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Journal Title: Cancer Prevention Research
Volume: Volume 8, Number 11
Publisher: American Association for Cancer Research | 2015-11-01, Pages 1061-1068
Type of Work: Article | Post-print: After Peer Review
Publisher DOI: 10.1158/1940-6207.CAPR-15-0212
Permanent URL: https://pid.emory.edu/ark:/25593/rthdm

Final published version: http://dx.doi.org/10.1158/1940-6207.CAPR-15-0212

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Accessed September 16, 2017 2:03 PM EDT
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Abstract

Aspirin has been shown to protect against colorectal neoplasms, however the optimal chemopreventive dose and underlying mechanisms are unclear. We aimed to study the relationship between prostanoid metabolites and aspirin’s effect on adenoma occurrence. We used data from the Aspirin/Folate Polyp Prevention Study, in which 1,121 participants with a recent adenoma were randomized to placebo or two doses of aspirin (81 or 325 mg/d) to be taken until the next surveillance colonoscopy, anticipated about 3 years later. Urinary metabolites of prostanoids (PGE-M, PGI-M, and dTxB2) were measured using LC/MS or GC/NICI-MS in 876 participants near the end of treatment follow-up. Poisson regression with a robust error variance was used to calculate relative risks and 95% confidence intervals. PGE-M, PGI-M, and dTxB2 levels were 28%, 37%, and 60% proportionately lower, respectively, in individuals who took 325 mg of aspirin compared to individuals who took placebo (all P<0.001). Similarly, among individuals...
who took 81 mg of aspirin, PGE-M, PGI-M, and dTxB₂ were, respectively, 18%, 30%, and 57% proportionally lower compared to placebo (all P<0.005). None of the metabolites or their ratios were statistically significantly associated with the risk of adenoma occurrence. The effect of aspirin in reducing adenoma risk was independent of prostanoid levels. Aspirin use is associated with lower levels of urinary prostanoid metabolites. However, our findings do not support the hypothesis that these metabolites are associated with adenoma occurrence, suggesting that COX-dependent mechanisms may not completely explain the chemopreventive effect of aspirin on colorectal neoplasms.

**Keywords**

aspirin; prostaglandins; thromboxanes; colorectal neoplasms; randomized controlled trial

**INTRODUCTION**

There is strong evidence from observational studies and randomized clinical trials that aspirin and other nonsteroidal anti-inflammatory drugs (NSAIDs) are protective against colorectal neoplasms (1–5), however the dose-response relationship and underlying antineoplastic mechanisms are not clear. Pooled results from four clinical trials with 2,967 participants randomly assigned to doses of aspirin from 81 to 325 mg daily showed a borderline statistically significant 15% decrease in any adenoma occurrence, and a statistically significant 29% decrease in advanced adenoma occurrence with higher-dose aspirin supplementation (≥300 mg/d)(6). Interestingly, in a combined analysis of the two trials (2, 5) [including ours (2)] that tested higher (300 or 325 mg/d) and lower (≤160 mg/d) doses of aspirin versus placebo, lower-dose aspirin showed statistically significantly greater risk reduction for all colorectal adenomas than higher-dose aspirin (6).

Prostanoids, important mediators of human physiology, are a subclass of eicosanoids consisting of the prostaglandins (PGs), the prostacyclins, and the thromboxanes. They are generated from the precursor arachidonic acid by cyclooxygenase (COX) enzymes: COX-1, constitutively expressed in the luminal gastrointestinal tract and responsible for the production of cytoprotective PGs; and COX-2, inducible by a variety of factors such as growth factors and mitogens, and a central element of the inflammatory response (7). The increased activity of COX-2 plays an important role in colorectal carcinogenesis (8, 9); the involvement of COX-1 has been less investigated, but also appears to be involved (10). Of the PGs, prostaglandin E₂ (PGE₂) is likely to be the primary mediator of COX pro-carcinogenic effects (8).

Thromboxane A₂ (TXA₂) and prostacyclin (PGI₂) are other prostanoids, both well known to regulate cardiovascular homeostasis: TXA₂ stimulates vasoconstriction and platelet aggregation, whereas prostacyclin PGI₂ promotes vasodilatation and inhibits platelet aggregation (8). In addition to their cardiovascular effects, both PGI₂ and TXA₂ appear to affect carcinogenesis. Thromboxane A₂ activation of platelets leads to the release of mediators of carcinogenesis, and TXA₂ itself seems to act as a pro-carcinogenic factor (10–13). On the other hand, PGI₂ seems anticarcinogenic (14): lower levels are seen in
neoplastic tissue (15–17), *in vitro* and *in vivo* studies suggest interference with carcinogenesis (12, 18), and studies of the prostacyclin synthase gene point to anti-neoplastic effects as well (19, 20).

Low doses of aspirin are sufficient to reduce PGE\(_2\) levels in the colorectal mucosa, with no additional increase in inhibition with doses above 81 mg per day (21–25). There have been no studies of the effects of aspirin on PGI\(_2\) or TXA\(_2\) in the large bowel mucosa, but higher doses are generally required to substantially modulate systemic prostacyclin production (26–29), while lower doses are effective in inhibiting platelet TXA\(_2\), and increasing the ratio of PGI\(_2\) to TXA\(_2\) (30–32). If prostacyclin has anti-neoplastic properties, aspirin doses above 81 mg would then lose preventive potency from the inhibition of this potentially anti-neoplastic prostanoid.

To understand the role of PGE\(_2\), PGI\(_2\), and TXA\(_2\), in colorectal carcinogenesis, we conducted secondary analyses in the Aspirin/Folate Polyp Prevention Study (AFPPS) (2, 33), a clinical trial of aspirin and/or folic acid for the prevention of occurrence of colorectal adenomas. Our aims were to assess the effect of two doses of daily aspirin intake on urinary prostanoid metabolites; to investigate whether urinary prostanoid metabolites are associated with risk for recurrent adenoma diagnosed during treatment; and to examine whether a hypothesized aspirin dose-dependent PGE\(_2\) to PGI\(_2\) trade-off plays a role in colorectal adenoma development.

**MATERIALS AND METHODS**

**Design**

These data were collected as part of the AFPPS, a randomized, double-blind, placebo-controlled, three-by-two factorial trial testing whether oral aspirin (81 or 325 mg daily) or folic acid (1 mg daily) reduces the risk of new colorectal adenomas (clinicaltrials.gov identifier: NCT00272324) (2, 33).

**Recruitment, randomization, treatment, and follow-up**

Details of subject eligibility, recruitment, randomization, treatment, and follow-up, and study outcomes have been previously described (2, 33). Briefly, patients with a recent history of colorectal adenomas were recruited between during 1994–1998 from nine clinical centers in the US and Canada. Eligible subjects were between 21 and 80 years of age, in good health, had a complete colonoscopy within 3 months before enrollment with no known polyps left in the bowel, and had received a recommendation for a 3-year follow-up colonoscopy by their regular medical practitioner.

At enrollment, eligible subjects completed a questionnaire regarding demographics, medical history, and lifestyle habits. All participants were asked to avoid the use of aspirin, NSAIDs, and supplements containing folic acid for the duration of active treatment. Each subject underwent a three-month, single-blind run-in period on 325 mg of aspirin per day. Only subjects with at least 80% compliance and no adverse effects of aspirin were randomly assigned to receive aspirin placebo, low-dose aspirin (81 mg/d), or high-dose aspirin (325 mg/d), and to receive folate placebo or folate (1 mg/d). A total of 1,021 full factorial
subjects were randomized, as well as 100 “aspirin-only” subjects who were recruited before
the folic acid component of the study was added.

By protocol, participants were to remain on study treatment until their anticipated
surveillance colonoscopy, about three years after the qualifying exam. Every 4 months
during study treatment subjects completed a questionnaire regarding compliance with study
agents, use of medications and supplements, large bowel endoscopy, and medical events. In
addition to a baseline blood sample, blood and spot urine samples were obtained late in the
third year of participation, and stored at −70°C. Compliance in the initial 3-year treatment
period was excellent: 87% to 95% of subjects took study pills at least 6 days per week
during those three years. All study aspirin treatment ended on September 28, 2001.

Study outcomes

The primary outcome — adenoma occurrence — was determined by colonoscopy and
confirmed by pathology review. All lesions removed from the large bowels of study subjects
were reviewed by a single study pathologist. Polyps were classified as adenomatous,
hyperplastic, serrated or other polyps; the degree of dysplasia and the extent of villous
component in each adenoma were recorded. Low-risk adenomas were defined as solitary
adenomatous polyps < 1 cm in greatest diameter with tubular histology. Advanced
adenomas were defined as adenomatous polyps with an estimated diameter of ≥1 cm, or at
least 25% villous component, any high-grade dysplasia, or invasive cancer. “High risk
findings” were one or more advanced adenomas or two or more low risk adenomas [for
comparability with previous studies (34, 35)].

Laboratory Measurements

Catabolism of PGE2 results in an end metabolite, 11-α-hydroxy-9,15-dioxo-2,3,4,5-tetranor-
prostone-1,20-dioic acid (PGE-M), that is excreted in the urine and stable for prolonged
periods when stored at −70°C (36). Measurement of urine PGE-M is a better measure of
systemic PGE2 production than plasma measurements (37, 38) because PGE2 in plasma is
rapidly metabolized in the lungs and consequently may not accurately reflect endogenous
prostaglandin production (39). We measured urinary PGE-M using liquid chromatography-
mass spectrometry (LC/MS) as previously described (40) on a Waters Acquity UPLC
coupled to a Thermo Scientific Quantum Vantage triple quadruple mass spectrometer.

2,3-dinor-6-keto-PGF1α (PGI-M), a major urinary metabolite of prostacyclin PGI2, was
measured using gas chromatography-negative-ion chemical ionization mass spectrometry
(GC/NICI-MS) as previously described (41) on an Agilent 5973 Inert Mass Selective
Detector coupled with an Agilent 6890n Network GC system (Agilent Labs, Torrance, CA).
11-dehydro-thromboxane B2 (dTxB2), a major urinary enzymatic metabolite of
thromboxane A2, was measured using GC/NICI-MS as previously described (42) using the
same instruments as described above for PGI-M.

PGE-M, PGI-M, and dTxB2 were successfully measured in 871, 816, and 854 samples,
respectively, in the Eicosanoid Core Laboratory at Vanderbilt University. The lower limit of
detection of PGE-M and PGI-M was 15 pg/mg creatinine, and of dTxB2, 20 pg/mg
creatinine. Inter-day and intra-day variability was less than 10% for all prostanoids assays.

Cancer Prev Res (Phila). Author manuscript; available in PMC 2016 November 01.
Quality control samples, from a pooled urine collection, were included with each batch analyzed. Between-batch coefficients of variation were 8.38%, 5.92%, and 7.32%, for the PGE-M, PGI-M, and dTxB₂ assays, respectively. Urinary prostanoid metabolite levels were expressed as pg/mg of analyte per mg of urinary creatinine. Urinary creatinine was measured using a chemical assay based on Jaffe’s reaction according to the manufacturer’s instructions (Enzo Life Sciences, NY, USA). Laboratory staff was blinded to the treatment group assignment of urine samples and the identity of quality control samples included in the study. Samples from each treatment group were randomly included in every batch.

Statistical Analyses

Levels of urinary biomarkers were natural log transformed to improve normality. Values below limit of detection (LOD) were replaced by LOD/2 [n = 4 (0.5%) for PGE-M, n = 61 (7.5%) for PGI-M, and n = 4 (0.5%) for dTxB₂]. Fisher exact tests (for categorical variables) and analysis of variance (for continuous variables) were used to compare randomized treatment groups. Correlations between biomarkers were calculated using Spearman’s correlation coefficient (ρ). We used modified Poisson regression with a robust variance estimate to calculate the relative risks (RR) and 95% confidence intervals (CIs) of having at least one adenoma (43). Covariates in multivariable models included age (continuous), sex, clinical center, number of lifetime adenoma before randomization (continuous), follow-up time (continuous), and aspirin treatment group assignment (placebo, 81 mg/d, or 325 mg/d). In secondary analyses, we further adjusted for randomized folic acid treatment, first degree family history of colorectal cancer (yes/no), body mass index (<25, 25 – 30, ≥30 kg/m²) at study enrollment, alcohol use (yes/no), smoking status (never/former/current). Inclusion of these covariates in the models did not substantially change the results; therefore, only the most parsimonious models are presented. Adjusted estimates are presented for one or more adenomas, low-risk adenomas, or advanced adenomas, and high-risk findings. The ratio of PGE-M to PGI-M was used as an indicator of trade-off between systemic PGE₂ and PGI₂ production. Since PGE₂ and thromboxane A₂ are hypothesized to be pro-carcinogenic, and PGI₂ possibly anti-carcinogenic, we used the ratio of (PGE-M + dTxB₂) to PGI-M as an indicator of the overall balance between potentially pro- and anti-carcinogenic effects of prostanoids. The P-value for trend was calculated using the continuous biomarker levels entered linearly into the models. An interaction by aspirin treatment assignment (placebo, 81 mg/d, or 325 mg/d) was assessed by including the cross product of the treatment variable and urinary biomarker, and evaluated using the Wald test. To compare our results with previous findings (35), we conducted an analysis of aspirin treatment effects within low (first tertile) and high (second and third tertiles) PGE-M levels. All statistical tests were two-sided. Statistical analysis was conducted using Stata (version 12.1, Stata Corp, College Station, TX).

RESULTS

There were 1,121 individuals randomized to aspirin or placebo, and 1,084 (97%) underwent a follow-up examination. A total of 876 participants (80.8% of with a follow-up exam) had at least one urinary prostanoid metabolite measurement; 328 had one or more recurrent adenoma at the end of treatment follow-up. The baseline characteristics of the subjects in the
three aspirin treatment arms with at least one biomarker measurement are shown in Table 1. Participants with missing urine measurements (n = 208) were comparable with those included in the analysis (n = 876) with respect to baseline characteristics and treatment randomization (data not shown). There were no statistically significant differences among the three arms with regard to demographic and lifestyle factors at the study entry. The three treatment arms were also similar with regard to the percentage of subjects who were randomly assigned to folic acid treatment.

Among all study participants, urinary PGE-M levels were weakly correlated with urinary PGI-M (ρ = 0.16, P < 0.001) and dTxB2 (ρ = 0.30, P < 0.001), with a somewhat stronger correlation between urinary PGI-M and dTxB2 (ρ = 0.41, P < 0.001). Similar correlations were found in each treatment group (data not shown). Urinary PGE-M, PGI-M and dTxB2 differed by sex, age, study center, and smoking status (Supplementary Tables 1 and 2). Aspirin treatment was associated with lower urinary prostanoid metabolites levels in a dose-dependent manner (Figure 1). Urinary PGE-M levels were proportionately 28% and 18% lower in participants who took 325 mg and 81 mg of aspirin, respectively, compared to participants who took placebo. Similar but weaker dose-dependent patterns were observed for PGI-M and dTxB2. However, the ratios of PGE-M to PGI-M, and of (dTxB2 + PGE-M) to PGI-M did not differ substantially by aspirin treatment group (Supplementary Table 3). Comparable aspirin-associated decreases in the levels of urinary prostanoid metabolites were observed in the data further stratified by the folic acid treatment arm (Supplementary Table 4).

There was no evidence that urinary PGE-M, PGI-M, or dTxB2 concentrations (or their ratios) were associated with risk for any adenoma: RR’s quartiles 2 through 4 were each close to 1.0, with narrow confidence intervals. Urinary PGE-M, PGI-M, and dTxB2 also had no material associations with low risk adenomas or high risk findings (Table 2). In contrast, the PGE-M, PGI-M, and to a lesser extent dTxB2, showed non-significant suggestions of decreasing RR trends with increasing urinary levels for advanced adenomas (Table 2). The ratios of PGE-M to PGI-M, and of (dTxB2 + PGE-M) to PGI-M were unassociated with risk of any adenomas class (Supplementary Table 5). None of the interactions of urinary prostanoid levels with aspirin treatment group were statistically significant (all P > 0.1). We repeated the analyses in the placebo group only and obtained similar findings (Supplementary Table 6).

In an exploratory analysis, in which we stratified the study participants according to PGE-M levels (Figure 2), aspirin treatment was associated with a significant reduction in any and high-risk adenoma occurrence among participants with low PGE-M (<5.34 ng/mg creatinine), but not among participants with high PGE-M (≥5.34 ng/mg creatinine) concentrations; however neither of the interactions were statistically significant. A similar pattern was suggested for low-risk adenoma, whereas for advanced adenoma the aspirin treatment effects did not seem to differ by PGE-M levels. Analyses stratified by sex (Supplementary Tables 7 and 8) suggested that among women, higher levels of PGE-M may be positively associated with adenoma occurrence; however, the results were based on a very small number of events and no statistically significant multiplicative interactions by sex were found.
Two participants in the low-dose aspirin group and three in the high-dose aspirin group were classified as aspirin resistant based on dTxB₂ > 1.5 ng/mg creatinine (>67.9 ng/mmol of creatinine)(44, 45). Among the 5 participants, one did not take the study treatment (0% compliance). The exclusion of these subjects from the analysis had no effect on the risk estimates (data not shown).

DISCUSSION

In this analysis of individuals participating in a randomized clinical trial, aspirin treatment was associated with lower levels of urinary prostanoids in a dose-dependent manner: participants who took high-dose aspirin had the lowest concentrations of PGE-M, PGI-M, and dTxB₂ compared to participants who took low-dose of aspirin or placebo. None of the urinary prostanoid metabolites or their ratios were significantly associated with risk of any measure of adenoma risk.

The levels of COX-derived PGE₂ and its synthase are increased in colorectal neoplasia as compared with histologically normal tissue.(16, 46, 47) PGE₂ has been shown to inhibit apoptosis, promote cellular proliferation, angiogenesis, and migration, and activate Wnt signaling (48, 49). In previous studies, the major urinary metabolite of PGE₂, PGE-M, has been strongly directly associated with the risk of colorectal cancer (50) and (contrary to our findings) with multiple and/or advanced adenoma (RR ranging from 1.66 to 2.19 comparing the highest vs. lowest quartiles) (34, 35). Conversely, in our study, the association pattern for PGE-M with advanced adenomas was non-significantly suggestive of an inverse association. In agreement with our results, previous studies have not found an association with single small tubular adenoma (34, 35) or any adenoma (35). There are no previous data regarding the associations of metabolites of PGI₂ and thromboxane with risk of colorectal cancer or adenomas of any type.

Our data suggested that aspirin treatment was associated with a significant reduction in adenoma occurrence in participants with low PGE-M, measured near the end of treatment. This is expected, since both adenoma occurrence and low PGE-M levels are associated with aspirin use and so are logically associated with each other. Trial participants on aspirin tended to have low PGE-M and also had the lowest adenoma risk. On the other hand, participants with high PGE-M tended to be placebo subjects (or aspirin subjects resistant to treatment) and had a higher risk for adenomas. Bezawada et al (35) also investigated this issue, and reported the opposite pattern: stronger aspirin/NSAIDs associations in women with higher urinary PGE-M. To the extent that their samples were truly baseline (taken before starting aspirin), their data are logical and do not conflict with ours. However, to the extent their data were taken “on treatment” (during aspirin/NSAID use) the two studies disagree. It is not clear whether their samples were truly baseline as they were collected at the same time as information on aspirin/NSAID use.

Thromboxane is essential to platelet activation, and the antiplatelet effect of aspirin through inhibition of thromboxane synthesis seems to be relevant to its anti-neoplastic actions (11). It has been proposed that activated platelets promote COX-2 upregulation in adjacent cells of the colorectal mucosa at sites of mucosal injury via paracrine lipid (e.g., TXA₂) or protein
(e.g., interleukin-1β) mediators (11). There may also be direct effects of TXA₂ itself, as mucosal levels stimulate proliferation, promote metastases, and mediate COX-2-dependent angiogenesis (13, 51–53). Elevated levels of TXA₂ and thromboxane synthase in colorectal neoplasms have been reported, (13, 17) and colon cancer cells that overexpress the TXA₂ synthase gene grow faster and exhibit more abundant vasculature (12). In our study, higher levels of dTxB₂, a major urinary metabolite of TXA₂, were not associated with the risk of any adenoma. In contrast to our initial hypothesis, the association pattern for dTxB₂ was suggestive of an inverse association for advanced adenoma at the end of study follow-up among all and placebo-only participants.

Prostacyclin PGI₂, a prostaglandin downstream of COX-1 and COX-2 (54), has important roles in cardiovascular homeostasis, and may also influence carcinogenesis. Expression of PGI₂ seems reduced in lung cancers (15), and administration of a PGI₂ analogue improves endobronchial dysplasia in former smokers (18). Colon cancer cells that overexpress the prostacyclin synthase gene (PTGIS) grow more slowly and with less developed vasculature than control inoculants (12). Epigenetic inactivation of the PTGIS is common in colorectal cancer (19), and reduced tumor tissue levels of PGI₂ compared with adjacent normal mucosa have been reported (16, 17). Furthermore, a polymorphism associated with reduced activity of the PTGIS gene has been associated with an increased risk of colorectal adenomas and a blunting of the protective effect of NSAIDs (20). Our results show that PGI-M, a major urinary metabolite of PGI₂, was not associated with any adenoma, but – in agreement with our initial hypothesis – higher levels of PGI-M were non-significantly inversely associated with advanced adenoma.

Although numerous epidemiological studies and several randomized clinical trials reported that aspirin is protective against colorectal neoplasms (1–5); it is unknown exactly how it exerts its anti-neoplastic effects, and why the lower aspirin dose seems more effective in preventing colorectal adenomas. The main proposed mechanism so far includes inhibition of COX enzymes. We also hypothesized that a trade-off between pro-carcinogenic PGE₂ and/or TXA₂, and putatively anti-carcinogenic PGI₂ may explain a protective effect of lower dose aspirin. However, our findings did not suggest that differential inhibition of prostanoids explains the paradoxical aspirin effects, nor support the COX-dependent anti-neoplastic effects of aspirin, as urinary prostanoid metabolites were not associated with risk for colorectal adenoma occurrence. It is possible that COX-independent effects of aspirin are involved, including inhibition of PPARδ (55) and oxidative DNA damage (56), modulation of polyamine metabolism (57) and Wnt signaling (58), activation of the NF-kB signaling pathway resulting in apoptosis (59), increase in leukotriene production as a result of shuttling of free arachidonic acid between COXs and lipoxygenases pathways (58), and activation of the NSAID-activated gene (NAG-1) (60), which is a member of the TGF-β family that has pro-apoptotic and anti-tumorigenic activities (61).

This study has several limitations. First, participants in this analysis had a previous history of at least one colorectal adenoma, potentially limiting the generalizability of the data. In particular, it is possible that the colorectal mucosa that has had prior adenoma may already have altered prostanoid signaling (e.g., a deficiency in 5-Hydroxyprostaglandin dehydrogenase [15-PGDH]), which may efface the differences in levels between those who
do, and do not, have a recurrent adenoma on a later colonoscopy. Second, we did not measure the prostanoids concentrations at the study entry, as we did not collect urinary samples at that time. However, the on-treatment measurements better reflect the subjects’ metabolic milieu during the period the adenomas were forming. Third, the ratios of PGE-M and dTxB2, individually and in combination, to PGI-M may not adequately capture trade-off between systemic productions of these prostanoids. Fourth, treatment effects of aspirin on the prostanoid synthesis in the colon mucosa are unclear, as we did not measure the expression of prostanoids in the colorectal tissue. Finally, we had a limited sample size to investigate risk of advanced adenoma occurrence.

The strengths of this study include the randomized, placebo-controlled trial design; high protocol adherence by study participants; the high follow-up rate; and the systemic collection of risk factor information at baseline and follow-up intervals as well as outcomes at the end of treatment. Another strength of this study is that we measured stable biomarkers of systemic production of prostanoids using highly sensitive assays in one laboratory. Finally, this study is the first human study to investigate the association between PGI-M and dTxB2 and risk of colorectal adenoma.

Overall, in this study, we found that low- and high-dose aspirin is associated with lower levels of urinary prostanoid metabolites in a dose-dependent manner. However, our findings provided no evidence that urinary prostanoid metabolites are associated with reduced risk of colorectal adenoma occurrence. In agreement with our original hypothesis, we observed a possible inverse association of metabolites of the possibly anti-carcinogenic prostanoid PGI2 with advanced adenoma occurrence. However similar inverse associations were observed for other potentially pro-carcinogenic prostanoids. Studies of colorectal neoplasms that measure changes in both systemic and tissue-specific levels of prostanoids in response to aspirin treatment are needed to confirm our results.

**Supplementary Material**

Refer to Web version on PubMed Central for supplementary material.

**Acknowledgements**

We thank the subjects in the trial for their dedication and cooperation.

**Funding:** This study was funded by the US Public Health Service, grant CA059005. Dr. Bradshaw was supported by National Institute of Health training award K12CA120780

**References**


Cancer Prev Res (Phila). Author manuscript; available in PMC 2016 November 01.


Figure 1.
Urinary PGE-M (A), PGI-M (B) and dTxB₂ (C) levels by aspirin treatment group, the Aspirin/Folate Polyp Prevention Study.
* P-value vs. placebo. ** The proportional difference in geometric means was calculated as ((treatment group – placebo group)/placebo group)*100%. *** P-value vs. 81 mg.
Figure 2.
Association of adenoma occurrence during study treatment within categories of urinary PGE-M levels by aspirin treatment group, the Aspirin/Folate Polyp Prevention Study. All models were adjusted for age (continuous), sex, center (categorical), number of previous adenomas (continuous), and follow-up time (continuous). Placebo was the reference group. Low PGE-M category = tertile 1, High PGE-M category = tertile 2 & tertile 3.
Table 1
Baseline selected characteristics of the study participants with at least one biomarker measurement.

<table>
<thead>
<tr>
<th>Baseline characteristic</th>
<th>Placebo (N = 296)</th>
<th>Aspirin 81 mg/d (N = 296)</th>
<th>Aspirin 325 mg/d (N = 284)</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, yrs, mean (SD)</td>
<td>57.5 (9.8)</td>
<td>57.2 (9.4)</td>
<td>57.6 (4.5)</td>
<td>0.88</td>
</tr>
<tr>
<td>Male sex, no. (%)</td>
<td>189 (63.9)</td>
<td>193 (65.2)</td>
<td>178 (62.7)</td>
<td>0.83</td>
</tr>
<tr>
<td>White, no. (%)</td>
<td>253 (85.5)</td>
<td>260 (87.8)</td>
<td>250 (88.0)</td>
<td>0.14</td>
</tr>
<tr>
<td>BMI, kg/m²*, mean (SD)</td>
<td>27.3 (4.5)</td>
<td>27.3 (4.5)</td>
<td>27.7 (4.8)</td>
<td>0.91</td>
</tr>
<tr>
<td>Overweight, no. (%)</td>
<td>140 (47.5)</td>
<td>142 (48.1)</td>
<td>130 (45.8)</td>
<td>0.89</td>
</tr>
<tr>
<td>Obese, no. (%)</td>
<td>63 (21.4)</td>
<td>60 (20.3)</td>
<td>70 (24.7)</td>
<td>0.41</td>
</tr>
<tr>
<td>Current smoker, no. (%)</td>
<td>41 (13.9)</td>
<td>40 (13.6)</td>
<td>41 (14.5)</td>
<td>0.56</td>
</tr>
<tr>
<td>Alcohol drinker, no. (%)</td>
<td>199 (67.3)</td>
<td>189 (63.9)</td>
<td>193 (68.0)</td>
<td>0.62</td>
</tr>
<tr>
<td>Multivitamin use, no. (%)</td>
<td>104 (35.1)</td>
<td>111 (37.5)</td>
<td>96 (33.8)</td>
<td>0.40</td>
</tr>
<tr>
<td>Family history of colorectal polyps, no. (%)</td>
<td>62 (26.8)</td>
<td>72 (29.2)</td>
<td>77 (34.1)</td>
<td>0.13</td>
</tr>
<tr>
<td>Adenoma characteristics (at baseline)**</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Number, mean (SD)</td>
<td>1.6 (0.9)</td>
<td>1.6 (1.0)</td>
<td>1.6 (1.1)</td>
<td>0.41</td>
</tr>
<tr>
<td>Advanced adenomas, no. (%)***</td>
<td>84 (36.1)</td>
<td>80 (34.9)</td>
<td>89 (39.6)</td>
<td>0.69</td>
</tr>
<tr>
<td>Proximal location, no. (%)</td>
<td>136 (54.4)</td>
<td>135 (54.0)</td>
<td>133 (55.0)</td>
<td>0.74</td>
</tr>
<tr>
<td>Folic acid treatment group, no. (%)</td>
<td>149 (51.0)</td>
<td>147 (49.7)</td>
<td>143 (50.7)</td>
<td>0.98</td>
</tr>
</tbody>
</table>

* N=1 in the placebo, and N=1 in the aspirin 81 mg/d group had missing BMI data.

** Data available for 268, 257, 243 participants in the placebo, aspirin 81 mg/d, and aspirin 325 mg/d treatment groups, respectively.

*** Data available for 233, 229, and 229 in the placebo, aspirin 81 mg/d, and aspirin 325 mg/d treatment groups, respectively.
### Table 2

Association of adenoma occurrence during study treatment with urinary PGE-M, PGI-M and dTxB₂ levels, the Aspirin/Folate Polyp Prevention Study.

<table>
<thead>
<tr>
<th>Biomarker / Quartile range</th>
<th>Any adenoma</th>
<th>Low-risk adenoma</th>
<th>High-risk findings</th>
<th>Advanced adenoma</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Cases/total N</td>
<td>RR**</td>
<td>95% CI</td>
<td>Cases/total N</td>
</tr>
<tr>
<td>PGE-M, ng/mg creatinine</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt;4.57</td>
<td>71/207</td>
<td>1.00 (ref)</td>
<td></td>
<td>36/172</td>
</tr>
<tr>
<td>4.57–7.37</td>
<td>77/207</td>
<td>1.00</td>
<td>0.78</td>
<td>1.29</td>
</tr>
<tr>
<td>7.37–11.38</td>
<td>86/210</td>
<td>1.00</td>
<td>0.78</td>
<td>1.28</td>
</tr>
<tr>
<td>≥11.39</td>
<td>93/211</td>
<td>1.07</td>
<td>0.83</td>
<td>1.37</td>
</tr>
<tr>
<td><em>P</em>&lt;sub&gt;trend&lt;/sub&gt;</td>
<td></td>
<td>0.92</td>
<td>0.53</td>
<td></td>
</tr>
<tr>
<td>PGI-M, ng/mg creatinine</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt;0.049</td>
<td>70/196</td>
<td>1.00 (ref)</td>
<td></td>
<td>33/159</td>
</tr>
<tr>
<td>0.049–0.082</td>
<td>82/194</td>
<td>1.14</td>
<td>0.90</td>
<td>1.45</td>
</tr>
<tr>
<td>0.082–0.124</td>
<td>76/194</td>
<td>1.05</td>
<td>0.82</td>
<td>1.36</td>
</tr>
<tr>
<td>≥0.125</td>
<td>80/196</td>
<td>1.04</td>
<td>0.81</td>
<td>1.35</td>
</tr>
<tr>
<td><em>P</em>&lt;sub&gt;trend&lt;/sub&gt;</td>
<td></td>
<td>0.92</td>
<td>0.22</td>
<td></td>
</tr>
<tr>
<td>dTxB₂, ng/mg creatinine</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt;0.107</td>
<td>68/199</td>
<td>1.00 (ref)</td>
<td></td>
<td>31/162</td>
</tr>
<tr>
<td>0.107–0.194</td>
<td>96/209</td>
<td>1.33</td>
<td>1.04</td>
<td>1.69</td>
</tr>
<tr>
<td>0.195–0.346</td>
<td>79/206</td>
<td>0.98</td>
<td>0.74</td>
<td>1.30</td>
</tr>
<tr>
<td>≥0.347</td>
<td>77/204</td>
<td>0.94</td>
<td>0.69</td>
<td>1.28</td>
</tr>
<tr>
<td><em>P</em>&lt;sub&gt;trend&lt;/sub&gt;</td>
<td></td>
<td>0.93</td>
<td>0.86</td>
<td></td>
</tr>
</tbody>
</table>

*Low-risk adenoma defined as solitary adenomatous polyp < 1 cm in greatest diameter and tubular or unknown histology (high-risk adenoma excluded from the analysis). High-risk adenoma defined as adenomatous polyp ≥1 cm in greatest diameter and/or tubulovillous, villous or high-grade dysplasia, histology or multiple (≥2) adenomatous polyps of any size or histology (low-risk adenoma excluded from the analysis). Advanced adenoma defined as adenomatous polyp with an estimated diameter of ≥1 cm, or at least 25% villous component, any high-grade dysplasia, or invasive cancer.

**Adjusted for age (continuous), sex, center (categorical), number of previous adenomas (continuous), follow-up time (continuous) and aspirin treatment group (placebo, 81 mg, 325 mg).

#P-values for interaction for aspirin treatment group were: for any adenoma, *P* PGE-M = 0.14, *P* PGI-M = 0.40, and *P* dTxB₂ = 0.50; for low-risk adenoma, *P* PGE-M = 0.13, *P* PGI-M = 0.22, and *P* dTxB₂ = 0.40; for high-risk adenoma, *P* PGE-M = 0.53, *P* PGI-M = 0.45, and *P* dTxB₂ = 0.63; and for advanced adenoma, *P* PGE-M = 0.62, *P* PGI-M = 0.33, and *P* dTxB₂ = 0.32.