Concomitant herpes simplex virus colitis and hepatitis in a man with ulcerative colitis: Case report and review of the literature

Varun K. Phadke, Emory University
Rachel Friedman-Moraco, Emory University
Brian Quigley, Emory University
Alton Farris III, Emory University
John Norvell, Emory University

Journal Title: Medicine
Volume: Volume 95, Number 42
Publisher: Lippincott, Williams & Wilkins: Medicine | 2016-10-01, Pages e5082-e5082
Type of Work: Article | Final Publisher PDF
Publisher DOI: 10.1097/MD.0000000000005082
Permanent URL: https://pid.emory.edu/ark:/25593/rtg24

Final published version: http://dx.doi.org/10.1097/MD.0000000000005082

Copyright information:
© 2016 the Author(s). Published by Wolters Kluwer Health, Inc. All rights reserved.
This is an Open Access work distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Accessed November 8, 2018 11:48 AM EST
Concomitant herpes simplex virus colitis and hepatitis in a man with ulcerative colitis

Case report and review of the literature

Varun K. Phadke, MD\textsuperscript{a,∗}, Rachel J. Friedman-Moraco, MD\textsuperscript{a}, Brian C. Quigley, MD\textsuperscript{b}, Alton B. Farris, MD\textsuperscript{b}, J. P. Norvell, MD\textsuperscript{c,d}

Abstract

Background: Herpesvirus infections often complicate the clinical course of patients with inflammatory bowel disease; however, invasive disease due to herpes simplex virus is distinctly uncommon.

Methods: We present a case of herpes simplex virus colitis and hepatitis, review all the previously published cases of herpes simplex virus colitis, and discuss common clinical features and outcomes. We also discuss the epidemiology, clinical manifestations, diagnosis, and management of herpes simplex virus infections, focusing specifically on patients with inflammatory bowel disease.

Results: A 43-year-old man with ulcerative colitis, previously controlled with an oral 5-aminosalicylic agent, developed symptoms of a colitis flare that did not respond to treatment with systemic corticosteroid therapy. One week later he developed orolabial ulcers and progressive hepatic dysfunction, with markedly elevated transaminases and coagulopathy. He underwent emergent total colectomy when imaging suggested bowel micro-perforation. Pathology from both the colon and liver was consistent with herpes simplex virus infection, and a viral culture of his orolabial lesions and a serum polymerase chain reaction assay also identified herpes simplex virus. He was treated with systemic antiviral therapy and made a complete recovery.

Conclusions: Disseminated herpes simplex virus infection with concomitant involvement of the colon and liver has been reported only 3 times in the published literature, and to our knowledge this is the first such case in a patient with inflammatory bowel disease. The risk of invasive herpes simplex virus infections increases with some, but not all immunomodulatory therapies. Optimal management of herpes simplex virus in patients with inflammatory bowel disease includes targeted prophylactic therapy for patients with evidence of latent infection, and timely initiation of antiviral therapy for those patients suspected to have invasive disease.

Abbreviations: ALT = alanine transaminase, AST = aspartate transaminase, CMV = cytomegalovirus, CT = computed tomography, ECCO = European Crohn’s and Colitis Organisation, FDA = Food and Drug Administration, HIV/AIDS = human immunodeficiency virus/acquired immunodeficiency syndrome, HSV = herpes simplex virus, IBD = inflammatory bowel disease, IHC = immunohistochemistry, IL = interleukin, PCR = polymerase chain reaction, TNF = tumor necrosis factor, UC = ulcerative colitis.

Keywords: case report, Crohn’s disease, herpes simplex virus, immunomodulatory therapy, inflammatory bowel disease, ulcerative colitis.

1. Introduction

Herpesvirus infections frequently complicate the clinical course of patients with inflammatory bowel disease (IBD).\textsuperscript{[1,2]} Immunosuppressive therapy increases the risk of herpesvirus reactivation and predisposes to more severe infections.\textsuperscript{[3]} Whereas cytomegalovirus (CMV) is a well-recognized cause of tissue-invasive disease in this population,\textsuperscript{[11]} herpes simplex virus (HSV) has only rarely been implicated.\textsuperscript{[15–7]} When HSV infections do occur they are usually limited to sites of local reactivation, for example, mucosal surfaces...
(such as the oropharynx, anogenital area, and eyes) and skin sites. Here we report the first case, to our knowledge, of a patient with IBD who developed disseminated HSV infection with concomitant colonic and hepatic involvement. We also review all of the previously reported cases of HSV colitis and hepatitis, as well as the published literature of HSV infections, in patients with IBD.

The patient provided informed consent for the use of clinical information and images for this case report.

2. Case report

A 43-year-old man with ulcerative colitis (UC) was transferred to our hospital for evaluation and management of refractory colitis symptoms and abnormal liver function tests. One year earlier he had been diagnosed with UC and started on maintenance therapy with oral balsalazide. He continued to have intermittent flares of his colitis which were well-controlled with short courses of systemic corticosteroids. After his last flare 6 months earlier, he was prescribed a trial of azathioprine, which he discontinued after a few weeks due to intolerance, followed by mesalamine suppositories. One month prior to admission he was asymptomatic.

Approximately 3 weeks prior to transfer he developed fatigue, decreased oral intake, diffuse crampy abdominal discomfort, and loose bloody stools, consistent with his prior UC flares. He was prescribed prednisone 40mg daily, but his symptoms did not improve after 1 week of this therapy. He was admitted to his local hospital. Laboratory testing was unremarkable (Table 1) and stool evaluation for bacterial and parasitic pathogens was unrevealing. A flexible sigmoidoscopy demonstrated “severe ulcerative colitis” without pseudomembranes; biopsies were not performed. He was treated with intravenous methylprednisolone and discharged with prednisone 60mg daily and a course of metronidazole—however, he was readmitted 1 week later for persistent symptoms. Laboratory testing on readmission was notable for: aspartate aminotransferase (AST), 136U/L; alanine aminotransferase (ALT), 32U/L; alkaline phosphatase, 32U/L; prothrombin time of 15.7 seconds (international normalized ratio, INR, of 1.25). Computed tomography (CT) of the abdomen and pelvis demonstrated pan-colitis. He was started on intravenous corticosteroids and empiric broad-spectrum antibiotics. For persistent colitis symptoms, he also received a dose of golimumab 48 hours after admission.

Over the next 96 hours, his serum aminotransferase levels and prothrombin time continued to increase (Table 1). A serologic evaluation for viral causes of acute hepatitis was notable for a positive CMV IgG and HSV-1 IgM; tests for hepatitis A and B and HSV-2 were negative. He subsequently underwent percutaneous liver biopsy, and then was transferred to our hospital for further management.

On arrival to our hospital, he was febrile to 39.6°C and in distributive shock. On physical examination, he had multiple small shallow ulcerations of the hard palate as well as diffuse abdominal tenderness to palpation. Laboratory testing was notable for pancytopenia and markedly elevated serum transaminases, with AST, 3072 U/L, and ALT, 3392 U/L (other results in Table 1). Blood cultures were obtained, swabs of his oral ulcers

### Table 1

**Progression of laboratory values of case patient.**

<table>
<thead>
<tr>
<th>Variable</th>
<th>Reference range, adults</th>
<th>3 weeks before transfer</th>
<th>11 days before transfer</th>
<th>4 days before transfer</th>
<th>2 days before transfer</th>
<th>On transfer</th>
<th>Hospital day 2</th>
<th>Hospital day 10</th>
<th>Discharge</th>
</tr>
</thead>
<tbody>
<tr>
<td>White-cell count, per mm³</td>
<td>4200–9100</td>
<td>16,300</td>
<td>21,200</td>
<td>7300</td>
<td>10,600</td>
<td>3400</td>
<td>21,200</td>
<td>5600</td>
<td>6300</td>
</tr>
<tr>
<td>Hemoglobin, g/dL</td>
<td>12.9–16.1</td>
<td>14.3</td>
<td>12.9</td>
<td>12.9</td>
<td>9.2</td>
<td>6.6</td>
<td>7.5</td>
<td>7.0</td>
<td>14.0</td>
</tr>
<tr>
<td>Platelet count, per mm³</td>
<td>150,000–400,000</td>
<td>340,000</td>
<td>324,000</td>
<td>300,000</td>
<td>197,000</td>
<td>127,000</td>
<td>104,000</td>
<td>140,000</td>
<td>207,000</td>
</tr>
<tr>
<td>Creatinine, mg/dL</td>
<td>0.7–1.2</td>
<td>1.0</td>
<td>0.8</td>
<td>1.1</td>
<td>1.1</td>
<td>0.9</td>
<td>0.81</td>
<td>0.38</td>
<td>1.00</td>
</tr>
<tr>
<td>Aspartate aminotransferase, U/L</td>
<td>15–41</td>
<td>18</td>
<td>13</td>
<td>98</td>
<td>316</td>
<td>3702</td>
<td>3726</td>
<td>55</td>
<td>20</td>
</tr>
<tr>
<td>Alanine aminotransferase, U/L</td>
<td>&lt;44</td>
<td>16</td>
<td>16</td>
<td>136</td>
<td>438</td>
<td>3392</td>
<td>2312</td>
<td>173</td>
<td>22</td>
</tr>
<tr>
<td>Alkaline phosphatase, U/L</td>
<td>&lt;32</td>
<td>73</td>
<td>50</td>
<td>62</td>
<td>57</td>
<td>77</td>
<td>122</td>
<td>156</td>
<td>63</td>
</tr>
<tr>
<td>Total bilirubin, mg/dL</td>
<td>0.3–1.2</td>
<td>1.0</td>
<td>0.8</td>
<td>1.6</td>
<td>0.8</td>
<td>0.9</td>
<td>1.7</td>
<td>1.6</td>
<td>1.1</td>
</tr>
<tr>
<td>Prothrombin time, seconds</td>
<td>9.4–12.5</td>
<td>–</td>
<td>–</td>
<td>15.7</td>
<td>–</td>
<td>16</td>
<td>19.5</td>
<td>12.8</td>
<td>10.6</td>
</tr>
<tr>
<td>International normalized ratio</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>1.25</td>
<td>1.52</td>
<td>1.85</td>
<td>1.22</td>
<td>1.01</td>
<td></td>
</tr>
</tbody>
</table>

g/dL = grams/dL, mg/dL = milligrams/dL, mm³ = cubic millimeter, U/L = units/Liter.

*Reference ranges used at Emory University Hospital."
were sent for viral culture, and serum polymerase chain reaction (PCR) assays for CMV and HSV were performed. A repeat CT of the abdomen and pelvis revealed diffuse colonic wall thickening and a micro-perforation in the distal sigmoid colon.

The patient received intravenous fluid boluses, transfusions of packed red cells, and empiric antimicrobial therapy with vancomycin, piperacillin/tazobactam, and intravenous ganciclovir. He underwent emergent total abdominal colectomy with end ileostomy as well as intraoperative liver biopsy. Blood cultures obtained on admission grew Escherichia coli and Citrobacter amalonaticus—his anti-bacterial therapy was changed to intravenous ertapenem. Viral culture of his oral ulcers grew herpes simplex virus (Fig. 1). A serum PCR assay was positive for HSV (cycle threshold value of 18.5, cutoff for positivity <39), but negative for CMV. Pathology slides sent from the referring hospital and intraoperative specimens from our hospital were reviewed. The colon had “chronic active colitis with ulceration” (Fig. 2A) and the liver had “patchy foci of nonzonal hepatocellular necrosis” (Fig. 2B). Histopathology and immunohistochemistry (IHC) of both the liver and colon were consistent with HSV infection (Fig. 2A–D); IHC stains for CMV and adenovirus were negative.

His antiviral therapy was changed to intravenous acyclovir for disseminated HSV infection. His steroids were tapered over a period of 1 month. At the time of discharge, 10 days after admission, his serum AST and ALT levels had decreased to 173 U/L and 55 U/L, respectively. He was discharged with oral valacyclovir 1000 mg 3 times daily to take for an additional 4 weeks, to be followed by indefinite suppressive therapy. Four months after discharge his serum transaminase levels had returned to normal. In the 1 year following discharge, he has remained clinically well with no evidence of relapse.

3. Discussion

3.1. Epidemiology of HSV infections in IBD

Herpes simplex virus 1 and 2 infections are common, with an estimated seroprevalence of 50% and 15% respectively in the general US population. Importantly, rates of infection vary between certain age, race, and socioeconomic groups—since no seroepidemiologic studies of latent HSV infection have been conducted specifically in patients with IBD, the prevalence in this particular population is unknown. Nevertheless, data from single

Figure 2. (A) Herpes simplex colitis, colectomy specimen. Deep ulceration in the colon (left) with adjacent nonulcerated mucosa (right). Hematoxylin and eosin stain, 20× overall magnification. (B) Herpes simplex virus hepatitis, intraoperative liver biopsy 1 day after initiation of antiviral therapy. Patches of nonzonal hepatocyte necrosis with minimal inflammation. Hematoxylin and eosin stain, 20× overall magnification. (C) Herpes simplex virus colitis. An immunohistochemical stain for herpes simplex virus (types 1 and 2, combined stain) darkly stains these inclusion-like structures (some denoted with arrows). Despite the apparently high background staining, note absence of staining within macrophage nuclei. No staining whatsoever was observed in the nonulcerated colonic mucosa. Immunohistochemical stain for herpes simplex virus, 1000× overall magnification. (D) Herpes simplex virus hepatitis. The patches of hepatocyte necrosis are highlighted strongly with an immunohistochemical stain for herpes simplex virus (types 1 and 2, combined stain). 20× overall magnification. Photo courtesy of Dr. Brian Quigley (Department of Pathology and Laboratory Medicine, Emory University).
center analyses and meta-analyses of multicenter trials indicate that HSV infections are fairly common among patients with IBD,[9-12] mirroring trends in the general population. In fact, the actual incidence is likely to be underestimated by these studies due to underreporting and inadequate duration of follow-up.

Use of immunomodulatory agents, particularly systemic corticosteroids and antimetabolites (e.g., azathioprine, 6-mercaptopurine, and methotrexate), has been associated with an elevated risk of localized mucocutaneous HSV reactivation,[9,10] similar to herpes zoster.[12] Invasive HSV infections have also been reported in patients taking azathioprine,[5-7,13] and methotrexate[14] for IBD. Post-marketing surveillance studies of IBD patients receiving newer immunomodulatory therapies (e.g., biologic therapies) less frequently evaluate the specific risk of HSV infections, as compared to herpes zoster, mycobacterial, and fungal infections.[11,13] As a result, the effect of biologic agents, such as tumor necrosis factor (TNF)-alpha antagonists, anti-integrin antibodies, or interleukin (IL)-12/23 receptor antagonists, on the incidence of HSV infections—both localized and invasive—among IBD patients is less clear. However, given the paucity of reported cases, the risk is likely lower than with antimetabolite therapy (Table 4).[11,13] Indeed, although there have been small case series of tissue-invasive HSV infections in patients receiving TNF-alpha antagonists and anti-integrin antibodies,[16-17] none have included patients with IBD.

Patient-specific immunologic factors can also influence the course of infections due to herpes simplex viruses. These may include individual-level variations in pathogen recognition receptors, major histocompatibility complex proteins, naturally occurring anti-viral peptides, and effector cell function.[18-21] Although IBD has been associated with a variety of genetic polymorphisms linked to a dysregulated immune response to enteric microbes, none of these genetic factors have been investigated specifically for their role in immunity to herpes simplex viruses.

3.2. Clinical manifestations of HSV infections in IBD

The most common clinical manifestations of HSV in IBD are similar to those in otherwise healthy individuals. Overall, mucocutaneous (i.e., oropharynx and anal/genital areas) disease predominates and is generally benign; however, progression to more invasive disease or widespread cutaneous dissemination has occasionally been described.[12,22,23] Importantly, oropharyngeal and anal/genital lesions due to HSV can mimic those seen with Crohn’s disease[24]; thus, it is important to exclude active HSV infection before initiating or escalating immunomodulatory therapy.[12] Herpes simplex viruses can also occasionally cause disseminated disease with end-organ involvement; however, there have been only a few reports of visceral HSV disease among patients with IBD. The gastrointestinal tract is a frequent site of HSV involvement in immunocompromised patients,[26] with the esophagus and rectum being the most frequently affected organs, followed by isolated reports of stomach and small bowel disease. Interestingly, the vast majority of these cases have been in patients with HIV/AIDS or among solid organ and bone marrow transplant recipients, and not those with IBD—we found only 1 report of HSV duodenitis in a patient with Crohn’s disease.[27] Involvement of other organs in IBD patients is even less common—to date only 2 cases of HSV meningoencephalitis[6,7] and a single case of HSV pneumonia/pneumonitis[5] have been reported, all in the setting of corticosteroid or anti-metabolite therapy.

Colitis due to HSV was first described in 1982[28] and since then only 16 cases have been reported in the literature.[4,19-24] Almost all of the cases occurred in immunocompromised hosts, nearly a third (n = 5) of whom were receiving corticosteroid therapy for IBD (Table 2).[4,33,39,41,42] In contrast, CMV is routinely implicated as an intestinal pathogen in patients with IBD. Indeed, among patients with IBD presenting with acute colitis, reported prevalence rates of CMV infection have ranged from 21% to 34%, and in those with steroid-refractory disease it can be as high 36%.[1] As is the case for CMV, HSV colitis in this population often mimics the underlying disease, presenting as fever, abdominal pain and diarrhea, rectal bleeding[4,39,42] or even paralytic ileus,[41] generally in the absence of suggestive mucocutaneous involvement. Endoscopic findings may be nonspecific and can include edema, erythema sometimes with petechiae, and well-demarcated ulcers of variable size and depth with or without purulent exudate, often separated by normal-appearing mucosa.[41,42]

The relative paucity of cases of HSV colitis as compared to CMV colitis (not only in the context of IBD) suggests a fundamental difference in the pathophysiology of end-organ involvement due to these viruses. Differences in the site of viral latency (e.g. neural ganglion cells for HSV vs leukocytes and endothelial cells for CMV) may account for the relative infrequency of colonic involvement due to HSV as compared to CMV. Alternatively, these viruses may differ in their tropism for colonic tissue. In studies that have exploited the oncolytic properties of herpes simplex viruses, investigators have shown diminished binding of HSV-1 to normal colonic tissue as compared to cancerous lesions.[43] Similarly, although HSV-1 has been used successfully in an ex vivo colonic explant model of virally induced immune activation in Crohn’s disease,[44] it is rarely detected in the large intestines of patients with IBD.[45,46]

Hepatitis due to HSV is also uncommon and almost exclusively occurs in the context of impaired cell-mediated immunity.[47] We found only 8 previous cases of HSV hepatitis in patients with IBD, all of whom were receiving systemic corticosteroids with or without anti-metabolite therapy (Table 3).[1,47-53] Similar to other patient populations, HSV hepatitis in patients with IBD tends to progress rapidly and is associated with a high case fatality rate, particularly if the diagnosis is not considered early enough in the disease course to allow for timely initiation of appropriate antiviral therapy.[151]

We identified only 3 prior reports[31,37,38] describing simultaneous colonic and hepatic involvement of disseminated herpes simplex virus infection (one of which did not report pathologic confirmation of hepatic involvement),[157] none of which involved a patient with IBD (Table 2). One of the cases was a neonate presumably with perinatal acquisition of HSV,[137] the second case was a patient who had received re-induction chemotherapy for acute lymphoblastic leukemia and developed reactivation HSV infection (prior history of herpes labialis) with multiorgan involvement,[38] and the third case was a liver transplant recipient who developed disseminated HSV with liver, colon, lung, and laryngeal involvement.[131]

3.3. Diagnosis of HSV infections in IBD

The gold-standard diagnostic test for tissue-invasive HSV disease, including colitis and hepatitis, is histopathologic evidence of herpes simplex virus infection, which can include characteristic eosinophilic intracytoplasmic inclusions (Fig. 2A and B) as well as positive immunohistochemical staining using labeled HSV-1- and
Published cases of herpes simplex virus (HSV) colitis in the English-language literature. Patients with inflammatory bowel disease (n = 4) in bold.

<table>
<thead>
<tr>
<th>Reference/year</th>
<th>Age/sex</th>
<th>Risk factor</th>
<th>HSV-1/HSV-2</th>
<th>Other organ involvement</th>
<th>Diagnostic test</th>
<th>Treatment</th>
<th>Outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td>Smith et al (2010)</td>
<td>50/M</td>
<td>CD, steroid therapy</td>
<td>HSV-2</td>
<td>None</td>
<td>Autopsy (histopathology, IHC) serology</td>
<td>Supportive</td>
<td>Died</td>
</tr>
<tr>
<td>Schunter et al (2007)</td>
<td>35/F</td>
<td>UC, steroid therapy</td>
<td>HSV-1</td>
<td>Perineal ulcers</td>
<td>Histopathology, viral culture (from rectum and perianal ulcer)</td>
<td>Supportive</td>
<td>Survived</td>
</tr>
<tr>
<td>Dray et al (2006)</td>
<td>69/F</td>
<td>CVID HSV-1</td>
<td>None</td>
<td>Viral culture (stool)</td>
<td>Acyclovir</td>
<td>Survived</td>
<td></td>
</tr>
<tr>
<td>Blaszyk et al (2006)</td>
<td>31/M</td>
<td>Liver transplant recipient</td>
<td>HSV-2</td>
<td>None</td>
<td>Histopathology, IHC, PCR</td>
<td>Acyclovir</td>
<td>Survived</td>
</tr>
<tr>
<td>Delis et al (2001)</td>
<td>2/M</td>
<td>Small bowel transplant recipient</td>
<td>HSV-2</td>
<td>None</td>
<td>Histopathology, PCR (colon biopsy)</td>
<td>Supportive</td>
<td>Died</td>
</tr>
<tr>
<td>Delis et al (2002)</td>
<td>28/F</td>
<td>Small bowel transplant recipient</td>
<td>HSV-1</td>
<td>Skin, liver, lung, colon</td>
<td>Histopathology, PCR (skin, lung, and colon)</td>
<td>Supportive</td>
<td>Died</td>
</tr>
<tr>
<td>Herget et al (2005)</td>
<td>22/F</td>
<td>Chemotherapy for acute leukemia</td>
<td>HSV-1</td>
<td>Skin, liver, lung, colon</td>
<td>Autopsy (histopathology, IHC)</td>
<td>Acyclovir</td>
<td>Died</td>
</tr>
<tr>
<td>Bissig et al (2003)</td>
<td>22/M</td>
<td>CD, steroid therapy</td>
<td>HSV-2</td>
<td>None</td>
<td>Autopsy (histopathology, IHC)</td>
<td>Acyclovir and foscarnet</td>
<td>Died</td>
</tr>
<tr>
<td>Norvell et al (2007)</td>
<td>28/F</td>
<td>UC, steroid therapy</td>
<td>HSV-1</td>
<td>None</td>
<td>Liver biopsy (histopathology, IHC)</td>
<td>Acyclovir</td>
<td>Survived</td>
</tr>
<tr>
<td>Levitsky et al (2008)</td>
<td>28/F</td>
<td>UC, steroid therapy</td>
<td>HSV-1</td>
<td>None</td>
<td>Liver biopsy (histopathology)</td>
<td>Acyclovir</td>
<td>Survived</td>
</tr>
<tr>
<td>Haag et al (2015)</td>
<td>19/F</td>
<td>CD, steroid therapy</td>
<td>HSV-2</td>
<td>None</td>
<td>Liver biopsy (HSV-1, viral PCR)</td>
<td>Acyclovir</td>
<td>Survived</td>
</tr>
</tbody>
</table>

HSV-2-specific monoclonal antibodies (Fig. 2C and D). In practice, particularly given the challenges of obtaining pathologic specimens from critically ill patients, a combination of noninvasive diagnostic tests is often used to support a clinical diagnosis of HSV end-organ disease and the decision to begin antiviral therapy.

A variety of noninvasive tests are available to aid in the diagnosis of tissue-invasive HSV infections.[54] Typically the most

Published cases of herpes simplex virus (HSV) hepatitis in patients with inflammatory bowel disease (IBD) in the English-language literature.

<table>
<thead>
<tr>
<th>Reference/year</th>
<th>Age/sex</th>
<th>Form of IBD</th>
<th>Immunosuppressive regimen</th>
<th>HSV-1/HSV-2</th>
<th>Other organ involvement</th>
<th>Diagnostic test</th>
<th>Treatment</th>
<th>Outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td>Shlien (1988)</td>
<td>16/F</td>
<td>UC</td>
<td>Steroids</td>
<td>HSV-1</td>
<td>Yes (oral &quot;aphthous&quot; ulcers, genital ulcers)</td>
<td>Autopsy (HSV-1)</td>
<td>Supportive</td>
<td>Died</td>
</tr>
<tr>
<td>Seksk et al (1999)</td>
<td>37/M</td>
<td>UC</td>
<td>Steroids</td>
<td>HSV-1</td>
<td>Yes (oral &quot;aphthous&quot; ulcers, genital ulcers)</td>
<td>Autopsy (HSV-1)</td>
<td>Acyclovir and foscarnet</td>
<td>Died</td>
</tr>
<tr>
<td>Bisig et al (2003)</td>
<td>22/M</td>
<td>CD</td>
<td>Steroids, tacrolimus, azathioprine</td>
<td>HSV-1</td>
<td>Yes (oral &quot;aphthous&quot; ulcers, genital ulcers)</td>
<td>Liver biopsy (histopathology, IHC)</td>
<td>Acyclovir</td>
<td>Survived</td>
</tr>
<tr>
<td>Norvell et al (2007)</td>
<td>28/F</td>
<td>UC</td>
<td>Steroids</td>
<td>HSV-1</td>
<td>Yes (oral &quot;aphthous&quot; ulcers, genital ulcers)</td>
<td>Liver biopsy (histopathology, IHC)</td>
<td>Acyclovir</td>
<td>Died</td>
</tr>
<tr>
<td>Haag et al (2015)</td>
<td>19/F</td>
<td>CD</td>
<td>Steroids, azathioprine, adalimumab</td>
<td>HSV-1</td>
<td>Yes (oral &quot;aphthous&quot; ulcers, genital ulcers)</td>
<td>Liver biopsy (HSV-1, viral PCR)</td>
<td>Acyclovir</td>
<td>Survived</td>
</tr>
</tbody>
</table>

CD = Crohn's disease, EM = electron microscopy, F = female, IHC = immunohistochemistry, M = male, NS = not specified, PCR = polymerase chain reaction, UC = ulcerative colitis.

This patient had also received a liver transplant.
readily available is HSV type-specific serology; however, given high background seroprevalence rates, a positive result is typically not useful for the diagnosis of suspected active disease, and a negative result does not exclude the possibility of a primary infection. As in our case, assays that detect IgM antibodies may suggest the diagnosis of early primary infection, especially if IgG antibodies are absent. Unfortunately, the sensitivity and specificity of these assays are limited by the fact that HSV IgM antibodies are not always detected, they can persist for months after primary infection, and they often reappear in the setting of reactivation.

Direct detection of HSV in non-invasively obtained specimens (e.g., scrapings of mucocutaneous lesions, respiratory or gastrointestinal tract secretions, etc.) using culture, direct immunofluorescence, or PCR-based assays can also support the diagnosis of end-organ involvement in the appropriate clinical setting. For instance, the appearance of characteristic herpetic mucocutaneous lesions in the context of mildly abnormal serum liver enzyme levels, transaminases and lactate dehydrogenase in particular—may herald impending fulminant HSV hepatitis and should prompt the initiation of timely antiviral therapy. However, concomitant skin findings are present in fewer than one-third of patients with HSV hepatitis, including those with biopsy. Similarly, less than half (n = 6) of all the previously reported cases of HSV colitis had suggestive mucocutaneous lesions at the time of presentation. More importantly, recovery of virus in the absence of lesions should be interpreted with caution since HSV reactivation and shedding is not uncommon in the context of critical illness and may or may not be indicative of active tissue-invasive HSV disease.

As illustrated by our case, visceral or disseminated HSV infections (particularly those with hepatic involvement) are frequently accompanied by some degree of HSV viremia, which can be detected using PCR assays of whole blood using HSV specific primers. Notably, in the United States none of these cases are currently approved by the Food and Drug Administration (FDA) for this application. Concomitant viremia was described in only one of the previously reported cases of HSV hepatitis in patients with IBD, and none of the cases of HSV colitis; however, HSV viremia has been used to predict hepatic involvement in other patient populations. Importantly, even the presence of HSV viremia is generally not sufficient to make a definitive diagnosis of visceral HSV disease because it too can be frequently detected in patients with non-HSV-related critical illness. Although quantification of viremia may be useful for differentiating between inconsequential reactivation and end-organ disease in this setting, this has not been studied in a prospective fashion.

### 3.4. Treatment and prevention of invasive HSV infections in IBD

There have been no prospective comparative studies of antiviral therapy for visceral or disseminated HSV infection. Acyclovir is the drug of choice for HSV infections and early treatment has been associated with improved outcomes for both HSV hepatitis and encephalitis. Primary mucocutaneous infections (i.e., occurring in a patient known to be HSV seronegative) in patients on immunomodulatory therapy for IBD should be treated promptly with antiviral therapy as this may prevent secondary complications. For tissue-invasive HSV infections (e.g., colitis, hepatitis, meningitis, etc.), most experts would recommend a minimum of 2 to 3 weeks of parenteral antiviral therapy. In the case of HSV hepatitis, a highly morbid disease with a high case fatality rate and no well-validated marker of disease activity, the duration may be significantly longer depending on the patient’s clinical course. After completion of a course of systemic antiviral therapy some experts advocate indefinite suppressive therapy with prophylactic doses of oral antiviral agents, particularly if further immunosuppressive therapy is anticipated in the future (i.e., post-transplantation), although this may lead to the subsequent emergence of acyclovir resistance. Our patient received a total of 6 weeks of antiviral therapy (intravenous acyclovir followed by oral valacyclovir) with a plan to continue lifelong suppressive therapy to prevent potential recurrence. Interestingly, four of the previously reported cases of HSV colitis were cured without specific antiviral therapy, though most of those patients did not have significant immunologic defects and ultimately required surgical resection of the diseased tissue before achieving symptom resolution.
In addition to initiating antiviral therapy, most experts as well as consensus guidelines from the European Crohn’s and Colitis Organization (ECCO) also recommend interruption of immunomodulatory therapy for IBD in the context of severe HSV infections.\(^{23,24,50}\) For more typical mucocutaneous infections immunomodulatory therapy may be continued, though there is a small risk for widespread dissemination and invasive disease.\(^{13,23,25}\) Patients with suspected hepatic involvement—particularly those with high-risk features, such as age >40 years, immunocompromised status, or evidence of fulminant hepatic failure (encephalopathy, coagulopathy, or thrombocytopenia)—should also be referred promptly for consideration for liver transplantation, as this may be life-saving.\(^{51}\)

Current guidelines make no formal recommendations regarding baseline serologic testing of patients with IBD for HSV or for routine antiviral prophylaxis, even when initiating immunomodulatory therapy.\(^{25,66}\) Given the observed increased rates of reactivation HSV disease—albeit mild—in patients receiving certain classes of immunomodulatory drugs, it has been suggested that patients with a previous history of frequent HSV outbreaks who are being prescribed immunomodulatory medications also receive suppressive antiviral therapy.\(^{65,67,68}\) though there are limited clinical data to support that recommendation. The likelihood of developing disseminated infection due to specific immunomodulatory agents has not been prospectively evaluated; however, based on post-licensure studies in other populations commonly prescribed similar therapies, the risk is likely to be greater with certain drug classes (Table 4). Consequently, it may be reasonable to offer secondary prophylaxis to HSV seropositive individuals or those with a history of recurrent localized mucocutaneous HSV infections who are going to receive prolonged courses of systemic corticosteroids, anti-metabolite therapy, or a calcineurin inhibitor. Patients with a previous history of disseminated or tissue-invasive HSV disease may be offered lifelong suppressive therapy/secondary prophylaxis, similar to the case patient, especially if any underlying or iatrogenic immunologic defect(s) cannot be reversed. However, the true risk of recurrence for tissue-invasive HSV disease has not been well established, secondary antiviral prophylaxis has variable efficacy depending on the site of disease, and ongoing antiviral therapy may lead to the development of drug resistance in immunocompromised patients.\(^{69}\)

4. Conclusion

Herpes simplex virus infections are common in patients with inflammatory bowel disease, but systemic dissemination and end-organ involvement have been infrequently reported. Patients with IBD are often at a higher risk for reactivation of HSV and tissue-invasive disease due to immunomodulatory therapy. Colitis and hepatitis due to HSV are particularly rare in this population, but HSV colitis should be considered in the differential diagnosis of refractory colitis symptoms, particularly if CMV disease has already been excluded. Treatment of suspected disseminated HSV infections should include prompt initiation of systemic antiviral therapy, with routine involvement of organ-specific and infectious disease specialists. Strategies for prevention of these infections include symptom-based and serologic screening of patients that are going to receive immunomodulatory therapy, especially with systemic corticosteroids or antimetabolites, with consideration of prophylactic antiviral therapy in those with a history of recurrent or complicated HSV infections.

References


