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Presentation of hepatocellular antigens

Arash Grakoui1 and Ian Nicholas Crispe2

The liver is an organ in which antigen-specific T-cell responses manifest a bias toward immune tolerance. This is clearly seen in the rejection of allogeneic liver transplants, and multiple other phenomena suggest that this effect is more general. These include tolerance toward antigens introduced via the portal vein, immune failure to several hepatotropic viruses, the lack of natural liver-stage immunity to malaria parasites, and the frequent metastasis of cancers to the liver. Here we review the mechanisms by which T cells engage with hepatocellular antigens, the context in which such encounters occur, and the mechanisms that act to suppress a full T-cell response. While many mechanisms play a role, we will argue that two important processes are the constraints on the cross-presentation of hepatocellular antigens, and the induction of negative feedback inhibition driven by interferons. The constant exposure of the liver to microbial products from the intestine may drive innate immunity, rendering the local environment unfavorable for specific T-cell responses through this mechanism. Nevertheless, tolerance toward hepatocellular antigens is not monolithic and under specific circumstances allows both effective immunity and immunopathology.

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According to ancient Babylonians and in the practice of Mesopotamian medicine, the liver is metaphorically regarded as the ‘seat of the living soul’ (The Evolution of Modern Medicine, William Osler). Ritualistic examination of the liver comprising assessment of its size and position, its color and the richness of its blood, was routinely interpreted as a measure of the inner invisible characteristics including emotions as well as illnesses. So while in the modern English language, the conceptualization of abstract emotion over the years has been linked to the heart, in ancient Babylonia, it all started with the liver. Pablo Neruda’s magnificent ‘Ode to the Liver’ is a modern manifestation of these ancient insights. Today, we know that the liver is the major organ responsible for more than 500 different functions; protein, carbohydrate and lipid metabolism, hormone production, plasma protein synthesis, decomposition of red blood cells, glycogen storage, bile production and detoxification to name a few – symbolically functioning as the organ responsible for cleansing of the soul.

Anatomically, it is interesting and unique that besides being the largest internal organ, the liver is traversed by both the hepatic artery and the portal vein, with the former carrying oxygen-rich blood from the aorta and the latter bringing in myriad antigens from the gut, spleen and pancreas and nutrients from the gastrointestinal tract. The hepatic blood exits from the sinusoids into the central vein where it is drained into the inferior vena cava. Perhaps in this antigen-rich context, it is not surprising that evolutionarily the liver has developed a tolerogenic environment in order to manage the panoply of antigens and their neo-antigenic metabolites. For tissue preservation, the liver must reduce the risk of immune activation in response to various oral and self-antigens. Importantly, the large microbial biomass found in the gut that serves as an important determinant of intestinal inflammation also directly influences the development of liver disease.1 In fact, the hyporesponsiveness to an oral antigen can be significantly reversed with the creation of a portal shunt.2 This tolerogenic aspect of the liver is consistent with the observation that liver allografts, unlike heart or kidney, can be accepted in various animal and human transplant settings in the absence of additional immunosuppression.3,4 Co-transplantation with the liver allows for protection and acceptance of other organs that normally would be rejected.5

Hepatic microvessels, known as sinusoids, slow down the blood flow and allow for the liver’s biochemical functions to be enacted by specialized cells, the hepatocytes. Hepatocytes comprise ~60% of hepatic cells and close to 90% of the liver volume. Liver sinusoidal endothelial cells (LSECs) form a fenestrated barrier separating the hepatocytes from direct blood

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flow and the immune cells that reside in the lumen. How does the liver balance between the need for tolerance to antigens that are metabolized in the liver and the requirement for immunity to pathogens that have developed hepatotropic propensity over the years? Are gut microbial antigens and/or neo-antigens derived from metabolic byproducts providing danger signals to induce inflammation? In addition to hepatocytes themselves controlling many aspects of inflammation, the assistance by other intrahepatic cells collaborate in managing the overall functioning of the liver. These nonparenchymal cells include lymphocytes, Kupffer cells, natural killer (NK) cells, NK-T cells, and stellate and dendritic cells (DC) that are assigned with sensing any perturbation in the liver architecture and milieu (Figure 1). The subject of this review is to delineate how immune sensing in the liver is shaped by the location, where the antigen is processed, how it is processed and to explore the influence of host innate receptors and pathogen subversion tactics as they act on hepatocytes and nonparenchymal cells in liver immunity.

**ANATOMY OF ANTIGEN PRESENTATION**

The presentation of antigens is controlled both by their location within the cell and by accessory signals that in turn are controlled through innate immune recognition mechanisms that sense pathogen-associated molecular patterns (PAMPs) such as viral and microbial nucleic acid motifs and distinctive glycolipids and specific conserved microbial proteins. A largely overlapping set of receptors activates similar signaling pathways in response to cellular injury through the recognition of damage-associated molecular patterns (DAMPs). Some of these receptors reside on the plasma membrane, for example Toll-like Receptor (TLR) 2 and TLR4, which recognize components of bacterial cell walls; some reside in endosomes, such as TLR3 and TLR9 that sense structural features of viral and microbial RNA and DNA respectively; and some are cytoplasmic, such as the retinoic acid-inducible–like receptors that sense viral RNA and the Nod-like receptors that sense bacterial lipids. Thus, in the liver, immune cell activation can be influenced by the cell in which the antigen is expressed, the cellular compartments in which it exists, the presence of innate immune signals that modulate antigen presentation via PAMP and DAMP receptors, and immune subversion mechanisms evolved by the pathogen to disable host defense.

Presentation of antigen to both CD4+ and CD8+ T cells depends on antigen processing, which is distinct and partitioned in different cellular compartments. Thus, viral capsid proteins and proteins that are contained within the virion can gain access to endosomes and to the processing pathway that degrades antigens and loads their peptides into major histocompatibility complex (MHC) class II molecules in a specialized compartment termed the MHC class II compartment. This route of processing is also open to virally encoded proteins that are secreted from infected cells, such as hepatitis B surface antigen in the case of hepatitis B infection. In contrast, non-structural viral proteins are synthesized only in the infected cell, and most often in the cytoplasm. These proteins can be degraded directly by the proteasome and loaded to the MHC class I molecules via a protein transporter complex, transporter of antigenic peptides, that traffics them into membrane-bound compartments. The antigens of nonviral pathogens are also located in both membrane-bound and cytoplasmic compartments. Thus, the bacterium *Listeria monocytogenes* is first taken up by endocytosis, but then disrupts the membrane of the endosome through the action of an enzyme listeriolysin, and enters the cytoplasm where it both migrates and spreads from cell to cell by exploiting the host cell cytoskeleton. Therefore, this pathogen evades the classic MHC class II pathway but *Listeria*-encoded antigens are available for the classical MHC class I-processing pathway. While *Listeria* is a virulent pathogen that infects the liver, its intracellular location together with its strong activation of innate immunity conspire to render it, in healthy individuals, a potential vaccine vehicle. In fact vaccines based on attenuated *Listeria* organisms can induce effective anticancer immunity, making them an exciting avenue for vaccinology and immunotherapy.

Malaria parasites enter hepatocytes by direct invasion of the bloodstream, which appears to be mediated by a prior interaction with Kupffer cells. Thus, their antigens expressed by the invasive stage, the sporozoite, are potentially accessible to the classical MHC class I pathway, but the parasites induce the formation of a parasitophorous vacuole, the membrane of which contains both host-encoded and parasite-encoded proteins. This vacuolar membrane mediates interaction between the parasite and the infected hepatocyte, but the extent

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**Figure 1** Anatomy of antigen presentation in the sinusoid. Plates of hepatocytes are separated from the blood flowing in the sinusoids by liver sinusoidal endothelial cells. In the blood space are recirculating natural killer cells, natural killer T cells, monocytes–macrophages and, adherent to the endothelial wall, the Kupffer cells. To interact directly with hepatocytes, CD8+ T cells must cross the endothelial barrier and enter the space of Disse. CD4+ T cells also enter this space but cannot directly interact with hepatocytes. However, they find other interaction partners such as DCs and stellate cells.
to which it controls antigen presentation is not understood. Genetically attenuated malaria parasites that can function as live vaccines may undergo developmental arrest before they form a parasitophorous vacuole; but late-arresting parasite variants that undergo partial differentiation within such a vacuole may also induce sterilizing immunity.14,15

**Licensing the Antigen-Presenting Cells**

Many important liver pathogens infect primarily hepatocytes. These include the hepatitis viruses (hepatitis A virus [HAV], hepatitis B virus [HBV], hepatitis C virus [HCV] and other less common viruses), cytomegalovirus and the globally important malaria parasite. Since these are intracellular pathogens, host defense depends primarily on T cells and in all these infections there is strong evidence that the cytotoxic CD8+ T cells are essential for host defense. To take two among many examples: depletion of CD8+ T cells from HBV-infected chimpanzees results in a resurgence of viremia16 and similarly abrogates immunity in mice primed with radiation-attenuated malaria parasites.17,18 Ones fully activated, cytotoxic CD8+ T cells undergo clonal expansion and may deliver their defensive function without support from other cell types, but for efficient primary activation, full effector function, survival and memory CD8+ T cells and the delivery of memory effector function, CD8+ T cells depend on an interaction with CD4+ T cells termed ‘help’. This interaction is mediated in several ways: through the direct delivery of supportive CD4+ T-cell-derived cytokines such as interleukin-2 (IL-2); through the enhanced function of specialized antigen-presenting cells (APCs) such as DCs, a mechanism termed licensing; and through a direct interaction between CD4+ and CD8+ T cells that requires the expression of CD40 on the CD8+ T cells.22

Among these mechanisms, ‘licensing’ is the most efficient because it can be mediated by sequential interaction of a rare antigen-specific CD4+ T cell and subsequently a rare antigen-specific CD8+ T cell with an APC. Both the licensing interaction between the CD4+ T cell and the APC, and the licensed interaction between the APC and the CD8+ T cell, depend on MHC-restricted antigen recognition, and this in turn means that for licensing to occur, the APC must express both MHC class I and class II. Among potential liver-resident APC, trafficking DC express both classes of MHC molecules. At a lower level, so do Kupffer cells and LSECs, but hepatocytes only express MHC class I. Therefore, hepatocytes cannot be licensed by CD4+ T cells.23 Instead, the full activation of a CD8+ T cell specific for a hepatocellular antigen depends on cross-presentation by an MHC class I+II+ cell (Figure 2).

Among liver-resident cells, the large macrophage population termed Kupffer cells would be an obvious candidate for the cross-presentation of hepatocellular antigens. However, the balance of evidence suggests that Kupffer cells are immunosuppressive. Interaction of Kupffer cells with CD8+ T cells in vitro causes proliferation,24,25 but they also secrete both IL-1024,25 and the immunosuppressive prostaglandin PGE2,26,27 and in vivo depletion of Kupffer cells impairs both oral tolerance28 and liver transplantation tolerance.29 It follows that there is no conflict between the abundance of Kupffer cells in HCV-infected livers and the persistence of HCV infection. If Kupffer cells were to be presenting or cross-presenting HCV-encoded antigens, tolerance would be the expected outcome.

The other liver-resident cell type that constitutively expresses MHC class II is the LSEC. These cells are very active in pinocytosis, sequester virions from the circulation and could readily cross-present both circulating antigens and hepatocellular antigens, whether these are released as soluble protein or in exosomes.2 Exosomes are membrane-bound cell fragments that transport proteins from one cell to another, and they have been implicated in the transfer of HCV-encoded proteins and viral RNA.33 LSECs may also cross-present cancer-derived antigens.34 However, the default outcome in the case of antigen presentation by LSECs is immune tolerance.34–37

These considerations lead to the conclusion that the potential of liver-resident cells to cross-present hepatocellular antigen does not lead to effective immunity. Instead, both LSECs and Kupffer cells may cross-present antigens, but the outcome is not ‘help’ but immune suppression. This raises a fundamental biological question: in the context of liver infection, are the LSECs and the Kupffer cells resistant to licensing mechanisms that act in other contexts? Or are they licensed, but in an alternative way that enhances their immunosuppressive potential? Experiments with purified LSECs and Kupffer cells in vitro have not so far clarified this issue, because there is no way to determine whether the cells were licensed, or unlicensed (or delicensed, or alternatively licensed) already at the time of isolation.
TOLERANCE DESPITE THE PRESENCE OF DANGER SIGNALS

Adaptive immune responses to pathogens occur in the context of ongoing innate immunity. The ‘Danger Model’ asserts that adaptive immunity is triggered by tissue damage, and the current understanding of this process is that injured cells release DAMPs that engage the same classes of receptors, as do PAMPs. These receptors engage two major signaling cascades: one via the adapter molecule MyD88 that activates many innate immune genes including those encoding Tumor Necrosis Factor (TNF)-α and IL-1α/β; the other via TRIF and IRF7 activates Interferon (IFN)-α/β, and thence a large number of interferon-sensitive genes, many of which encode antiviral proteins. Both DAMP and PAMP receptors are abundantly expressed by liver cells, so it would be expected that the activation of innate immunity is effective in the liver. The ‘Danger Model’ asserts further that these changes set the scene for adaptive T-cell immunity through increased expression of MHC class I and class II molecules, increased expression of co-stimulatory and intercellular adhesion molecules, and the switch from basal proteasomes to immunoproteasomes. All these changes can be driven by IFNs.

However, in the liver there are two confounding factors that derail this logically appealing model. First, the liver is exposed to low levels of bacterial products from the intestinal microbiota, and these include endotoxin (lipopolysaccharide, LPS) from gram-negative bacteria. While the level of LPS in the liver is increased in pathological states including alcohol toxicity, it is also detectable at baseline. Low-level exposure to LPS could act by inducing LPS tolerance, which is illustrated by experiments in which systemic administration of a low dose of LPS results in protection from a subsequent challenge with a lethal dose by modifying the immunobiology of liver cells. This process acts by down-regulating innate immune signaling pathways (Figure 3). Thus, the presence of low-level LPS in the liver could blunt the innate response to infection. In addition, documented effects of LPS on isolated liver cells suggest it may induce immunosuppressive molecules. Thus, exposure of Kupffer cells to low-dose LPS resulted in the secretion of IL-10, which in turn could modify the function of LSECs. In addition, liver APCs are not functionally mature in the same way as their counterparts in other organs, as identical treatments result in divergent immune responses. This could be due to the presence of a large quantity of anti-inflammatory cytokines in the liver or cell intrinsic properties that make these hepatic APCs less immunostimulatory.

A second consideration is the induction of negative feedback through IFNs. Innate immunity results in the secretion of both IFN-α/β by many cell types, and IFN-γ mostly by lymphocytes including NK cells and NK-T cells. While these IFNs induce MHC I/II, co-stimulation and the immunoproteasome activation, they also induce negative feedback inhibition that may result in immune tolerance. These effects are best documented for IFN-α/β, which suppressed the CD4+ T-cell response to blood-stage malaria parasites, while mice lacking IFN-α/β signaling were relatively resistant. Similarly, IFN-α/β suppressed the CD8+ T-cell response in virus infection. In LCMV infection in mice, inhibition of IFN-α/β caused increased IFN-γ secretion and an antiviral effect. However, IFN-γ may also suppress effective immunity, for example in pancreatic islet transplantation into the liver. This type of immune suppression induced by proinflammatory cytokines in the context of cancer has been termed adaptive resistance. Therefore, we would argue that immune tolerance provoked by inflammation should be called adaptive tolerance.

Suppression of T-cell immunity acts via many pathways, including immunosuppressive small molecules, cytokines and cell surface ligands; many of these are induced by IFNs. IFN-β induces the enzyme IDO1, which imposes T-cell tolerance through the depletion of tryptophan and the synthesis of kynurenine. IFN-γ gene transduction also induces the Ido1 gene as well as both Fas (CD95) and Fas ligand, the interaction of which results in T-cell apoptosis. Some other proinflammatory cytokines have similar effects. Thus, IFN-γ along with IL-12 and IL-17 up-regulated the immunosuppressive ligand PD-L1; IFN-γ along with TNF-α and TLR2, 3 and 4 ligands induced galectin-9.

There is evidence that all these immunosuppressive pathways are active in the liver. Thus, in HBV infection, IDO not only suppresses anti-hepatitis B surface antigen CD8+ T-cell activity but also promotes liver injury in fulminant hepatitis. In mice, deficiency of PD-L1 results in massive accumulation of CD8+ T cells in the liver, while antibody-mediated inhibition of PD-L1 signaling enhanced an anti-HBV CD8+ T-cell response. Similarly, in human HCV and HBV infection, dysfunctional T cells were enhanced by PD-L1 blockade. Likewise, both the Fas–FasL pathway and the galectin-9–Tim-3 pathway regulate T-cell immunity in viral hepatitis.

The IFN-induced immunosuppressive feedback pathways may actively promote T-reg activity. Thus, in HCV the interaction of galectin-9 with Tim-3 on CD4+ T cells promoted T-reg
correlated with the onset of viral clearance, while CD8 function only improved after virus was eliminated. This is a provocative observation that challenges the model, argued recently evolved to include inflammation as a mechanism. Although the first landmark observation linking liver disease and viral infection was made more than 40 years ago, our understanding of liver disease processes has only recently evolved to include inflammation as a mechanism rather than a consequence. Liver lymphocyte infiltration, increased levels of circulating LPS and elevated levels of pro-inflammatory cytokines such as TNF-α are key hallmarks of not just alcoholic liver disease but also nonalcoholic steato-hepatitis, and cirrhosis due to viral infection. Consistent with this idea, a recent study by one of our laboratories showed surprisingly that liver recruitment and activation of DCs was a general consequence of inflammation, irrespective of viral infection status. In fact, the immune manifestations of alcoholic liver disease and nonalcoholic steato-hepatitis are virtually indistinguishable through current diagnostic methods, suggesting that common core pathways of inflammation-fueled cirrhosis exist that may serve useful as therapeutic targets.

Recently, studies have alluded to the contribution of intestinal microbiota as prime suspects to promote metabolic diseases by driving low-level inflammation. In fact TLR/Nod-like receptors activation in resident Kupffer cells by low-level bacterial products (endotoxin) in the liver through TLR4 pathway is critical for liver inflammation induced by ischemia reperfusion, alcohol and viral components. Given the alarming correlation seen between changes in our diet to obesity, metabolic syndrome and nonalcoholic steato-hepatitis, gut permeability and elevated serum endotoxins have been shown to be critical components in promoting liver inflammation and disease progression. Interestingly, many of the diet-induced metabolic manifestations seem to be reversed in germ-free mice and/or in mice devoid of TLR signaling. In addition, gut microbiota also play an important role in the progression of liver disease to fibrosis as chemically induced treated mice with carbon tetrachloride showed increases in bacterial translocation. Furthermore, intestinal microbiota and signaling through TLR4 on hepatic stellate cells was shown to be critical for the development of fibrosis by modulation of TGFβ and subsequent unrestricted activation of Kupffer cells.

In summary, the liver is an environment where antigens are readily presented by diverse liver cell types, but the prevalence of background low-level exposure to innate immune stimuli results in continuous feedback inhibition of T-cell immunity through many parallel pathways. However, the inextricable connectivity between the intestinal microbiota and the liver also selects for an inflammatory thermostat that shifts the balance from tolerance to inflammatory-induced liver disease when gut microbial antigens and/or neo-antigens provide unchecked danger signals to the liver. In Neruda’s words (translated by H. Morales and W. Hochman) ‘one small cell goes astray, the pilot flies in the wrong sky, the tenor shrinks to a whisper, the astronomer loses his planet’. Such is the central role of the liver in immunity.

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5 Labaied M, Harupa A, Dumpit RF, Coppens I, Mikolajczak SA, Kappe SH. Genetically engineered, attenuated *Salmonella* vaccine vectors that express different molecular forms of human papilloma virus-16 (HPV-16) E7 induce qualitatively different liver antigens


