL cells (20%). (d–i) The CLL/SLL cells show immunoreactivity for CD20 (D), CD5 (E) and CD23 (F) while the LCS cells are positive for CD1a (G), S100 (H) and langerin (I). (j) and (k) A FISH analysis using probes for centromere6 (green) and 6q23 (red) revealed the CLL/SLL (J) and the LCS cells (K) both lost the 6q23 signal.

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p38 MAPK-mediated pathway in vitro. Whether similar mechanism is responsible for the potential role of BRAF mutation in the transformation of CLL/SLL to LCS needs to be addressed by further investigation.

To date, there has been only one documented synchronous CLL/SLL and LCS case. In the present case, the LCS component was identified when sheets of large epithelioid cells distinctly different from the background CLL cells were found and showed positivity for Langerhans cell antigen markers, that is, CD1a, S100 and langerin. LCH or Langerhans cell hyperplasia were ruled out by the high mitotic count, the high proliferation index (MIB-1) and focal necrosis in these epithelioid cells. Of note, CD1a immunostain only showed partial positivity. It is uncertain whether the extent of CD1a positivity is reversely proportional to the aggressiveness of LCS, although the published LCS cases seem strongly positive for CD1a.

The FISH analysis with CLL panel showed that among six sets of probes, five of them (11q23, 13q14, and 13q34, centromere12, IGH, 17p13) showed normal signals both in CLL/SLL and LCS, but the sixth set (centromere6 and 6q23) showed loss of 6q23 signal in both CLL/SLL and LCS, which suggested that the LCS shared the same clonality with the CLL/SLL. So far there is only one publication that investigated cytogenetic changes of histiocytic/dendritic cell sarcomas arising from...
SLL/CLL.[8] In that study, all six cases of histiocytic/dendritic sarcomas (including one LCS) showed 17p abnormalities but only two cases of corresponding SLL/CLL had the same change. The authors proposed that the high frequency of 17p abnormalities (contains TP53 gene) might be a potential risk factor for transformation of CLL/SLL. In this case, 6q23 deletion, but not 17p abnormalities, was detected in both the CLL/SLL and the LCS components. Studies showed that 6q23 deletion was found in 5% of CLL patient.[20] The gene in this region, c-MYB, controls the proliferation and differentiation of hematopoietic stem and progenitor cells. Loss-of-function study on c-MYB shows that there is a ten-fold increase in the number of hematopoietic stem cells.[21, 22] The exact role of c-MYB in leukemogenesis and transdifferentiation is not clear.

BRAF V600E mutation has been shown in many types of malignancy and the best known ones are melanoma and papillary thyroid carcinoma. Interestingly, recent studies showed that BRAF V600E mutation occurred in 38% to 57% of LCH and 54% of Erdheim–Chester disease, but not in Rosai–Dorfman disease, juvenile xanthogranuloma, histiocytic sarcoma, xanthoma disseminatum, IDCS, and necrobiotic xanthogranuloma.[4, 3] Our result demonstrates, for the first time, that LCS can bear the BRAF V600E mutation. It would be immature to predict the significance of such a molecular defect in LCS with only one case, but our finding certainly opens an opportunity to expand our knowledge of the genetics and tumorogenesis of LCS. Notably, a BRAF V600E mutant inhibitor vemurafenib has been approved for treating advanced melanoma. This novel drug is able to improve the progression-free survival of unresectable melanomas drastically.[24] Dramatic efficacies of this drug in both multisystemic and refractory Erdheim–Chester disease and LCH that harbor BRAF V600E mutation have also been reported recently.[23] Therefore, it is reasonable to deduce that similar treatment would be applicable in the management of LCS harboring the same mutation.

Conclusion

Taken together, we reported a case of synchronous and clonally related LCS arising from CLL/SLL. More importantly, for the first time, BRAF V600E mutation was detected in LCS, which opened the door for potential clinical trial of the BRAF V600E mutant inhibitor vemurafenib on LCS with this mutation. Larger case series, of course, would be needed in order to achieve this goal.

References


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