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BMJ Open  Pneumococcal colonisation density: a new marker for disease severity in HIV-infected adults with pneumonia

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ABSTRACT

Objective: A high genomic load of Pneumococcus from blood or cerebrospinal fluid has been associated with increased mortality. We aimed to analyse whether nasopharyngeal colonisation density in HIV-infected patients with community-acquired pneumonia (CAP) is associated with markers of disease severity or poor outcome.

Methods: Quantitative lytA real-time PCR was performed on nasopharyngeal swabs in HIV-infected South African adults hospitalised for acute CAP at Chris Hani Baragwanath Hospital, Soweto, South Africa. Pneumonia aetiology was considered pneumococcal if any sputum culture or Gram stain, urinary pneumococcal C-polysaccharide-based antigen, blood culture or whole blood lytA real-time PCR revealed pneumococci.

Results: There was a moderate correlation between the mean nasopharyngeal colonisation densities and increasing CURB65 scores among all-cause patients with pneumonia (Spearman correlation coefficient r=0.15, p=0.06) or with the Pitt bacteraemia score among patients with pneumococcal bacteraemia (p=0.63). In patients with pneumococcal pneumonia, nasopharyngeal pneumococcal colonisation density was higher among non-survivors than survivors (7.7 vs 6.1 log10 copies/mL, respectively, p=0.02) and among those who had pneumococci identified from blood cultures and/or by whole blood lytA real-time PCR than those with non-bacteraemic pneumococcal pneumonia (6.6 vs 5.6 log10 copies/mL, p=0.03). Nasopharyngeal colonisation density correlated positively with the biomarkers procalcitonin (Spearman correlation coefficient r=0.37, p=0.001), proadrenomedullin (r=0.39, p=0.008) and copeptin (r=0.30, p=0.01).

Conclusions: In addition to its previously reported role as a diagnostic tool for pneumococcal pneumonia, quantitative nasopharyngeal colonisation density also correlates with mortality and prognostic biomarkers. It may also be useful as a severity marker for pneumococcal pneumonia in HIV-infected adults.

INTRODUCTION

Community-acquired pneumonia (CAP) and influenza are the second leading natural cause of death in South Africa, responsible for 6.6–7.5% of deaths between 2009 and 2011.1 Streptococcus pneumoniae is the most common aetiology of CAP in adults regardless of setting and clinical severity.2 This is also the case for HIV-infected adults and has been confirmed in various aetiologic studies from Africa.3–8 In HIV-infected adults, Pneumococcus has been estimated to be implicated in 20% of bacterial pneumonias, 40% of pneumonias with an identified organism and 70% of pneumonias with positive blood cultures.9 Pneumococcal pneumonia carries a mortality of about 10–15% with slightly higher mortality in patients with bacteraemic pneumococcal pneumonia,10–14 which has been roughly unchanged since the advent of antibiotic therapy.15 Outcomes are worse for HIV-infected patients with pneumococcal pneumonia in some studies9 but not others.9
Information on prognosis is useful to assign limited medical resources including treatment and site-of-care decisions or to intensify therapy. The prognosis for patients with pneumonia is a complex interplay between host and microbiological factors. Current prognostic scoring systems such as the CURB65 score or the Pneumonia Severity Index (PSI) are based on underlying conditions, clinical and laboratory characteristics. Interestingly, these scores have not been validated in patients with HIV infection even though HIV is a recognised risk factor for poor outcome in patients with pneumococcal pneumonia. Strikingly, these scores also do not take into account microbiological data. Novel biomarkers are increasingly used to provide prognostic information in patients with pneumonia, correlate with clinical severity scores and might provide additional information beyond clinical scores.

Since organism burden has been difficult to assess in bacterial infections prior to more widespread use of quantitative molecular techniques, relatively little information is available on its prognostic impact. A high genomic load of *Pneumococcus* from cerebrospinal fluid and blood, respectively, has been associated with poor outcome and increased mortality in Malawian children with pneumococcal meningitis and Malawian children and European adults with bacteraemic pneumococcal pneumonia. Non-invasive specimens would be preferable due to ease of sample collection and due to low diagnostic yields of blood specimens. We recently proposed quantitative nasopharyngeal (NP) colonisation density as a novel diagnostic tool for pneumococcal aetiology in HIV-infected adults with pneumonia. In this study, we aimed to analyse whether NP colonisation density is associated with clinical and biological markers of disease severity or poor outcome in the same cohort of patients with pneumonia.

**Methods**

Details of the analysed patients were previously described. In brief, we enrolled adults (age ≥18 years) who were hospitalised at Chris Hani Baragwanath Hospital, Soweto, South Africa, with acute X-ray confirmed pneumonia CAP. Active tuberculosis or current antituberculous treatment at diagnosis were exclusion criteria for enrolment, whereas patients with current or recent antibiotic therapy were not excluded. *S. pneumoniae* was identified by colony morphology, optochin susceptibility and bile solubility. Urine was tested with the immuno chromatographic BinaxNow *S. pneumoniae* test (ICT) for pneumococcal C-polysaccharide according to the manufacturer’s instructions. Quantitative lytA realtime (rt) PCR was performed on NP swab as described. For whole blood testing, nucleic acids were extracted from 200 µL of clinical samples using the QIAamp DNA Blood Mini Kit (Qiagen, the Netherlands) and eluted in 100 µL of elution buffer. 5 µL of extracted DNA was subjected to a triplex rtPCR for *Pneumococcus* (lytA), *Haemophilus influenzae* type b, and *Staphylococcus aureus* using the IQ Multiplex Powermix (Bio-Rad, USA). Owing to the lack of a diagnostic gold standard, pneumococcal pneumonia (SP-CAP) was considered present if a composite diagnostic standard was positive in patients with CAP, that is, if pneumococci were detected by either blood culture, urine ICT, good quality (ie, >25 neutrophils and <10 epithelial cells per high-power field) sputum Gram stain or culture, or whole blood lytA rtPCR. In an expanded composite diagnostic (eSP-CAP) we included a quantitative NP density >8000 copies/µL as an additional criterion for pneumococcal pneumonia. Antimicrobial activity in urine as evidence of previous antimicrobial use was measured as previously described.

HIV counselling and testing were offered to all patients with unknown or negative HIV serostatus. Testing was performed using the Architect HIV Ag/Ab Combo assay (Abbott) and, if positive, was confirmed by Elecsys HIV combi assay (Elecsys 2010 analyzer, Roche). The main study population included only patients who were already known to be HIV-infected or newly diagnosed with HIV infection.

The CURB65 score was calculated as according to Lim et al. For patients with bacteraemic pneumonia, that is, positive blood culture or whole blood lytA rtPCR, we calculated the Pitt bacteriaemia score. Infection-related and prognosis-related biomarkers were measured in batch with a highly sensitive research assay for procalcitonin (BRAHMS PCT LIA sensitive) and commercial assays for mid-regional proadrenomedullin (BRAHMS MR-proADM KRYPTOR), mid-regional proatrial natriuretic peptide (BRAHMS MR-proANP KRYPTOR) and copeptin (BRAHMS Copeptin KRYPTOR, all Thermo Scientific Biomarkers, Hennigsdorf, Germany) after storage at −70°C. C reactive protein (CRP) was measured immediately on site by the routine diagnostics laboratory (CRP immunoturbidometry, 717 automated analyzer; Boehringer Mannheim/Hitachi, Mannheim, Germany) if requested by the treating physicians.

Continuous variables were compared with two-sided pooled t tests or a Mann-Whitney-Wilcoxon test, as appropriate, and proportions calculated with Pearson’s χ² test or Fisher’s exact test, as appropriate. For relationship between continuous variables, we used Spearman correlation coefficients. One-way analysis of variance (ANOVA) and Kruskal-Wallis tests were applied for comparisons of continuous dependent variables between more than two categorical independent variables, as appropriate. Univariate and multivariate analyses were performed to identify predictors of hospital mortality. Variables significant in the univariate analysis and those of special interest were included in the multivariate analysis. The area under the receiver operating characteristic curve (AUC-ROC) was calculated as indicator for diagnostic accuracy to predict hospital mortality.

Values ≤0.05 were considered significant.

All patients and controls provided written informed consent.
RESULTS

Of 514 patients with acute clinical pneumonia, 370 had X-ray confirmed pneumonia (CAP). Analyses were restricted to those 280 patients with CAP in whom HIV-infection was present or newly diagnosed. Based on the composite diagnostic criteria, SP-CAP was diagnosed in 99 (35.4%) of 280 patients; 75 had either a positive urine ICT or blood culture, or good quality (ie, >25 neutrophils and <10 epithelial cells per high-power field) sputum Gram stain or culture with pneumococcus; 58 patients had a positive whole blood lytA rrtPCR. A lytA rrtPCR from NP >8000 copies/mL was present in 126 patients, resulting in a diagnosis of pneumococcal CAP based on the expanded composite diagnostic in 150 (53.6%) of 280 patients. The in-hospital case death rate was not different between those with (11.7%) and those without (14.1%; p=0.53) pneumococcal aetiology.

Mean pneumococcal colonisation density was not significantly different between patients with and those without antimicrobial activity in urine (3.36 (95% CI 2.82 to 3.89) copies/mL vs 3.95 (95% CI 3.30 to 4.60); p=0.16). There was also no difference in in-hospital mortality between those with and those without antimicrobial activity in urine (13.2% vs 11.5%; p=0.69).

Correlation between colonisation density and clinical scores

As a prognostic marker for mortality, we correlated NP colonisation with the CURB65 score. Colonisation density measured by lytA rrtPCR did not significantly correlate with increasing CURB65 classes either among cases with CAP (Spearman correlation coefficient r=0.15, p=0.06) or among cases with SP-CAP (Spearman correlation coefficient r=0.14, p=0.19). The mean colonisation density of cases with bacteraemic SP-CAP did not vary significantly for different Pitt bacteraemia scores (0: 6.5 log_{10} copies/mL; 1: 6.8 log_{10} copies/mL; 2: 7.0 log_{10} copies/mL; 3: 4.8 log_{10} copies/mL; p=0.63). Hospital mortality did not vary significantly between CURB65 scores (0:13.1%; 1: 11.1%; 2: 11.3%; 3: 25.0%; 4: 0%; p=0.56) or—among those with bacteraemic pneumococcal pneumonia—between Pitt bacteraemia score points (0: 20.7%; 1: 10.7%; 2: 16.7%; 3: 0%).

Correlation between colonisation density and bacteraemia

Presence of pneumococci in blood (by culture or whole blood lytA rrtPCR) among cases with SP-CAP was associated with a higher pneumococcal NP colonisation density than if SP-CAP was non-bacteraemic (6.6 vs 5.6 log_{10} copies/mL, p=0.03; figure 1). Similarly, growth of S. pneumoniae from blood cultures was associated with higher NP colonisation density among patients with SP-CAP than those without (7.1 vs 6.1 log_{10} copies/mL, p=0.04).

Correlation between colonisation density and biomarkers

The levels of the biomarkers procalcitonin, proadrenomedullin and CRP, but not of copeptin and ProANP were significantly higher for patients with SP-CAP than for patients with CAP without pneumococcal aetiology (table 1).

There was a positive correlation between increasing NP colonisation density among those colonised with *Pneumococcus* and increasing procalcitonin values in cases with CAP (Spearman correlation coefficient r=0.37, p<0.0001) and in cases with SP-CAP (r=0.37, p=0.001; figure 2A). Similarly, there was a significant positive correlation between increasing NP colonisation density among those colonised with *Pneumococcus* and increasing proadrenomedullin values in cases with CAP (r=0.34, p<0.0001) and in cases with SP-CAP (r=0.40, p=0.008; figure 2B). A similar positive correlation was observed between increasing copeptin and increasing NP colonisation density among those colonised with *Pneumococcus*.

Figure 1  Association of nasopharyngeal colonisation density with bacteraemia and death. Nasopharyngeal colonisation density according to bacteraemia (either positive blood cultures or positive lytA real-time (rt) PCR from whole blood) and survival in HIV-infected patients with pneumococcal pneumonia. +/° represent the mean; length of the box represents the IQR between the 25th and 75th centiles; horizontal line in the box represents median; whiskers represent minimum and maximum values.
coloniisation density among those colonised with *Pneumococcus* and in patients with CAP (r=0.18, p=0.03) and in patients with SP-CAP (r=0.30, p=0.01; figure 2C).

There was a trend for a positive correlation between increasing NP colonisation density among those colonised with *Pneumococcus* and increasing ProANP values in patients with CAP (r=0.17, p=0.047), but less so in patients with SP-CAP (r=0.12, p=0.30; figure 2D). In contrast, there was no significant correlation between increasing NP colonisation density and CRP values for patients with CAP (r=0.13, p=0.51) or for patients with SP-CAP (r=0.35, p=0.17; figure 2E).

**Correlation between colonisation density and in-hospital mortality**

Among patients with SP-CAP who had pneumococcal colonisation in the nasopharynx, mean NP colonisation density was significantly higher in those who died (n=8; 7.72 log₁₀ copies/mL) than those who survived (n=76; 6.11 log₁₀ copies/mL; p=0.02). All non-survivors had very high NP colonisation densities, which were at least 6.4 log₁₀ copies/mL and thus more than 2.5 log₁₀ copies/mL higher than the cut-off of 3.9 log₁₀ copies/mL proposed to diagnose pneumococcal aetiology (figure 1).

In univariate analysis the levels of copeptin, ProANP, proadrenomedullin and procalcitonin and the NP pneumococcal colonisation density were significantly associated with hospital mortality in patients with CAP (table 2). In multivariable logistic regression, proadrenomedullin (aOR 1.196, 95% CI 1.011 to 1.414; p=0.04) and quantitative *lytA* rtPCR from NP (aOR 1.870; 95% CI 1.047 to 3.342; p=0.03) were independent predictors of hospital mortality (table 2).

In patients with SP-CAP, receipt of highly active antiretroviral therapy (HAART) and increasing levels of the biomarkers copeptin, ProANP, proadrenomedullin and increasing NP pneumococcal colonisation density were associated with hospital mortality (table 2). In a multivariable logistic regression model, there were trends for increasing age (aOR 1.09; 95% CI 0.99 to 1.20; p=0.10) and quntitative *lytA* rtPCR from NP (aOR 2.10; 95% CI 0.95 to 4.65; p=0.07) as predictors of mortality (table 2).

The prognostic accuracy as determined by the ROC-AUC curves showed the best prediction of hospital mortality from all-cause pneumonia for the prognostic biomarkers proadrenomedullin, ProANP and copeptin, with lower AUC observed for quantitative *lytA* rtPCR from NP (see online supplementary table S3, figure 3A). In patients with SP-CAP, proadrenomedullin and quantitative *lytA* rtPCR from NP had the highest AUC (0.77 and 0.74, respectively, figure 3B).

**DISCUSSION**

This study shows, for the first time, that NP colonisation density as measured by quantitative *lytA* rtPCR was predictive of bacteraemia and mortality in HIV-infected South African adults with CAP. Colonisation density correlated moderately with prognostic biomarkers indicative of poor outcome.

Despite the theoretical limitation being that molecular diagnostics cannot distinguish between dead and alive bacteria, we previously reported a good correlation between quantitative *lytA* rtPCR and quantitative NP cultures. A pneumococcal colonisation density cut-off >8000 copies/mL for HIV-infected adults with CAP was a highly sensitive and specific diagnostic tool for pneumococcal aetiology. This supported the pathophysiological

<table>
<thead>
<tr>
<th>Table 1 Biomarkers in HIV-infected patients with community-acquired pneumonia <em>Based on composite diagnostic standard</em></th>
<th>Based on expanded composite diagnostic standard†</th>
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</thead>
<tbody>
<tr>
<td><strong>Pneumococcal aetiology</strong></td>
<td><strong>Non-pneumococcal aetiology</strong></td>
</tr>
<tr>
<td><strong>Procalcitonin (ng/mL)</strong> 18.8 (n=84)</td>
<td>6.93 (n=153)</td>
</tr>
<tr>
<td><strong>Proadrenomedullin (MR-proADM; nmol/L)</strong> 2.9 (n=80)</td>
<td>1.9 (n=156)</td>
</tr>
<tr>
<td><strong>Proatrial natriuretic peptide (MR-proANP; pmol/L)</strong> 119.4 (n=81)</td>
<td>120.9 (n=156)</td>
</tr>
<tr>
<td><strong>Copeptin (pmol/L)</strong> 73.0 (n=82)</td>
<td>46.2 (n=155)</td>
</tr>
<tr>
<td><strong>CRP (mg/L)</strong> 282.9 (n=20)</td>
<td>134.4 (n=27)</td>
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</table>

Since C reactive protein (CRP) values were available only when requested by the treating physicians, we compared patients with available values for CRP to patients without available CRP values. There was no difference in mean age, in mean nasopharyngeal colonisation density and no difference in pneumococcal diagnosis. However, patients with an available CRP value had a significantly higher (30.6%) in-hospital mortality compared to patients without a CRP value (15.3%) (p=0.02).

Pneumococcal and non-pneumococcal aetiology were defined according to composite diagnostic standard (pneumococci were detected by either blood culture, urine immunochromatographic BinaxNow Streptococcus pneumoniae test, good quality (ie, >25 neutrophils and <10 epithelial cells per high-power field) sputum Gram stain or culture, or whole blood *lytA* rtPCR). (rt, real-time).

†Pneumococcal and non-pneumococcal aetiology were defined according to composite diagnostic standard (pneumococci were detected by either blood culture, urine immunochromatographic BinaxNow Streptococcus pneumoniae test, good quality (ie, >25 neutrophils and <10 epithelial cells per high-power field) sputum Gram stain or culture, or whole blood *lytA* rtPCR or *lytA* rtPCR from nasal swab >8000 copies/mL.)
concept of a critical pneumococcal colonisation density in the nasopharynx, at which the risk of pneumococcal microaspiration and subsequent lower respiratory tract infection increase. Owing to the lack of lung specimens or bronchoalveolar lavage samples representing the direct site of infection, we instead chose to correlate NP colonisation density and biomarkers in pneumococcal pneumonia. Correlation between nasopharyngeal colonisation density and biomarkers among HIV-infected cases with SP-CAP who are colonised with pneumococcus. Pneumococcal aetiology (SP-CAP) in patients with CAP was defined as pneumococcal detection from either blood culture, urine immunochromatographic BinaxNow Streptococcus pneumoniae test, good quality (ie, >25 neutrophils and <10 epithelial cells per high-power field) sputum Gram stain or culture, or whole blood lytA real-time (rt) PCR. Counts are depicted after logarithmic transformation.

Figure 2 Correlation between nasopharyngeal colonisation density and biomarkers in pneumococcal pneumonia. (A) Procalcitonin. (B) MR-proADM. (C) Copeptin. (D) MR-proANP. (E) C reactive protein. Correlation between nasopharyngeal colonisation density and biomarkers among HIV-infected cases with SP-CAP who are colonised with pneumococcus. Pneumococcal aetiology (SP-CAP) in patients with CAP was defined as pneumococcal detection from either blood culture, urine immunochromatographic BinaxNow Streptococcus pneumoniae test, good quality (ie, >25 neutrophils and <10 epithelial cells per high-power field) sputum Gram stain or culture, or whole blood lytA real-time (rt) PCR. Counts are depicted after logarithmic transformation.
### Table 2: Predictors of in-hospital mortality in HIV-infected adults with CAP and SP-CAP

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<tr>
<th></th>
<th>CAP Univariate analysis</th>
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<th>SP-CAP Univariate analysis</th>
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<th>SP-CAP Multivariable analysis</th>
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<tbody>
<tr>
<td></td>
<td>OR</td>
<td>95% Confidence limit</td>
<td>p Value</td>
<td>OR</td>
<td>95% Confidence limit</td>
<td>p Value</td>
<td>OR</td>
<td>95% Confidence limit</td>
<td>p Value</td>
</tr>
<tr>
<td><strong>Pneumococcal aetiology (vs not)</strong></td>
<td>0.806</td>
<td>0.378–1.719</td>
<td>0.5766</td>
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<tr>
<td><strong>Bacteraemic pneumococcal pneumonia (vs not)</strong></td>
<td>1.289</td>
<td>0.585–2.841</td>
<td>0.5285</td>
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<td><strong>Male (vs female)</strong></td>
<td>1.083</td>
<td>0.527–2.225</td>
<td>0.8292</td>
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<tr>
<td><strong>Age</strong></td>
<td>1.028</td>
<td>0.992–1.065</td>
<td>0.1307</td>
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<tr>
<td><strong>Cotrimoxazole prophylaxis (vs not)</strong></td>
<td>0.211</td>
<td>0.028–1.603</td>
<td>0.1327</td>
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<tr>
<td><strong>CD4 count</strong></td>
<td>0.999</td>
<td>0.996–1.002</td>
<td>0.4752</td>
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<tr>
<td><strong>HAART (vs not)</strong></td>
<td>0.838</td>
<td>0.238–2.948</td>
<td>0.783</td>
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<td><strong>CURB65</strong></td>
<td>1.137</td>
<td>0.782–1.654</td>
<td>0.5007</td>
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<tr>
<td><strong>Copeptin</strong></td>
<td>1.005</td>
<td>1.002–1.006</td>
<td>0.0013</td>
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<tr>
<td><strong>MR-proANP</strong></td>
<td>1.005</td>
<td>1.003–1.008</td>
<td>&lt;0.0001</td>
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<tr>
<td><strong>PCT</strong></td>
<td>1.013</td>
<td>0.996–1.030</td>
<td>0.1226</td>
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<tr>
<td><strong>MR-proADM</strong></td>
<td>1.260</td>
<td>1.123–1.412</td>
<td>&lt;0.0001</td>
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<tr>
<td><strong>Log$_{10}$ lytA rtPCR NP</strong></td>
<td>1.284</td>
<td>0.985–1.672</td>
<td>0.0642</td>
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**Pneumococcal and non-pneumococcal aetiology were defined according to composite diagnostic standard (pneumococci were detected by either blood culture, urine immunochromatographic BinaxNow Streptococcus pneumoniae test, good quality (ie, >25 neutrophils and <10 epithelial cells per high-power field) sputum Gram stain or culture, or whole blood lytA real-time (rt) PCR).**

**ORs are reported as increase per single unit of the respective risk factor, for example, per year (age), per point (CURB65 score), per cell/µL (CD4 count).**

**CAP, community-acquired pneumonia; SP-CAP, pneumococcal pneumonia.**
colonisation density with prognosis. Despite the biological plausibility that clinical severity and risk of poor outcome correlate with a higher organism density at the site of infection, there are surprisingly few data to support this hypothesis in patients with pneumonia. Major reasons might be the difficulty of measuring the organism burden prior to the wider application of quantitative molecular methods and obtaining the appropriate specimen for this measurement. Studies in the first half of the 20th century using quantitative blood cultures reported a dose–effect relationship between organism burden and mortality.26 27 As a surrogate marker for bacterial load in children with bacteraemic SP-CAP in Spain, a shorter time to positivity of blood cultures correlated with risk of hospitalisation, admission to the intensive care unit (ICU), meningitis or other non-pneumonic sources of bacteraemia and with a higher number of positive blood cultures.28 In children with pneumococcal meningitis in Malawi, higher DNA loads as measured with a lytA rtPCR from cerebrospinal fluid and blood were found in more non-survivors than survivors (8.2 vs 7.5 log₁₀ copies/mL, p=0.03; and 3.8 vs 3.2 log₁₀ copies/mL, p=0.02; respectively). All children with SP-CAP survived and had lower DNA loads (0.42 log₁₀ copies/mL) than meningitis survivors and non-survivors.20 Genomic loads were higher for HIV-infected children (n=59) than HIV-uninfected children (n=39) (p=0.04 for CSF; p=0.6 for blood). In addition, blood bacterial DNA load correlated with plasma cytokines such as tumor necrosis factor-α, interleukin (IL)-1β, IL-6 and IL-10.20 In a study of 45 Dutch adults with CAP, quantitative pneumococcal load measured with lytA PCR from whole blood was higher in patients with bacteremia and in those who fulfilled criteria of the systemic inflammatory response syndrome. It correlated positively with markers for severity of disease including length of stay, CRP levels and mental status changes.21 In a Spanish study of 93 adults with pneumococcal CAP whole blood lytA copies independently predicted septic shock and receipt of mechanical ventilation.13 High lytA copy numbers (≥10⁵/mL) were risk factors for bacteraemia (OR=6.3), septic shock (OR=8.0), acute kidney injury (OR=7.0), mechanical ventilation (OR=10.5), acute respiratory distress syndrome (OR=14.8), rapid radiological spread (OR=22.8) and mortality (OR=5.4).13 A recent study showed that HIV and influenza infections were associated with higher lytA load in blood among South African patients with acute lower respiratory tract infections and demonstrated 3.6-fold increased mortality if pneumococcal load was ≥10⁵/mL.29 Plural fluid genomic pneumococcal load measured with pby rtPCR, which is a less specific target than lytA,30 in children with SP-CAP correlated with positive cultures, higher biomarkers such as CRP and neutrophils and with a longer length of hospital stay.31 In another study, high blood lytA copy numbers in children ≤5 years of age with CAP were predictive of a parapneumonic effusion. Children with empyema had higher values than those with simple parapneumonic effusion. Serotype 19A was associated with high bacterial loads as well as parapneumonic effusion or empyema.32 A limitation of applying quantitative molecular methods to invasive specimens in these studies is that only the relatively small subset of pneumococcal disease with documented invasiveness can be evaluated: approximately 10–25% of SP-CAP in adults33 34 and < 10% in children35 36 are bacteraemic. Therefore, application of quantitative molecular methods in NP swabs not only improves ease of specimen collection but also markedly increases the number of patients who could be assessed in this way. In this regard NP swabs are also superior to sputum, which frequently cannot be obtained in sufficient quantity and quality.37 In adults from New Zealand
Correlation with a variety of severity markers with the presence of bacteraemia and with mortality all support NP colonisation density as a prognostic tool in HIV-infected adults with SP-CAP. Persistence or increase of bacterial loads over time by serial monitoring might indicate insufficient treatment, antibiotic resistance or insufficient focus control, while documented decreases of the bacterial load could allow discontinuation of antibiotic therapy. Waterer and Rello suggested that a high organism burden could indicate a risk for antibiotic-induced bacterial lysis, cytokine release and an unfavourable proinflammatory state with subsequent haemodynamic instability, which might benefit from immunomodulation. Therefore, a high colonisation density might be an indication for combination treatment with macrolides or other antibiotics that down-regulate pneumolysin production, or for alternative anti-inflammatory treatments.

A limitation of our study includes the lack of data, particularly arterial blood gas analysis, to calculate the PSI. Owing to the setting, very few patients were admitted to ICU, which unfortunately makes this a not useful outcome measure. As there were few HIV-uninfected patients in our study, our results should be confirmed in a larger study of HIV-uninfected adults even though similar trends were found regardless of HIV status in our study (data not shown).

CONCLUSIONS

As previously reported, the quantitative lytA rtPCR from NP swabs is a very promising tool to diagnose pneumococcal pneumonia. In addition, this study shows that the same assay also conveys some prognostic information as it correlated with bacteraemia, survival and prognostic biomarkers. How exactly it could be implemented and how it might change management, such as site of care, antibiotic choices and duration, needs to be determined.

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