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Fungal-derived semiochemical 1-octen-3-ol disrupts dopamine packaging and causes neurodegeneration

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Parkinson disease (PD) is the most common movement disorder and, although the exact causes are unknown, recent epidemiological and experimental studies indicate that several environmental agents may be significant risk factors. To date, these suspected environmental risk factors have been man-made chemicals. In this report, we demonstrate that exposure to genetic, biochemical, and immunological studies that the common volatile fungal semiochemical 1-octen-3-ol disrupts dopamine levels and causes dopamine neuron degeneration in Drosophila melanogaster. Overexpression of the vesicular monoamine transporter (VMAT) rescued the dopamine toxicity and neurodegeneration, whereas mutations decreasing VMAT and tyrosine hydroxylase exacerbated toxicity. Furthermore, 1-octen-3-ol also inhibited uptake of dopamine in human cell lines expressing the human plasma membrane dopamine transporter (DAT) and human VMAT ortholog, VMAT2. These data demonstrate that 1-octen-3-ol exerts toxicity via disruption of dopamine homeostasis and may represent a naturally occurring environmental agent involved in parkinsonism.

Significance

Poor air quality from fungal growth in water-damaged, moldy buildings/residences is correlated with a negative impact on human health. The volatile organic compound 1-octen-3-ol is commonly emitted by molds and is responsible for much of the distinctive moldy odor associated with fungal colonization. Using a Drosophila model, we demonstrate via genetic, biochemical, and immunological studies that 1-octen-3-ol causes dopamine neuron degeneration through disruption of dopamine handling. These data demonstrate that 1-octen-3-ol exerts toxicity via disruption of dopamine homeostasis and may represent a naturally occurring environmental agent involved in parkinsonism. Moreover, it provides possible insights into reported movement disorders associated with human exposure to fungi and their volatile organic compounds.


The authors declare no conflict of interest.

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that low concentrations of a natural fungal volatile organic compound lead to disruption of dopamine homeostasis and also interact with genetic variants in dopamine biosynthesis to increase dopaminergic neurodegeneration.

**Results**

**Exposure to 1-Octen-3-ol Shortens Survival Span and Affects Locomotion of Wild-Type *D. melanogaster***. To evaluate the toxic effects of a low dose (0.5 ppm) of 1-octen-3-ol, we exposed wild-type *w^{1118} melanogaster* flies to a pure form of volatile-phase 1-octen-3-ol. The time to 50% mortality of adult flies exposed to 0.5 ppm of 1-octen-3-ol was ∼19 d (Fig. 1A). Furthermore, exposure of wild-type flies to 0.5 ppm of 1-octen-3-ol caused locomotor defects as assessed by a negative geotaxis assay as early as 24 h, with a further decline in locomotor activity upon continuous exposure to 1-octen-3-ol until day 7 (Fig. 1B).

**Exposure to 1-Octen-3-ol Induces Loss of Dopaminergic Neurons and Decreases Dopamine Levels in Head Extracts.** To test whether 0.5 ppm of 1-octen-3-ol causes loss of dopaminergic neurons, we used transgenic *TH-GAL4; UAS-GFP* adults (18). The morphology and number of posteriorly located dopaminergic neurons in these transgenic fly brains were assessed after a 24-h exposure to 0.5 ppm of 1-octen-3-ol. Exposure to 1-octen-3-ol significantly decreased the number of all of the subgroups of dopaminergic neurons, except for PPL1 (Fig. 2A and B). We then performed HPLC to quantify the effect of 0.5 ppm of 1-octen-3-ol on levels of dopamine and its metabolites. A significant decrease in dopamine levels (28%) was observed concomitantly with an increase in DOPAC levels (40%) compared with control flies. These data suggested that a potential impairment of dopamine handling is associated with exposure to 1-octen-3-ol (Fig. 2C).

**1-Octen-3-ol Exposure Inhibits Dopamine Uptake in HEK-Dopamine Transporter/VMAT Ortholog Cells via Its Action on the Dopamine Transporter.** To determine whether the increased DOPAC observed following exposure to 1-octen-3-ol was the result of impaired dopamine transport, we measured the effect of 1-octen-3-ol on [3H]dopamine (DA) uptake in human embryonic kidney (HEK) cells stably expressing human dopamine transporter (DAT) and human VMAT ortholog VMAT2. We exposed 1, 3, and 10 ppm of 1-octen-3-ol for 2 h to these cell lines and found no significant effect on DAT/VMAT activity at 1 ppm, but 3 and 10 ppm reduced the uptake to a significant extent. Exposure to 10 ppm of 1-octen-3-ol for 2 h reduced uptake by 95% in HEK-DAT cells. The experiment was repeated in HEK-DAT/VMAT2 cells that express human DAT and VMAT2 with 10 ppm, and similar results were observed (Fig. 3A and B). Protein assays confirmed that the loss of uptake was not due to a loss of cells after exposure to 1-octen-3-ol. The near-complete inhibition of DAT prevents the assay from detecting any effect of 1-octen-3-ol on VMAT2. Additionally, methods to measure VMAT2 uptake were not compatible with the airborne exposure setup. Therefore, we took a more direct approach to measuring VMAT2 activity utilizing a stable cell line expressing human VMAT2.

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**Fig. 1.** Common fungal VOC 1-octen-3-ol exposure truncates survival span of wild-type and induces mobility defects. The data were collected daily until all of the flies exposed to 1-octen-3-ol were dead, and the percentage mortality of dead flies was calculated. (A) The 50% mortality for wild-type flies is shown (n = 125), and data are from three independent experiments. (B) Forty-eight-hour-old wild-type flies were exposed to 0.5 ppm of 1-octen-3-ol, and time to cross a 5-cm distance was recorded after 1 and 7 d of continuous exposure. There was a significant difference in the locomotory activity between wild-type, unexposed, and exposed flies assessed with a negative geotaxis assay at days 1 and 7. Error bars represent SEM, and the significant difference between wild-type exposed and unexposed flies is indicated; *P < 0.05,* **P < 0.005.

**Fig. 2.** Effect of 1-octen-3-ol on dopamine pools in head extracts of *Drosophila* flies. (A) Exposure of *TH-GAL4; UAS-GFP* to 0.5 ppm of 1-octen-3-ol for 24 h led to a decrease of GFP-expressing TH neurons in adult brains. The PPM1 and PPL1 subgroups of DA neurons are indicated by an arrow and circle, respectively, in unexposed and 1-octen-3-ol-exposed adult brains. (B) Scoring for the average number of subgroups of dopaminergic neurons for control and 0.5-ppm 1-octen-3-ol-exposed *TH-GAL4; UAS-GFP* adult brains. Except for PPL1, there was a significant decrease in all of the subgroups of dopaminergic neurons in brains exposed to 0.5 ppm of 1-octen-3-ol (n = 10–15). (C) Exposure to 0.5 ppm of 1-octen-3-ol for 24 h caused a decrease in dopamine pools with a subsequent increase in DOPAC levels in head extracts (n = 200 for each group; data represent values from three independent experiments). Error bars represent the SE of the mean, and the significant difference between control and 1-octen-3-ol-exposed brains is indicated; *P < 0.05,** *P < 0.005.
approach to assessing the putative role of VMAT in the effects of 1-octen-3-ol.

**VMAT Mutants Are Sensitive Whereas Transgenic Strains for VMAT Are Resistant to 0.5 ppm of 1-octen-3-ol.** The *Drosophila* VMAT gene contains two splice variants, dVMAT-A and -B, where dVMAT-A is expressed in all dopaminergic, serotonergic, and octopaminergic neurons and functions similarly to mammalian VMAT2 (19, 20). To assess the potential for 1-octen-3-ol to exert dopaminergic toxicity through disruption of dopamine handling in vivo, we first exposed two loss-of-function mutant lines for dVMAT-A, l(2)SH0459/+ and Delta14/+, to 0.5 ppm of 1-octen-3-ol (21). The survival span was significantly reduced to 8 and 10 d in l(2)SH0459/+ and Delta14/+ dVMAT mutants, respectively, compared with 17 d for wild-type flies (Fig. 4A). The unexposed wild-type flies (***) and dVMAT mutant strains l(2)SH0459/+ and Delta14/+ used as controls for these experiments failed to show any death during this period. We then overexpressed UAS-dVMAT in dopaminergic neurons using the dopamine driver TH-GAL4. Upon exposure to 0.5 ppm of 1-octen-3-ol, average survival of overexpressed transgenic flies TH-GAL4-UAS-dVMAT was about 3.5 d longer than that of control flies TH-GAL4-UAS-dVMAT. Similarly, the overexpression of UAS-dVMAT using the pan-neuronal driver elav-GAL4 led to an extension of survival duration by 4 d compared with control flies (Fig. 4B).

HPLC analysis of dopamine and DOPAC levels in the head extracts of flies overexpressing dVMAT in dopaminergic neurons (TH-GAL4; UAS-dVMAT) exposed to 0.5 ppm of 1-octen-3-ol failed to show the alteration of DA and DOPAC levels (Fig. 4C) that was observed in wild-type flies (Fig. 2C). We further determined the protective effect of dVMAT against 1-octen-3-ol by evaluating the status of dopaminergic neurons overexpressing dVMAT. The overexpression of dVMAT prevented the 1-octen-3-ol–mediated loss of dopaminergic neurons (Fig. 4D and E) that was seen in wild-type flies (Fig. 2A and B).

**Heterozygous tyrosine hydroxylase Mutant Flies Are Sensitive to 1-Octen-3-ol and Demonstrate Defective Locomotion Compared with Wild-Type Flies.** Considering that 1-octen-3-ol exposure led to alteration of dopamine homeostasis and dopamine neuron loss, we then investigated whether 1-octen-3-ol mediates the variation in the activity of the gene responsible for dopamine biosynthesis, tyrosine hydroxylase (TH), encoded by *pale* (ple) locus in *Drosophila*. We exposed two heterozygous loss-of-function mutant strains of the ple gene, ple* c and ple* d, known to have mutations in the *tyrosine hydroxylase* gene and to possess low dopamine pools (22) to 0.5 ppm of 1-octen-3-ol. Heterozygous mutant strains ple* c and ple* d exhibited reduced tolerance to 0.5 ppm of 1-octen-3-ol with a 50% mortality obtained at 13.8 and 12.4 d, respectively, compared with wild-type flies exposed to the same concentration of 1-octen-3-ol. Unexposed control flies did not show any death during the observed time period (Fig. 5A).

We also measured movement deficits in wild-type w* , ple* , and ple* mutant flies after 1 and 7 d of exposure to 0.5 ppm of 1-octen-3-ol. After 1 d of exposure to 0.5 ppm of 1-octen-3-ol, there was no significant difference in the average time for exposed wild-type and ple mutants to cross a 5-cm distance. However, after 7 d of continuous exposure to 1-octen-3-ol, the average time required for ple mutants was about 1.5-fold higher than for wild-type flies (Fig. 5B). In the absence of 1-octen-3-ol, both ple mutants demonstrated climbing ability similar to wild type at both time points.
as a powerful signal molecule in insect attraction and deterrence (26–28), and has been hypothesized to contribute to the adverse health effects seen in moldy and water-damaged buildings and houses (29). The exact concentration of any given fungal VOC in water-damaged, moldy buildings is difficult to measure because concentrations vary depending on the ventilation rate, moisture, temperature, and other parameters. However, reported concentrations of 1-octen-3-ol in moldy buildings and classrooms range from 0.2 μg/m^3 (0.00004 ppm) up to 900 μg/m^3 (0.16 ppm) (15, 30–32).

Currently, only limited data are available on the toxicity of 1-octen-3-ol. Kreja and Seidel (33) reported the cytotoxic and mutagenic potential of 6.4 mM (820 ppm) liquid-phase 1-octen-3-ol for A549 human lung cell cultures. The lethal concentration 50% (LC50) for the A549 human lung cell line determined with the colony-forming ability assay was 3.8 mM (486 ppm), whereas with 3(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium 50 bromide and methylene blue assays it was 3.4 mM (435 ppm) and 2.1 mM (268 ppm), respectively. More recently, we showed that 100 ppm of volatilized 1-octen-3-ol was 80 times more toxic than toluene in human embryonic stem cells (34). Human volunteers exposed to 1.9 ppm of 1-octen-3-ol for 2 h experienced irritative symptoms, along with increases in inflammatory markers, eosinophil cationic protein, myeloperoxidase, lysozyme, and albumin in the nasal secretions (35). In this report, survival duration was truncated in *Drosophila* exposed to 1-octen-3-ol vapor at lower concentrations (0.5 ppm). Exposed flies exhibited locomotory deficits and loss of dopamine neurons, which were accompanied by loss of dopamine and an increase in 3,4-dihydroxyphenylacetic acid (Figs. 1–3).

The increased DOPAC levels suggest that 1-octen-3-ol may interfere with the packaging of dopamine into synaptic vesicles, which increases autooxidation of dopamine and generation of oxidative damage and reactive dopamine metabolites such as 3,4-dihydroxyphenylacetaldehyde (36–38). Indeed, we previously found that an increase in reactive oxygen species is observed in head extracts from 1-octen-3-ol–exposed flies (16). Previous studies modeling Parkinson disease in *Drosophila* demonstrated that overexpression of the vesicular monoamine transporter promotes packaging of dopamine into vesicles, thereby lowering its cytoplasmic concentrations (19, 39). Human genetic studies have found single nucleotide polymorphisms (SNPs) within the promoter region of VMAT2, in particular gain-of-function haplotypes, are also associated with decreased risk of PD in women (40). Furthermore, in mice, reduced VMAT2 function produces dopamine-mediated toxicity and neurodegeneration in the nigrostriatal dopamine system, and restoration of VMAT2 function may be an important intervention in the treatment of PD (41–44). Therefore, impaired vesicular packaging of dopamine may represent a common pathological mechanism of PD (45).

To determine whether the 1-octen-3-ol–mediated alterations in dopamine and DOPAC levels were due to interference with vesicular storage of dopamine, we exposed HEK cells expressing human DAT alone or DAT and VMAT2 to 1-octen-3-ol. Because the inhibition of DAT by 1-octen-3-ol prevented determination of direct effects on VMAT2, airborne exposure to 1-octen-3-ol, which is most relevant to human exposure, was not possible with our assay. Therefore, to directly assess the role of VMAT2 in the dopaminergic neurotoxicity of 1-octen-3-ol in vivo, we used a genetic approach in *Drosophila*.

The mechanism of dopamine uptake and packaging in synaptic vesicles and their regulated release from these vesicles is well-conserved between *Drosophila* and mammalian systems (46). *Drosophila* DAT and VMAT share high sequence similarity with mammalian DAT and VMAT2 (19, 47). The dVMAT gene has two splice variants, dVMAT-A and -B; dVMAT-A is expressed in all dopaminergic, serotonergic, and octopaminergic neurons in *Drosophila* (19, 20). To assess the role of dVMAT in 1-octen-3-ol–
induced DA toxicity, we first exposed two heterozygous loss-of-function mutant lines of dVMAT, (2)SH0459/+ and Delta14/+, and found decreased survival times of these strains in the presence of 1-octen-3-ol (Fig. 5A). Conversely, overexpression of dVMAT in neurons was sufficient to delay 1-octen-3-ol–mediated mortality by over 3.5 d compared with control flies (Fig. 5B). This finding is similar to those of Lawal and coworkers (39), who found that overexpression of VMAT in Drosophila protected against the toxicity of a variety of pesticides linked to PD. HPLC analysis of transgenic TH-GAL4; UAS-dVMAT flies (i.e., those where VMAT was overexpressed in DA neurons) showed restoration of DA and DOPAC levels (Fig. 5C). Moreover, no detectable decreases in the number of dopaminergic neurons or detectable morphological changes were observed between the unexposed and exposed brains of TH-GAL4; UAS-dVMAT flies (i.e., those where VMAT was overexpressed in DA neurons) showed restoration of DA and DOPAC levels (Fig. 5D and E). Taken in concert, these data suggest that disruption of dopamine handling is a likely mechanism for the dopaminergic toxicity of 1-octen-3-ol.

The genes encoding the enzymes regulating the dopamine biosynthesis pathway of Drosophila are also well-conserved with those of mammals (48). The tyrosine hydroxylase (TH) gene is the rate-limiting step in the dopamine biosynthesis pathway and is responsible for the hydroxylation of tyrosine into 3,4-dihydroxy-l-phenylalanine, which is then converted into dopamine by dopa decarboxylase (49). Pharmacological inhibition of TH, catecholamine uptake, and α-adrenoceptor antagonists at 1 mM concentration results in failure of viable progeny whereas lower concentration (0.1 mM) significantly delayed the development, indicating the essential function of catecholamines in Drosophila development and viability (50). Drosophila tyrosine hydroxylase is encoded by the pale locus (51), and mutations in the pale gene result in decreased dopamine pools in adult heads, even in the heterozygous state (22). Compared with wild type, ple mutants showed increased susceptibility toward various types of stress, including the dopaminergic toxicant paraquat, indicating that dopamine functions as a key element in the stress response (22, 52). Here we also found that 1-octen-3-ol was more toxic in the ple heterozygous mutant lines. Wild-type flies survived 3–6 d longer than the heterozygous mutant strains ple− and ple+ (Fig. 5). Although these mutants failed to show any locomotory defects after 1 d of exposure to 1-octen-3-ol, there was a significant modulatory function of the mutant flies after a 7-d exposure to 1-octen-3-ol (Fig. 5B). The sensitivity of ple heterozygous mutant flies to the exposure of 1-octen-3-ol likely reflects the modulatory function of dopamine in the pathways governing survival and locomotory behavior, as seen previously with paraquat exposure (22).

In summary, these data demonstrate that 1-octen-3-ol damages the dopamine system, most likely through disruption of dopamine handling. These findings are of particular interest given recent epidemiological studies that have raised the concern of neuropsychological impairments and movement disorders in human populations exposed to moldy and water-damaged buildings (13, 14, 53, 54). Increased incidence of PD is seen in rural populations (55), where it is usually attributed to pesticide exposure. However, the prevalence of mold and mushroom in these environments may provide another plausible risk factor for the development of PD. Furthermore, 1-octen-3-ol is known to be present in human sweat (56). Being an oxidative product of linoleic acid, an essential fatty acid, its excessive endogenous production in the body may contribute to human vulnerability to developing PD. Our studies in Drosophila suggest that the common fungal VOC 1-octen-3-ol may also contribute to Parkinson disease, particularly in people who have a genetic susceptibility. Further epidemiological studies will be needed to test this hypothesis. Additionally, the toxic effects we have demonstrated for 1-octen-3-ol implies that other plant and microbial VOCs should be screened for their possible neurotoxicity.

Materials and Methods

Drosophila Strains. All stocks were cultured on Ward’s instant Drosophila medium (blue), and all experiments were performed at 25 °C. Wild-type y+, w+, a yellow-body and white-eyed strain that carries otherwise wild-type genes, was used for all of the experiments unless otherwise stated. The following mutant fly strains were used: for tyrosine hydroxylase, pale−/+ and pale−/− (22); for dVMAT mutants, (2)SH0459/+ and Delta14/+. The following mutant fly strains were used: for tyrosine hydroxylase, pale−/+ and pale−/− (22); for dVMAT mutants, (2)SH0459/+ and Delta14/+. The transgenic reporter strains used were TH-GAL4; UAS-GFP, elav-GAL4; UAS-GFP, and TH-GAL4; UAS-dVMAT. Except for TH-GAL4, elav-GAL4, and dVMAT mutant and transgenic lines, which were gifts from Janis O’Donnell (University of Alabama, Tuscaloosa, AL), Venugopal Reddy (Rutgers, The State University of New Jersey, Piscataway, NJ), and David Krantz (University of California, Los Angeles, CA), respectively, all mutant and transgenic strains were obtained from the Bloomington Stock Center.

Exposure of Flies to 1-Octen-3-ol. Adult 48-h posteclosion flies were fed on 1% (wt/vol) agar media supplemented with 5% (wt/vol) sucrose in 250-mL glass flasks. The racemic form of 1-octen-3-ol (98%) was purchased from Sigma-Aldrich (OS294-25G) in liquid form. The exposure of 1-octen-3-ol was carried out using the method published in Inamdar et al. (16) with minor modifications. The flies were exposed to vapors from undiluted aqueous 1-octen-3-ol at 0.5 ppm (vol/vol) concentration.

Mobility Assay. The mobility assays were based on negative geotaxis behavior. The assays were performed using a published protocol (16). The time taken by a single fly to cross a 5-cm distance in 8 s was recorded. Control and pale mutant flies exposed to 0.5 μL of 1-octen-3-ol were tested at 1 and 7 d of exposure. The average value was calculated from three trials in both assays with 1 min of rest between the trials.

Confocal Microscopy. Confocal microscopy experiments were done to monitor the effect of 1-octen-3-ol on the dopaminergic neurons in D. melanogaster using the transgenic lines TH-GAL4; UAS-eGFP and TH-GAL4; UAS-dVMAT. The brains from age-matched controls and 1-octen-3-ol–exposed flies were dissected and mounted on slides to examine the number and status of GFP-expressing dopaminergic neurons. The final image was obtained as an average of 2 zones of sections at 10–12 sections using a Zeiss LSM 710 confocal microscope. The number of dopaminergic neurons was scored directly under the microscope by visualizing GFP expression to acquire the quantitative data on the status of dopaminergic neurons exposed to 1-octen-3-ol.

HPLC Analysis. One hundred heads from frozen w118 flies were homogenized in 100 μL of 0.1 N perchloric acid and then centrifuged at 18,000 × g for 10 min at 4 °C. The pellets were kept for protein assay and the supernatants were filtered through 0.2-μm filters. Ten microliters of supernatant was injected onto a HPLC with electrochemical detection (Waters) for neurochemical analysis of DA and its metabolite DOPAC. The components were separated on a cation-exchange column (MD-150, 150 × 3.2 mm column, Bio-Rad) using an isocratic mobile phase (MD-TM mobile phase; ESA Biosciences) containing 2.2 mM NaCl pumped at a constant flow rate of 0.5 mL/min. The compounds were quantified by electrochemical detection using a glassy carbon working electrode, 2.0-mm diameter in situ silver reference electrode (Flow Cell, 2mm GC WE, ISAAC; Waters). The pellets in the bottom of the tubes were dried overnight in an oven at 30 °C. The dried pellets were dissolved in 100 μL of 0.5 N NaOH in a sonicating water bath for 1 h at 37 °C, and then 400 μL of H2O was added to bring the final concentration of NaOH to 0.1 N. The protein concentration for each sample was determined with a bichinchoninic acid assay reagent kit (Pierce) at 562 nm with a SpectraMax microplate reader (Molecular Devices) using BSA as a standard. The data were expressed in ng/mg of protein.

Whole-Cell [3H]DA Uptake. The human kidney transgenic cell lines HEK-DAT and HEK-DAT/VMAT2 were used. Our protocol was modified from Bernstein et al. (57). Cells were plated at 2×104 cells per well. Twenty-four hours after the experiments were performed. Cells were exposed to 1-octen-3-ol by the airborne exposure method previously described (15) for 2 h. After exposure, cells were washed with 100 μL of uptake buffer (4 mM Tris, 6.25 mM Hepes, 120 mM NaCl, 5 mM KCl, 1.2 mM CaCl2, 1.2 mM MgSO4, 5.6 mM Na-glucose, 1.7 mM ascorbic acid, 1 μM pargyline, pH 7.4). Cells were incubated with 90 μL of uptake buffer with or without 10 μM nonfenesine for 15 min. After incubation, 10 μL of uptake buffer was added and followed by 2 μL of [3H]DA uptake buffer for a final concentration of 20 nM [3H]DA and 1 μM DA was added. Cells were incubated at 37 °C for 10 min. Nonspecific uptake was determined in the presence of 10 μM nonfenesine. Uptake was terminated by aspirating...
upstage buffer and washing each well twice with 100 μL of ice-cold uptake buffer. Cells were lysed in 0.1 N NaOH and transferred to vials containing 3 mL of scintillation mixture. Radioactivity was counted using a Beckman LS6500 liquid scintillation counter.

Statistical Analysis. All of the graphs were plotted using Prism 5 (GraphPad), and data were analyzed using a one-tail Student t test or one-way/two-way ANOVA (Dunnett’s posttest or Bonferroni posttest) wherever appropriate. We have described the details of the statistical analysis in the figure legends.

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