Establishment of Patient-derived Xenografts in Mice.

Dongkyoo Park, Emory University
Dongsheng Wang, Emory University
Guo Chen, Emory University
Xingming Deng, Emory University

Journal Title: Bio-Protocol
Volume: Volume 6, Number 22
Publisher: Bio-protocol LLC. | 2016-11-20, Pages e2008-e2008
Type of Work: Article | Post-print: After Peer Review
Publisher DOI: 10.21769/BioProtoc.2008
Permanent URL: https://pid.emory.edu/ark:/25593/rwx1g

Final published version: http://dx.doi.org/10.21769/BioProtoc.2008

Accessed July 9, 2019 2:03 PM EDT
Establishment of Patient-derived Xenografts in Mice

Dongkyoo Park1, Dongsheng Wang2, Guo Chen1, and Xingming Deng1,*

1Division of Cancer Biology, Department of Radiation Oncology, Emory University School of Medicine and Winship Cancer Institute of Emory University, Atlanta, GA, USA
2Department of Hematology and Medical Oncology, Emory University School of Medicine and Winship Cancer Institute of Emory University, Atlanta, GA, USA

Abstract

Patient-derived xenograft (PDX) models for cancer research have recently attracted considerable attention in both the academy and industry (Hidalgo et al., 2014; Wilding and Bodmer, 2014). PDX models have been developed from different tumor types including lung cancer to improve the drug development process. These models are used for pre-clinical drug evaluation and can be used for the predictive results of clinical outcomes because they conserve original tumor characteristics such as heterogeneity, complexity and molecular diversity (Kopetz et al., 2012). Additionally, PDX model provides the potential tool for the personalized drug therapy. In this protocol, we present methods for the establishment of PDX in mice using primary tumor tissues from patients with small cell lung cancer (SCLC).

Materials and Reagents

1. Sterile alcohol prep pads (Covidien, catalog number: 6818)
2. Petri dishes (Corning, Falcon®, catalog number: 353003)
3. Tissue adhesive (3M, catalog number: 1469SB)
4. 6 weeks old SCID mice (Charles River Laboratories International, catalog number: 236) or athymic nude mice (Envigo, catalog number: 069)
5. Ketamine hydrochloride/xylazine hydrochloride solution (Sigma-Aldrich, catalog number: K113)
6. Phosphate-buffered saline (PBS) (Mediatech, catalog number: 21-040-CV)
7. 70% ethanol (Decon Labs, catalog number: 2401)

*For correspondence: xdeng4@emory.edu.
1Ensure that all procedures should be undertaken in biological safety cabinet.
2Ensure that any necrotic tumor tissue is not used for transplant (Figure 2).
3Ensure that only early-passage PDXs (less than the 5th passage) can be used for the studies.
Equipment

1. Stainless steel sterile scalpels (Integra LifeSciences, Miltex®, catalog number: 4-423)
2. Operating scissors (Sklar Surgical Instruments, catalog number: 13-1045)
3. Micro dissecting forceps (Roboz Surgical Instrument, catalog number: RS-5135)
4. Biological safety cabinet (Labconco, catalog number: 3460001)
5. CO₂ chamber in animal facility
6. Heating pad (Sunbeam Products, catalog number: 000765-500-000)

Procedure

1. Collect pieces of primary tumors by surgery or biopsy procedures (F0).
2. Anesthetize the SCID mice using ketamine hydrochloride/xylazine hydrochloride solution.
3. Remove hair from the dorsal region.
4. Cut the tumor (F0) into pieces.
5. Make a shallow incision in the dorsal region of SCID mouse using a scissor and then implant subcutaneously two pieces of primary tumor into the interscapular fat pad of SCID mouse within 3 h after collecting primary tumor tissues from a patient. These mice in the engraftment phase are called F1 mice (SCID mice are used for F1 mice due to higher engraftment rates than those of athymic nude mice).
6. Move the F1 mice to a warm area on heating pad and monitor them during recovery.
7. Return F1 mice to the routine mouse sterile housing.
8. When the tumor grows and reaches around 1.5 cm (4–10 weeks), prepare the additional transplant.
9. Place the F1 mouse in the CO2 chamber for 5 min to euthanize it.
10. Move the euthanized mouse into biological safety cabinet and clean the whole body of mouse using sterile alcohol prep pads.
11. Cut off skin around the tumor area.
12. Resect the tumor and transfer the tumor to a sterile Petri dish containing PBS (Video 1).
13. Cut the tumor into even size piece (i.e., 2 mm³) (Video 1).
14. Anesthetize the athymic nude mice like described in step 2.
15. Implant a piece of tumor into each experimental athymic nude mouse like described in step 5. Apply tissue adhesive to cut surfaces of the skin. Mice in this expansion phase are called F2 mice. After 2–4 weeks, the tumors could be visible as in Figure 1 and Video 2.

16. Move the F2 mice to a warm area and monitor them like described step 5.

17. Return the F2 mice to the routine mouse sterile housing.

18. When the tumor volume reaches 100–150 mm$^3$, mice can be used for downstream application.

Acknowledgments

We thank Dr. Taofeek K. Owonikoko, Assistant Professor and Guojing Zhang, Research Specialist at Emory University for providing primary lung tumor tissues. This work is supported by NCI, National Institutes of Health grants 1R01CA193828, 2R01CA136534 and 1R01CA200905-01A1.

References


Figure 1. Establishment of PDX model
A. Photographs of representative small cell lung cancer (SCLC) PDX in mice (F3 mice). B. H&E histology of tumor tissues from SCLC PDX in mice. TKO stands for Dr. Taofeek K. Owonikoko who provided primary lung tumor tissues.
Figure 2.
Photographs of representative non-necrotic tissue and necrotic tissue
Video 1.
The cutting of tumor into small even piece for engraftment
Video 2.
The tumor implantation in mice