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Structural and functional insights into alphavirus polyprotein processing and pathogenesis

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Alphaviruses, a group of positive-sense RNA viruses, are globally distributed arboviruses capable of causing rash, arthritis, encephalitis, and death in humans. The viral replication machinery consists of four nonstructural proteins (nsPs) produced as a single polyprotein. Processing of the polyprotein occurs in a highly regulated manner, with cleavage at the P2/3 junction influencing RNA template use during genome replication. Here, we report the structure of P23 in a precleavage form. The proteins form an extensive interface and nsP3 creates a ring structure that encircles nsP2. The P2/3 cleavage site is located at the base of a narrow cleft and is not readily accessible, suggesting a highly regulated cleavage. The nsP2 protease active site is over 40 Å away from the P2/3 cleavage site, supporting a trans cleavage mechanism. nsP3 contains a previously uncharacterized protein fold with a zinc-coordination site. Known mutations in nsP2 that result in formation of noncytopathic viruses or a temperature sensitive phenotype cluster at the nsP2/nsP3 interface. Structure-based mutations in nsP3 opposite the location of the nsP2 noncytopathic mutations prevent efficient cleavage of P23, affect RNA infectivity, and alter viral RNA production levels, highlighting the importance of the nsP2/nsP3 interaction in pathogenesis. A potential RNA-binding surface, spanning both nsP2 and nsP3, is proposed based on the location of inactivating sites and adaptive mutations. These results offer unexpected insights into viral protein processing and pathogenesis that may be applicable to other polyprotein-encoding viruses such as HIV, hepatitis C virus (HCV), and Dengue virus.

Alphaviruses are small, enveloped RNA viruses that infect various vertebrates, including humans, horses, birds, and rodents (1). Transmission occurs via an invertebrate vector, usually mosquitoes. In humans, typical alphavirus infection can result in rash, arthritis, encephalitis, and death. Within the past decade, infection by Chikungunya virus (CHIKV), a member of the genus alphavirus, caused massive epidemics in Africa and Southeast Asia, with positive cases reported in Europe and the United States, raising the possibility of an emerging threat (2). Additionally, CHIKV often presents with nonclassic symptoms that go undiagnosed, making global pathogenicity difficult to monitor (3).

The alphavirus genome consists of single-stranded, positive-sense RNA of ~9–11 kb with a 5’ cap structure and 3’ polyadenosine tail. The genome contains two cistrons. The first, located in the 5’ two-thirds of the genome, encodes the viral replication machinery, termed nonstructural proteins (nsPs), whereas the structural proteins that form virus particles are encoded in the second cistron. The genomic RNA is used as an mRNA for the translation of the nsPs and as a template for the synthesis of complementary negative-sense RNA. The negative-sense strand is a template for subgenomic RNA containing the second cistron and progeny, genomic RNA. Because genomic and subgenomic RNAs are made in vast excess, the negative-sense RNA strand is considered a replication intermediate.

The alphavirus replication machinery is composed of four nonstructural proteins (nsP1 to -4), which are expressed as one of two polyproteins (P123 or P1234). P1234 is expressed as a read-through of an opal termination codon at the end of nsP3. These precursor polyproteins are cleaved by a protease within nsP2 (1, 4). After translation of P1234, cleavage at the P3/4 junction occurs either in cis or trans, followed by the P1/2 junction, which occurs in cis only (4, 5). Both P123+nsP4 and nsP1+P23+nsP4 preferentially synthesize negative strand viral RNA (6, 7). The final cleavage event between P23 produces fully mature nsPs and switches RNA template for synthesis of positive-sense genomic and subgenomic RNAs. The correlation between P23 cleavage and the switch from negative- to positive-sense RNA production is poorly understood.

In mammalian cells, alphavirus infection inhibits host gene expression, causes severe cytotoxic effects (CPEs), disrupts the cellular response to IFN, and leads to cell death (8–11). Mature nsP2 contains an amino-terminal RNA helicase domain, a central protease domain that catalyzes all cleavages between the nonstructural proteins, and an inactive RNA methyltransferase-like (MT-like) domain (12). During infection, a portion of nsP2 localizes to the nucleus (13, 14) and plays a role in shutting off host-cell transcription and viral cytopathogenicity (9, 10). Two genetically distinct groups of alphaviruses termed New World and Old World have emerged because geographic isolation (8). In Old World alphaviruses, such as Sindbis virus (SINV) and Semliki Forest virus (SFV), transcriptional shutoff is nsP2-dependent (15). Even in the context of a replicon lacking the structural proteins, the nsPs of SINV and SFV alone can efficiently shut off host transcription and induce severe CPE (16).

Mutations that cause alphaviruses to establish a persistent infection with minimal or no CPE have been identified (16–20). A key feature of these mutants is their inability to inhibit host-cell transcription (10, 15). The best-characterized noncytopathic mutations map to nsP2 proline 726 (P726) within the MT-like domain (10). During infection, a portion of nsP2 localizes to the nucleus (13, 14) and plays a role in shutting off host-cell transcription and viral cytopathogenicity (9, 10). Two genetically distinct groups of alphaviruses termed New World and Old World have emerged because geographic isolation (8).

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The authors declare no conflict of interest.

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Data deposition: The atomic coordinates and structure factors have been deposited in the Protein Data Bank, www.pdb.org (PDB ID code 4GUA).

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the overall importance of nsP2 to viral replication, as well as host-cell interaction (11, 16).

The function(s) of nsP3 in alphavirus replication remains unresolved. Based on sequence conservation, nsP3 is organized into three domains: an amino-terminal macro or X domain (23), a central alphavirus-specific region, and a hypervariable-sequence carboxyl terminus. Macro domains have been shown to bind ADP-ribose (24) with certain viral macro domains also retaining ADP-ribose−1′′-phosphatase activity (25, 26). Recent work in SFV shows that residues just after the macro domain of nsP3 play a role in positioning of the P23 cleavage site (27). Additionally, the carboxyl-terminal region of nsP3 contains numerous phosphorylation sites targeted by yet unknown cellular kinases (28, 29).

Here, we present the structure of an uncleaved P23 precursor protein spanning the nsP2 protease domain through to the central, zinc-binding domain (ZBD) of nsP3 from SINV (Fig. L4). The P23 cleavage site is located in a narrow cleft formed between nsP2 and nsP3 that is inaccessible for proteolysis. Surprisingly, all of the previously reported nsP2 noncytopathic mutants lie at the interface between nsP2 and nsP3. Mutagenesis of residues in nsP3 opposite of nsP2 P726 showed inefficient cleavage of P23, delayed onset of pathogenesis and host gene shutoff, and altered viral RNA synthesis. Because P23⁎pro-zbd copurified with large amounts of RNA, we propose a potential RNA-binding surface that extends across both nsP2 and nsP3 and hypothesize that cleaving the P2/3 junction may alter the RNA-binding surface, thereby contributing to template switching by the replication machinery. Our results provide a better understanding of the mechanism of viral polyprotein processing and pathogenesis.

Results
Alphavirus P23⁎pro-zbd Structure. The SINV P23 precursor protein extending from the nsP2 protease domain to the nsP3 central domain was shown to be fully intact by SDS/PAGE and mass spectrometry despite the presence of a competent nsP2 protease active site (Fig. S1 A and B). Sequence alignment of the central domain of nsP3 shows numerous absolutely conserved cysteine residues (Fig. S2), suggesting the presence of an ion-bridging site. Various purified preparations of the middle region of nsP3 were analyzed by quantitative X-ray fluorescence for common transition metals found in metalloproteins (30, 31) and shown to have one zinc ion per molecule. Crystals of P23⁎pro-zbd diffracted to 2.85-Å resolution and the structure was determined by molecular replacement with structures of nsP2 pro and nsP3 macro domain (32) (Table S1). The resulting electron density allowed for an unambiguous trace of the entire polypeptide chain, including the previously uncharacterized nsP3 central region containing the zinc-binding site. The location of the zinc ion was confirmed by anomalous difference maps (Table S1 and see Fig. S4B).

SINV P23⁎pro-zbd shows a highly compact structure, with the four domains (protease and MT-like from nsP2; macro and ZBD from nsP3) arranged with each domain occupying a vertex of a rectangle (Fig. 1 B−D). The polypeptide chain progresses around the perimeter of the rectangle such that the nsP2 protease and nsP3 ZBD are adjacent to one another. The carboxy terminus of the P23⁎pro-zbd construct is pointing toward the protease active site (Fig. 1 B−D). A previously described SINV mutant (ts7) defective in RNA synthesis at nonpermissive temperatures contains a F312 to serine mutation in nsP3 (33). This residue is located in the hydrophobic core and surrounded by residues from the macro, ZBD, and MT-like domains (Fig. 1E). Mutation of this residue would likely destabilize the core and disrupt domain interaction, given rise to the RNA− phenotype at the nonpermissive temperature.

In a previously described nsP2pro⁎ structure, the six carboxy-terminal amino acids were disordered (12). In the P23⁎pro-zbd structure presented here, the entire polypeptide between nsP2 and nsP3 is ordered, permitting modeling of the P2/3 cleavage site. The P2/3 cleavage site is located at the base of a narrow cleft about 11−13 Å wide formed by the MT-like domain of nsP2 and the macro domain of nsP3 (Fig. 1 F and G). The P2/3 cleavage site is located at the base of a narrow cleft formed by the MT-like and macro domains. Three amino acids on either side of the scissile bond (arrow) are shown in stick representation. The view in F is similar to that in B, whereas in G is rotated 90° about a horizontal axis from F, looking directly into the cleft.

Fig. 1. Overview of the alphavirus replication machinery and P23⁎pro-zbd structure. (A) Schematic representation of the alphavirus replication machinery with emphasis on domain organization of nsP2 and nsP3. The crystallization construct consists of four domains: protease (blue) and methyltransferase-like (teal) domains of nsP2 and the macro (yellow) and zinc-binding (red) domain of nsP3, encompassing amino acids 1011−1675 of the P1234 polyprotein. Amino acid numbering provided from the amino terminus of nsP2 and nsP3 are also noted in parentheses. (B−D) Ribbon diagram of P23⁎pro-zbd colored according to A. A gray sphere denotes the position of the zinc ion. The P23 cleavage site, nsP2 protease active site, and ADP-ribose−binding site are labeled with an arrow, asterisk (*), and filled circle, respectively. (C) Ribbon diagram of P23 rotated 90° about a vertical axis from the view in B. D) Ribbon diagram of P23 rotated 180° about a vertical axis from the view in B. Box shows location of the view in E. (E) Previously described nsP3 temperature-sensitive mutant ts7 (F312S) highlighted in green with the immediately surrounding amino acids located in the macro, ZBD, and MT-like domains. nsP2 residues are shown in italics. (F and G) The P2/3 cleavage site is located at the base of a narrow cleft formed by the MT-like and macro domains. Three amino acids on either side of the scissile bond (arrow) are shown in stick representation. The view in F is similar to that in B, whereas in G is rotated 90° about a horizontal axis from F, looking directly into the cleft.
ZBD in nsP3 Is Essential for Viral Replication. After the P2/3 cleavage site, the polypeptide chain continues into the nsP3 macro domain. The ADP ribose–binding site within the macro domain is solvent-exposed and points away from the other four domains. The polypeptide exits the last helix of the macro domain adjacent to the P2/3 cleavage site and continues into a stretch of nsP3 that lacks secondary structure but forms a twist (Fig. S3). The nsP3 macro domain contains an antiparallel α-helical bundle, two parallel β-strands, and a previously unknown zinc-coordination site (Fig. S4B). The zinc is coordinated by C263, C265, C288, and C306 of nsP3. C263 and C265 are located in the loop between the last two α-helices, whereas C288 and C306 are at the carboxyl termini of the two parallel β-strands (Fig. S3C). Each of the zinc coordinating cysteines is invariant among all alphavirus sequences (Fig. S2). Structural comparison of the nsP3 ZBD against all known folds in the Protein Data Bank using PDBFold (37) and Dali servers (38) failed to produce any statistically significant hits, and no metalloprotein folds were identified. The spatial arrangement of secondary structural elements that contribute to nsP3 zinc-binding site, i.e., two cysteines in a loop between alpha helices and the other two cysteines at the end of two parallel β-strands, represents a structural scaffold for zinc-ion coordination (39).

Site-directed mutagenesis was performed to examine the functionality of the four conserved cysteine residues involved in zinc coordination in nsP3 (Fig. S4A). All four SINV mutants (C263A, C265A, C288A, and C306A) failed to produce productive infection in BHK cells, as evaluated by infectious center and plaque assays (Fig. S4C). A conserved cysteine residue in nsP3 (C247) unrelated to the zin-binding site served as a control and C247A mutant produced infectious particles to near wild-type levels. Zinc-coordination mutants failed to express the non-structural polyprotein and replicate virus, thereby supporting the structural role of the zinc ion (Fig. S4D).

The coordination site forms a distorted tetragon due to the presence of two highly conserved serine residues (S289 and S290) below the zinc atom (Fig. S2). Mutation of these serine residues did not alter viral replication levels. Furthermore, a double serine to alanine mutant (S289A+S290A) resulted in decreased viral RNA infectivity, but overall viral titers were comparable to wild type (Fig. S4C).

Importance of P23 Interface for Polyprotein Processing and Viral Pathogenesis. The interface between the macro and ZBD of nsP3 has a buried surface area of 570 Å², whereas the linker connecting the two domains is extended, making nsP3 into a ring-like structure with an inner diameter of 15–18 Å (Fig. 2A). Together, nsP2 and nsP3 share an extensive interface with 3,000 Å², which is at the end of two parallel β-strands (Fig. S5B). The two cysteines in a loop between alpha helices and the other two cysteines at the end of two parallel β-strands, represents a structural scaffold for zinc-ion coordination (39).

Several groups have identified mutations in the carboxyl-terminal portion of nsP2 that result in noncytopathic virus and persistent infection (16–20). These mutations reduce viral RNA replication levels with minimal effects on host gene expression and viral yields. All of the noncytopathic mutations map to the surface of nsP2 at the nsP3 interface (Fig. 2D). The mutations follow the encircling path made by the macro to ZBD linker. Mutagenesis of P726 in SINV to amino acids with large side chains (Phe, Tyr, Leu, Arg, or Gln) resulted in numerous steric clashes with nsP3, whereas Ser, Thr, Ala, and Val had fewer conflicts. Moreover, the P726G mutation could destabilize the α9–β11 loop, causing it to interfere with P23 interaction. Attempts to purify P23pro–ZBD with a P726S or P726L mutation resulted in poorly soluble and aggregated protein, supporting the importance of this position.

To evaluate how destabilizing P23 interface may contribute to changes in viral pathogenesis and replication, site-directed mutagenesis to the corresponding surface of SINV nsP3 in close proximity to nsP2 P726 were created (Fig. 2E). The side chains of V162 and L165 of nsP3 are ~4.6 and 3.3 Å away from nsP2 P726 side chain, respectively (Fig. 2F). V162 and L165 of nsP3 were mutated to one of the following: Ala, Asp, Glu, Phe, Arg, or Trp, as nsP2 P726 to Ser, Gly, or Leu mutants were used for comparison. RNA infectivity measured by infectious center assay on BHK cells were reduced in P726L, L165D, L165E, and L165R mutants compared with the wild-type SINV (Fig. 3A). P726L mutation in nsP2 reduced RNA infectivity to only 2% of wild-type levels, whereas the three nsP3 L165 mutants averaged an RNA infectivity at 8% of wild type. These four mutants also displayed a small size plaque phenotype in infectious center assay (Fig. S5 A–D). In addition,
reached wild-type levels of virus titers. All V162 mutants behaved similarly to wild type in both RNA infectivity and viral titer and were not pursued further. None of the nsP3 mutants displayed a noncytopathic phenotype in the TSG/puromycin N-acetyltransferase (PAC) replicon background (16), although onset of CPE in L165 to D, E, and R mutants was delayed.

Radiolabeling experiments were performed to evaluate both viral and host protein and RNA production over the course of infection and to analyze efficiency of host-cell transcriptional and translational shutoff caused by SINV nsP3 mutations. Wild-type SINV is able to halt host translation completely within 6 h postinfection, whereas P726S and P726G mutants were unable to do so even up to 12 h postinfection, as indicated by persistent actin production (Fig. S6). nsP2 P726L and the four selected nsP3 L165 mutants (Asp, Glu, Arg, Trp) were able to shut off host translation, although at a slightly delayed time postinfection compared with wild type (Fig. S6).

Tritium-labeled uridine was used to track viral and host RNA production over the course of infection. In the presence of actinomycin D, both genomic and subgenomic viral RNA production can be clearly observed in wild-type SINV starting as early as 2 h postinfection and remaining stable until 12 h (Fig. 3C). Levels of subgenomic and genomic RNA in P726G and P726S mutants are reduced compared with wild type. nsP2 P726L and the four nsP3 L165 mutants exhibit a slow initial rate of transcription, especially of genomic RNA; however, these mutants reach wild-type levels by 4 h postinfection. Thereafter, the levels of RNA replication in the L165 mutants continue to increase, and even surpass, wild-type RNA replication, especially subgenomic RNA (Fig. 3C). RNA labeling in the absence of actinomycin D showed complete inhibition of host cellular transcription by 8 h postinfection as demonstrated by decreasing 28S and 18S cellular rRNA (Fig. 3D). P726G and P726S mutants exhibited a lack of host-cell transcriptional shutoff with rRNA persisting up to 12 h postinfection. The nsP2 P726L mutant and the four nsP3 L165 mutants showed a delay in host transcription inhibition but eventually achieved complete shutoff 12 h postinfection (Fig. 3D).

Given that residues R159 and E163 of SFV nsP3 have been shown to affect P2/3 cleavage (27), we assayed the effect of the SINV nsP3 mutations on cleavage activity. nsP3 linker region mutants L165D and L165E revealed drastic inefficiency in P23 cleavage that was not seen in L165S and L165R mutants (Fig. 3F and Fig. S7). The presence of uncleaved P23 in the nsP3 L165D and L165E mutants started 6 h postinfection and persisted up to 12 h, whereas P726 mutants, as well as nsP3 L165R and nsP3 L165W, showed substantially lower amounts of uncleaved P23. Western blots probed with anti-nsP3 antibodies were used to confirm the presence of uncleaved P23 (Fig. 3F). nsP3 appears as a smear, indicative of alternate phosphorylation states and possibly other post-translational modifications (40). The nsP2 P726 mutants may also accumulate precleavage forms but at very low concentrations that do not build up over time, as in the case of the nsP3 linker mutants.

Discussion
The structure of alphavirus precursor polyprotein P23pro-zbd presented here reveals the arrangement of four domains (protease, MT-like, macro, and ZBD) in a precleavage form. Based on the structure of P23pro-zbd, several inferences can be made about the mechanism of alphavirus polyprotein cleavage. First, the carboxyl terminus of the P23pro-zbd structure is pointing toward the nsP2 protease active site about 34–45 Å away. It is possible that the large, flexible carboxyl region of nsP3 could span the distance, placing the P3/4 junction in the protease active site for cis cleavage (34). Second, the 40 Å distance between the P2/3 cleavage site and the nsP2 protease active site supports the current trans model of cleavage (5, 34, 35). Third, the inaccessibility of the P23 cleavage site itself indicates that access is tightly regulated. It is possible the activator sequence present in the extreme
amino terminus of nsP2, which becomes exposed after cleavage at the P1/2 site, may be responsible for this regulation.

During the course of alphavirus infection, a portion of nsP2 is localized to the nucleus, whereas nsP3 remains cytoplasmic (13, 14, 41). At the moment, it is unclear how nsP2 would dissociate post-cleavage from the extensive, encircling grasp of nsP3. Comparison of the domains in P23pro-zbd with the isolated structures of nsP2pro (12) and nsP3 macro domain (32) demonstrated that the protease and macro domains were highly similar with a root mean square deviation (rmsd) of <0.6 Å, whereas the MT-like domains have an rmsd of 1.3 Å for similar α carbon. A superposition of the nsP2 structures shows the discrepancy between nsP2pro and P23pro-zbd is limited to specific regions within the MT-like domain (Fig. 4A and B). α-Helices α8, α10, and α11, as well as β-strands β11, β12, and β13, between the two structures superimpose well with an rmsd of <1 Å, although the β13-α10 loop moves 5 Å because of interactions with the nsP3 linker. In addition, the nsP3 linker interacts with α9, causing the helix to rotate 8° with the carboxyl-terminal end of the helix moving 3.8 Å to form a single continuous helix with α8. This movement of the α9 helix causes the β-sheet to become more concave. The distortion of the MT-like domain in the P23pro-zbd becomes more pronounced the further the secondary structures are from α9, with α7 and β9 moving about 4 Å from their corresponding positions in nsP2pro. Perhaps movement of the MT-like domain facilitates separation of nsP2 and nsP3 post-cleavage, which is triggered by binding viral proteins or RNA to the concave side of the nsP2 MT-like β-sheet.

Previously described noncytopathic mutations at the surface of nsP2 contacting the nsP3 linker region appear by molecular modeling to destabilize P23 interaction, decreasing efficiency of the RNA replication complex during virus infection. Altering the corresponding nsP3 residue (L165), which contacts nsP2 P726, did not produce virus with the same noncytopathic phenotype. In contrast to the nsP2 P726G and P726S mutants, nsP3 L165 mutants had enhanced RNA replication levels, indicating the apparent stabilization of the replication complex. Despite the large, bulky nature of a tryptophan substitution at L165, the virus remained similar in phenotype to wild type. The L165D and L165E mutants, however, exhibited the most extreme phenotype in terms of reduced RNA infectivity and inefficient P23 cleavage. The surface of nsP2 surrounding P726 is both basic and hydrophobic (Fig. 2F), suggesting that the P23 interface may be stabilized by the introduced acidic amino acid at L165 through interaction with the surface of nsP2. This stabilization could cause the interface to become “locked in,” preventing the flexibility needed for conformational change and, thus, disallowing nsP2 access to the P2/3 cleavage site.

The buildup of uncleaved P23 is evidence that the nsP3 linker region, which encircles nsP2, may function in positioning and recognition of the P2/3 cleavage site. Given the nsP3 linker residues and nsP2 P726 are within reasonable distance to the P2/3 cleavage site (21 and 17 Å from Cβ of L165 or P726 to the cleavage site, respectively), it is plausible that mutation of the nsP3 linker region shifts local structural features, altering recognition of the cleavage site (Fig. 2F). A previously described E163R mutation in SFV nsP3 that altered P23 cleavage is completely solvent-exposed (Fig. 2E) (27). These results may indicate that the nsP3 linker residues surrounding nsP2 P726 are recognized by the nsP2 protease or another factor necessary for P2/3 cleavage.

Cleavage between P2/3 dictates template use by the alphavirus replication machinery. During purification, P23pro-zbd bound extensive amounts of bacterial RNA, which could only be removed by high concentrations of salt (Fig. S1A and C). The electrostatic potential of the P23pro-zbd structure identified a large basic surface that centers on the zinc-binding site of nsP3 and extends to nsP2 (Fig. 4C). Many zinc metalloproteins are involved in nucleic acid-binding and gene regulation (42); the localization of a basic patch in close proximity to the zinc-binding site suggests a potential role in RNA binding. Several molecules of sulfate and 2-(N-morpholino)ethane sulfonic acid (MES), used in crystallization, were bound to P23pro-zbd along the basic surface, which may indicate potential binding sites for the phosphate groups of nucleic acids (Fig. 4C–E). In fact, a molecule of MES occupies the ADP ribose–binding site in each of the three macro domains in P23pro-zbd in the asymmetric unit, as well as in the macroH2A1.1 structure (Fig. 4E) (43). The alphavirus genome contains conserved sequence elements (CSEs) that serve as promoters for production of positive- and negative-sense genomic RNA, as well as subgenomic RNA. Mutations in a 51-nt-long CSE in the nsP1-coding region of SINV disrupted the RNA structure without affecting translation (44). Adaptive mutations (E118K of nsP2 and T286I of nsP3) were shown to restore replication of the mutated CSE. T286 is 2 aa before one of the zinc-coordinating cysteine residues (C288) (Fig. 4F) and is located on the basic surface of the ZBD. T286 is 6.1 Å away from a sulfate ion that is coordinated by three highly conserved amino acids in nsP3: Y267, Y267, and R276, and R276, and surrounding residues. The close-up view corresponds to the box in E.

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of viral families, there is lack of structural information regarding the precursor forms. Currently, alphavirus P23<sup>pro-zbd</sup> and poliovirus 3CD (45) are the only structures of polypeptide precursor from a viral replication machinery. Both alphavirus P23<sup>pro-zbd</sup> and poliovirus 3CD structures contain the protease domain responsible for cleaving the precursor, which is thought to proceed by a trans mechanism. However, 3CD has no intramolecular contacts and a solvent accessible cleavage site that is regulated by intermolecular contacts, whereas P23<sup>pro-zbd</sup> has an extensive interface and an inaccessible cleavage site. The results presented here further expand our current understanding of viral polyprotein processing, replication, and cytopathogenesis, which may be applicable to other positive-sense RNA viruses.

**Materials and Methods**

Complete materials and methods are provided in *SI Materials and Methods.*

**Structure Determination.** P23<sup>pro-zbd</sup> of the SINV strain Toto1101 was expressed in Escherichia coli as a fusion with GST. The protein was purified by sequential chromatography over GST affinity, hydroxypatite, and heparin columns. Crystals of P23<sup>pro-zbd</sup> were grown by vapor diffusion against 2 M ammonium sulfate and 0.1 M MES (pH 6.5). Crystals were transferred to a cryoprotectant solution containing 2 M ammonium sulfate, 0.1 MES (pH 6.5), 3% (vol/vol) xylitol, and 5% (vol/vol) trimethylamine N-oxide and then flash-cooled in liquid nitrogen. Diffraction data were collected at the zinc K absorption edge (1.2824 Å) to 2.85 Å resolution at beam line X25 at Brookhaven National Synchrotron Light Source (Table SI). Phases were soaked with molecular replacement using the nsP2<sup>−</sup> (12) and nsP3 macro structures (32). The final model contains 3 molecules of P23<sup>pro-zbd</sup> 3 zinc ions, 31 sulfate ions, and 5 molecules of MES. Atomic coordinates have been deposited in the Protein Data Bank under PDB ID code 4UQA.

**VIROLOGICAL Studies.** Infectious center and plaque assays were performed similarly to those described in ref. 46. Protein and RNA radiolabeling were performed as described in ref. 9. Harvested protein from BHK-J cells at time points postinfection were used in Western blotting experiments, and membranes were probed with anti-nsP2, anti-nsP3, and anti-GAPDH antibodies.

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