**In Vitro** Assessment of the Immunological Significance of a Human Monoclonal Antibody Directed to the Influenza A Virus Nucleoprotein

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Influenza A viruses cause annual epidemics and occasionally pandemics. Antibodies directed to the conserved viral nucleoprotein (NP) may play a role in immunity against various influenza A virus subtypes. Here, we assessed the immunological significance of a human monoclonal antibody directed to NP in vitro. This antibody bound to virus-infected cells but did not display virus-neutralizing activity, complement-dependent cell cytotoxicity, or opsonization of viral antigen for improved antigen presentation to CD8+ T cells by dendritic cells.
NP, horseradish peroxidase, and 3,3′,5,5′-tetramethylbenzidine, plates were read with a Tecan Infinite 200 reader (Tecan, Giessen, the Netherlands). In addition to the controls provided by the manufacturer, negative-control ferret serum and positive-control human serum were used as controls. A proportional difference in optical density (OD) value between sample and negative control of <0.6 was interpreted as positive as indicated by the manufacturer.

To test whether the monoclonal antibody was able to bind to NP present on the surface of influenza A virus-infected cells, A549 adenocarcinoma human alveolar basal epithelial cells, Epstein-Barr virus (EBV)-transformed human B cells, and Madin-Darby canine kidney (MDCK) cells were inoculated with influenza A(H3N2) virus RESVIR-9 with a multiplicity of infection (MOI) of 3. Influenza A(H3N2) virus RESVIR-9 is an influenza vaccine strain obtained after reassortment between influenza virus A/Nanchang/933/95 (HA, NA, NP) and influenza virus A/PR/8/34 (the remaining five gene segments). After incubation for 16 to 18 h at 37°C with 5% CO2, cells were trypsinized and subsequently incubated for 30 min with 10, 20, and 40 µg/ml of MoAb NP or isotype control (BD, Alphen a/d Rijn, the Netherlands) at 4°C. After washing, cells were stained with fluorescein isothiocyanate (FITC)-labeled MoAb directed against human IgG (Dako, Glostrup, Denmark). Subsequently, cells were fixed with Cytofix (BD, Alphen a/d Rijn, the Netherlands) and then assessed by flow cytometry with a FACSCanto II cytometer. The data were analyzed with FACSDiva software (BD, Alphen a/d Rijn, the Netherlands). All experiments were performed in duplicate, and uninfected cells were used as negative controls. As shown in Fig. 1, MoAb NP bound to virus-infected cells of all three cell lines, while no binding was observed to uninfected control cells. In addition, virtually no binding of the isotype control antibodies was observed (Fig. 1A). In addition, time course experiments indicated that NP could already be detected on the surface of infected cells within 1 h after inoculation (data not shown), suggesting the binding of free NP present in the inoculum. The findings confirm those originally obtained by Yewdell et al., who demonstrated the presence of NP on the surface of infected P815 cells by using a MoAb NP of murine origin (15).

In addition, the cellular localization of NP in virus-infected A549 cells was analyzed by confocal microscopy using a Zeiss LSM700 microscope in combination with Zeiss LSM Zen 2010 software (Carl Zeiss, Sliedrecht, the Netherlands). The same procedure as that described for flow cytometry was used, except that cells were also incubated with PKH-26 (Sigma-Aldrich, St. Louis, MO) to visualize cellular membranes. Also using this method, binding of MoAb NP to extracellular NP was observed on virus-infected cells, while virtually no binding was observed with the isotype control (Fig. 1B). Of interest, binding of MoAb NP to NP was not evenly distributed over the whole cell surface but displayed a patchy pattern, which is in accordance with the pattern of NP staining of infected mouse cells with a mouse monoclonal antibody specific for NP (15). Permeabilization of the infected cells prior to incubation with the NP-specific antibody resulted in the typical nuclear and cytoplasmic staining of NP, and also on these cells, binding of MoAb NP on the surface of cells was observed (data not shown).

To assess the broadness of reactivity of MoAb NP, A549 cells were inoculated with influenza A viruses A/Puerto Rico/8/1934 (H1N1), A/Swine/Netherlands/25/1980 (H1N1), A/Netherlands/246/2008 (H1N1), A/Netherlands/602/2009 (H1N1pdm09), A/HongKong/2/1968 (H3N2), and A/Netherlands/348/2007 (H3N2) and influenza B viruses B/Netherlands/151/2001 (Victoria lineage) and B/Netherlands/080/2002 (Yamagata lineage) at an MOI of 3 and subsequently tested by flow cytometry as described above. FITC-labeled MoAbs directed against the NP of either influenza A or influenza B virus (Imagen immunofluorescence test; Thermo Fisher Scientific, Waltham, MA) were used as positive controls.

Binding of the MoAb NP was observed to cells infected with any of the influenza A viruses but not to those infected with influenza B viruses. This confirms the broad reactivity of this antibody for influenza A viruses. However, differences were observed in the extent of binding, which might be due to differences in the biology of infection or due to differences in the affinity of the antibody for the respective NPs (Fig. 2) (20, 21).

Although the MoAb NP is highly cross-reactive and can bind to virus-infected cells, the role of this NP-specific MoAb in immunity against influenza remains elusive. Therefore, we performed in vitro experiments to assess possible roles of the MoAb NP in immunity against influenza A virus.

First, the virus-neutralizing capacity of the MoAb NP was
tested using the infection reduction assay with influenza A viruses RESVIR-9 (H3N2) and A/PR/8/34 (H1N1) as described previously (22). Using this assay, which is more sensitive than the conventional virus neutralization assay (22), no reduction of infection was observed using 2.5, 5.0, 10.0, and 20.0 μg/ml of MoAb NP in contrast to postinfection ferret sera raised against the homologous strains that were used as positive controls. These findings indicate that this NP-specific MoAb is not capable of preventing infection of cells, which confirms previous results obtained with murine monoclonal antibodies specific for NP (Fig. 3A) (15).

Second, we tested whether the MoAb against NP could mediate complement-dependent cytotoxicity (CDCC) as described previously for antibodies against the matrix 2 protein and the conserved region of the stalk protein of the hemagglutinin (10, 23). To this end, A549 cells were infected with influenza A virus RESVIR-9 (H3N2) at an MOI of 3 and 30,000 cells were incubated per well of a V-bottom 96-well plate for 3 h at 37°C in a humid atmosphere of 5% CO2. An interval of 3 h was used since it was demonstrated in pilot experiments that the recognition of NP on the surface of cells reached a plateau within 3 h after inoculation and this relatively short duration of infection resulted in the highest number of live cells with NP on the surface. Subsequently, cells were incubated with either 0, 20, or 40 μg/ml of MoAb NP for 30 min in phenol red-free RPMI medium containing 5% fetal calf serum, 100 IU/ml penicillin, 100 μg/ml streptomycin, 2 mM glu-
tamine, and 20 μM β-mercaptoethanol. As a positive control, we used purified IgG of a human donor in which the presence of antibodies against the hemagglutinin of the RESVIR-9 virus was demonstrated using the hemagglutination inhibition (HI) assay (HI titer of 40). Then, 1:20-diluted, Low-Tox guinea pig complement (Cedarlane, Burlington, Ontario, Canada) was added, and after incubation for 2 h, complement-dependent lysis of cells was determined using the Cytotox 96 nonradioactive cytotoxicity assay (Promega, Leiden, the Netherlands) according to the manufacturer’s recommendations. All experiments were performed in quintuplicate, and uninfected cells were used as negative controls. Antibody-specific lysis was calculated by subtracting the percent specific lysis measured for uninfected cells from the percent specific lysis of infected cells. Percent specific lysis was calculated using the following formula: (% lysis by antibody and complement — % lysis by complement only)/(% maximum lysis — % lysis by complement). Also in this assay, no activity of the MoAb NP was observed despite the use of high antibody concentrations of MoAb NP, in contrast to equivalent volumes of the positive-control IgG (Fig. 3B). These findings suggest that this antibody to the NP does not mediate CDC, in contrast to antibodies against the hemagglutinin or the matrix 2 protein (10, 23). The differences are most likely explained by the fact that the hemagglutinin and matrix 2 protein are anchored in the cell membrane, while this has not been demonstrated and most likely is not the case for the nucleoprotein. Of interest, Yewdell and colleagues observed low levels of CDC using a murine MoAb directed against the NP, which is in contrast to our observations (15). This might be related to differences between the assays used or the MoAb NP used.

It has been suggested that the protective effect of NP-specific antibodies observed in mice can be attributed to the formation of immune complexes that are processed by antigen-presenting cells, which subsequently results in accelerated and enhanced virus-specific CD8+ T cell responses. Indeed no protection was observed in the absence of memory T cells (14, 24). Therefore, we wished to investigate the effect of MoAb NP on opsonization of viral antigen, possibly resulting in improved uptake, processing, and antigen presentation to virus-specific CD8+ T cells by professional antigen-presenting cells. To test this possible mode of action, a split virion preparation of influenza A virus RESVIR-9 was produced by treating concentrated and purified virus stocks with decanoyl-N-methylglucamide (Mega-10; Sigma-Aldrich, Zwijndrecht, the Netherlands) for 30 min. After dialysis against phosphate-buffered saline (PBS) for 24 h, the protein concentration was determined using the BCA protein assay kit (Thermo Scientific Pierce Protein Biology Products, Rockford, IL), and full inactivation of the virus was confirmed as described previously (25). An 0-, 0.5-, or 1-μg amount of the split virion preparation was incubated with or without 1 μg MoAb NP for 1 h at 37°C and subsequently added to 100,000 human monocyte-derived dendritic cells (DC) from HLA-B*3501+ donors, which were prepared as described previously with monocytes derived from two different healthy blood donors (26). After incubation of DC with antigen and with or without MoAb NP for 16 to 18 h at 37°C, 30,000 cells of a CD8+ T cell clone specific for the HLA-B*3501-restricted epitope NP411-426 (LPFEEKSTMV) were added. Following incubation for 6 h in the presence of Golgistop (BD, Alphen a/d Rijn, the Netherlands), the production of gamma interferon (IFN-γ) by cells of the CD8+ T cell clone was assessed by flow cytometry using a FACScanto II cytometer. The data were analyzed by FACSDiva software essentially as described previously, and specific production of IFN-γ was calculated by subtracting the percentage of IFN-γ+ T cells of medium controls from the DC incubated with the split virion preparation (28). Stimulation of the NP-specific CD8+ T cells with dendritic cells incubated with the split virion preparation induced IFN-γ production. However, preincubation of the split virion antigen preparation with MoAb NP did not increase the percentage of IFN-γ-producing T cells (Fig. 3C and D). These findings suggest that this MoAb NP does not improve uptake, processing, and antigen presentation to virus-specific CD8+ T cells by DC in vitro. In addition, no increased upregulation of costimulatory molecules CD80, CD83, and CD86 was observed after incubation of DC with split virion and MoAb NP compared to that with DC incubated with the split virion preparation only (data not shown). However, MoAb NP might improve endocytosis and presentation via the major histocompatibility complex (MHC) class II molecule to CD4+ T cells. Collectively, we have demonstrated that MoAb NP bound to viral nucleoprotein on the surface of virus-infected cells, but we were not able to demonstrate biological activity of immunological significance for this human antibody directed to the NP in various in vitro test systems.

The exact source and trafficking route of NP resulting in the patchy appearance on the surface of virus-infected cells are still unknown. In pilot studies using an NP-specific mouse monoclonal antibody, we observed that NP was already detectable on the surface of infected cells within 1 h after inoculation. Furthermore, we were not able to prevent this appearance by using the protein transport inhibitor brefeldin A, suggesting that it is not an active process, which was already suggested previously (15). Possibly, NP detected on the cell surface is acquired after release by other virus-infected cells or accumulated from the viral inoculum.

Of course, the in vitro test systems used here do not reflect the situation in vivo. In addition, the biological effect of NP-specific antibodies may be dependent on the epitopes that are recognized, and more antibodies against the NP should be tested to address this issue. Furthermore, the heavy chain constant region of the MoAb of the present study was of the IgG1 subclass, which may not be the same as that of the original antibody. However, IgG1, which is the most abundant subclass, is able to activate complement and to bind to Fc receptors on various cells of the immune system (29, 30). It cannot be excluded, however, that other subclasses may display different biological activity, and therefore, more subclasses should be tested. Thus, antibodies against the influenza A virus nucleoprotein might still play a role in heterosubtypic immunity, for example by cooperating with T cells to eliminate virus-infected cells in the lung (14). More research is necessary to address the potential of NP-specific antibodies to afford heterosubtypic immunity in humans, to elucidate their relative contribution to this type of immunity, and to elucidate the mechanisms of protection.

REFERENCES


