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Journal Title: Journal of Virology
Volume: Volume 89, Number 18
Publisher: American Society for Microbiology | 2015-09-01, Pages 9368-9382
Type of Work: Article | Final Publisher PDF
Publisher DOI: 10.1128/JVI.01178-15
Permanent URL: https://pid.emory.edu/ark:/25593/rmzk7

Final published version: http://dx.doi.org/10.1128/JVI.01178-15

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Accessed January 14, 2018 9:50 PM EST
P2X1 Receptor Antagonists Inhibit HIV-1 Fusion by Blocking Virus-Coreceptor Interactions

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ABSTRACT

HIV-1 Env glycoprotein-mediated fusion is initiated upon sequential binding of Env to CD4 and the coreceptor CXCR4 or CCR5. Whereas these interactions are thought to be necessary and sufficient to promote HIV-1 fusion, other host factors can modulate this process. Previous studies reported potent inhibition of HIV-1 fusion by selective P2X1 receptor antagonists, including NF279, and suggested that these receptors play a role in HIV-1 entry. Here we investigated the mechanism of antiviral activity of NF279 and found that this compound does not inhibit HIV-1 fusion by preventing the activation of P2X1 channels but effectively blocks the binding of the virus to CXCR4 or CCR5. The notion of an off-target effect of NF279 on HIV-1 fusion is supported by the lack of detectable expression of P2X1 receptors in cells used in fusion experiments and by the fact that the addition of ATP or the enzymatic depletion of ATP in culture medium does not modulate viral fusion. Importantly, NF279 fails to inhibit HIV-1 fusion with cell lines and primary macrophages when added at an intermediate stage downstream of Env-CD4-coreceptor engagement. Conversely, in the presence of NF279, HIV-1 fusion is arrested downstream of CD4 binding but prior to coreceptor engagement. NF279 also antagonizes the signaling function of CCR5, CXCR4, and another chemokine receptor, as evidenced by the suppression of calcium responses elicited by specific ligands and by recombinant gp120. Collectively, our results demonstrate that NF279 is a dual HIV-1 coreceptor inhibitor that interferes with the functional engagement of CCR5 and CXCR4 by Env.

IMPORTANCE

Inhibition of P2X receptor activity suppresses HIV-1 fusion and replication, suggesting that P2X signaling is involved in HIV-1 entry. However, mechanistic experiments conducted in this study imply that P2X1 receptor is not expressed in target cells or involved in viral fusion. Instead, we found that inhibition of HIV-1 fusion by a specific P2X1 receptor antagonist, NF279, is due to the blocking of virus interactions with both the CXCR4 and CCR5 coreceptors. The ability of NF279 to abrogate cellular calcium signaling induced by the respective chemokines showed that this compound acts as a dual-coreceptor antagonist. P2X1 receptor antagonists could thus represent a new class of dual-coreceptor inhibitors with a structure and a mechanism of action that are distinct from those of known HIV-1 coreceptor antagonists.
cells stably expressing DSP1 (293T-DSP1 cells) (28) were gifts from H. Hoshino, N. Hosoya, and A. Iwamoto (University of Tokyo, Tokyo, Japan). NP2 cells were grown in minimal essential medium (MEM) (Cellgro) supplemented with 10% FBS, 100 U/ml penicillin-streptomycin, and 4 μg/ml blasticidin (Bioworld, Atlanta, GA).

**Primary macrophage culture.** Peripheral blood was obtained from healthy volunteer donors according to a protocol approved by the Emory University Institutional Review Board (IRB). Peripheral blood mononuclear cells (PBMCs) were obtained from the buffy coat after Ficoll density centrifugation and were negatively selected by using magnetically activated cell sorting (MACS) monocyte isolation kit II (Miltenyi Biotec, Bergish Gladbach, Germany). Monocytes were differentiated into macrophages upon culturing for 7 days in RPMI 1640 medium supplemented with 10% FBS, 100 U/ml penicillin-streptomycin, 2 mM glutamine, 1% sodium pyruvate, 1% nonessential amino acids, and 5 μg/ml granulocyte-macrophage colony-stimulating factor (GM-CSF; Cell Sciences, Canton, MA, USA).

**Plasmids.** The pCAGGS-HXB2-Env vector for X4-tropic envelope expression was provided by J. Binley (Torrey Pines Institute, CA), the pBal26-Env vector for R5-tropic envelope expression was a gift from P. Clapham (University of Massachusetts, Worcester, MA), and pHPG-R3A-Env was a gift from J. Hoxie (University of Pennsylvania). The pMDG-VSVG plasmid expressing VSV-G was a gift from J. Young (Salk Ins, La Jolla, CA). The HIV-1 proviral clone pRΔEnv lacking the env gene was obtained from D. Trono (University of Geneva, Geneva, Switzerland), and pcRev and PMM310-Blam-Vpr expressing Rev and β-lactamase (Blam-Vpr), respectively, were obtained from the ARRRP (29). The pcDNA3.1-P2X1R-WT and pcDNA3.1-P2X1R-dell. (l351-deleted) plasmids encoding wild-type human P2X1R and its dominant-negative mutant, respectively, were kindly provided by M. Hoylaerts (University of Leuven, Leuven, Belgium) (30).

**Reagents and inhibitors.** The P2X1R inhibitors NF279 and NF449, the P2X7R inhibitor NF449, and thapsigargin were obtained from Tocris Bioscience (R&D Systems, Minneapolis, MN). ATP, oxidized ATP (oATP), α,β-methylene ATP (α,β-m-eATP), arsate from potato, the CXCR4 antagonist AMD3100, the calcium ionophore A23187, and poly-I-lysine were obtained from Sigma-Aldrich (St. Louis, MO). The calcium ionophore ionomycin was obtained from Calbiochem (EMD Millipore, Billerica, MA). The cell-permeable calcium indicator Fluor-AM was obtained from Invitrogen. The BMS-806 compound was synthesized by Chemical Corp. (Baltimore, MD). The C52L recombinant peptide was a kind gift from Min Lu (University of New Jersey). The human chemokine stromal cell-derived factor 1 alpha (SDF-1α) was obtained from Cell Sciences (Canton, MA), the synthesized CCR5 receptor agonist RANTES (regulated on activation, normal T cell) was provided by Dr. M. Grotz (Helmholtz Zentrum Muenchen, Germany), and the CCR5 antagonist TAK-779 (31) were obtained from the ARRRP. The concentration of ATPr was quantified by a luminescence assay, using the commercial ATP-Plite kit (Perkin-Elmer) according to the manufacturer’s recommendations. Luminescence was measured by using a TopCount NXT plate reader, and ATP concentrations of samples were determined by comparison with ATP standards.

**Virus production.** Pseudoviruses containing Blam-Vpr were produced as described previously (10). Briefly, a 10-cm dish of HEK 293T/17 cells was transfected with JetPrime reagent (Polyplus Transfection, Illkirch-Graffenstaden, France) by using the following plasmids: the viral envelope glycoprotein vector (3 μg), pRΔEnv (2 μg), pMM130-Blam-Vpr (2 μg), and pcRev (1 μg). Twelve hours after transfection, the medium was replaced with phenol red-free DMEM growth medium, and pseudoviruses were harvested 48 h after transfection. The supernatant was centrifuged for 5 min at 350 × g, filtered through a 0.45-μm filter membrane, and concentrated by using the LentX reagent (Clontech Labora-
tories, Madison, WI) according to the manufacturer’s recommendations. Pseudoviruses were then aliquoted and stored at −80°C. Infectious titers were determined by a β-galactosidase assay in TZM-bl cells (32).

**Results**

**P2X1 receptor antagonists, but not P2X7 or P2Y11 receptor antagonists, inhibit HIV-1 fusion.** We first confirmed and expanded previously reported findings that P2X1R inhibitors selectively block HIV-1 fusion (13–15, 24). The effects of antagonists of P2X1 (NF279 and NF449), P2Y11 (NF340), and P2X7 (A740003) receptors on HIV-1 fusion with HeLa-derived and CD4+ T cells were tested. In agreement with our previously reported findings (24), the P2X1R antagonists NF279 and NF449 suppressed fusion between particles pseudotyped with CCR5-tropic (HXB2), CCR5-tropic (Bal26), and dual-tropic (R3A) HIV-1 Env glycoproteins and TZM-bl cells in a dose-dependent manner (Fig. 1A to C). Both compounds were nearly equally potent and were more effective against fusion mediated by laboratory-adapted HXB2 Env than against fusion mediated by Bal26 or R3A Env. We have previously shown that this inhibition was not due to downregulation of CD4 or coreceptor expression (24). Both NF279 and NF449 inhibited HXB2 pseudovirus (HXB2pp) fusion with lymphoid CEM-A cells (Fig. 1E). However, high concentrations of NF279 were required to block fusion with CEM-A cells compared to TZM-bl cells, and NF449 was considerably less potent than NF279. NF279 also inhibited HXB2pp and Bal26pp fusion with CEM-A and Jurkat cells engineered to express CCR5 (data not shown).

In contrast to P2X1R antagonists and consistent with our previously reported results (24), the P2X7 and P2Y11 receptor antagonists A740003 and NF340 exhibited marginal to no inhibitory activity against HIV-1 fusion with TZM-bl or CEM-A cells (Fig. 1). Importantly, none of the tested P2X7R or P2Y11R antagonists considerably affected VSVpp fusion (Fig. 1D) or the viability of TZM-bl and CEM-A cells (Fig. 1A and E). These results appear to further support a specific role for P2X1R in HIV-1 entry/fusion.

**Extracellular ATP or elevated cytosolic calcium levels are not required for HIV-1 fusion.** P2X receptors are activated upon the release of cellular ATP (e.g., see reference 34). We therefore asked whether depletion of extracellular ATP, presumably released by cells upon HIV binding (13, 14), or addition of exogenous ATP could modulate HIV-1 fusion. To address this question, virus-cell fusion experiments were carried out in the presence of a high dose of the ATP-degrading enzyme apyrase. Apyrase did not considerably affect HXB2pp or Bal26pp fusion with TZM-bl cells (Fig. 2A) at a concentration that effectively degraded 1 mM ATP within 30 min at 37°C (Fig. 2A and B). The addition of 0.5 μM ATP, a concentration that is known to activate P2X1R (35), did not augment virus-cell fusion. A relatively low multiplicity of infection
was used in these experiments in order to avoid saturation of the BlaM signal. Even 100 μM ATP did not considerably facilitate HIV-1 fusion (data not shown). Likewise, neither apyrase nor exogenous ATP modulated HXB2pp fusion with CEM-A cells (Fig. 2C). The stable ATP analogs α,β-meATP, which is thought to selectively activate P2X1R (35), and oATP, a general inhibitor of purinergic receptors, did not noticeably affect fusion with either cell type. Whereas BaL26pp fusion was modestly promoted by ATP and α,β-meATP, this effect was also observed in samples treated with apyrase (Fig. 2A), suggesting that the modest en-
hancement of fusion in the presence of these reagents was not related to P2X1R activation.

Since much lower concentrations of ATP are required to activate P2X1R than P2X4R and P2X7R (35), the above-described results do not rule out the possibility that very low doses of ATP released locally through pannexin-1 hemichannels (36, 37) activate P2X1R. We therefore asked if downstream effects of P2X1R activation, specifically the elevation of the cytosolic calcium concentration, can modulate HIV-1 fusion. The calcium concentration in the cytoplasm during HIV-1 entry/fusion was increased by adding the calcium ionophore ionomycin or the sarco/endoplasmic reticulum calcium

FIG 2 Extracellular ATP or an elevated cytosolic calcium concentration is not required for HIV-1 fusion. (A) Fusion of HXB2pp or BaL26pp with TZM-bl cells was allowed to proceed in the absence or in the presence of 10 U/ml of apyrase, 0.5 μM ATP or 100 μM α,β-methyleneATP or oATP, 2 μM thapsigargin, 10 μM ionomycin, and 30 μM NF279. (B) Control for apyrase activity. ATP (1 mM) was incubated with 10 U/ml apyrase for 30 min at 37°C. The concentrations of ATP in mock-treated and apyrase-treated samples were quantified by using an ATPlite kit. (C) HXB2pp fusion with CEM-A cells under the conditions described above for panel A. Data are mean normalized BlaM activities (± standard deviations) from three independent experiments carried out in triplicate. (D) Cell viability measured in the same samples by an MTS assay. (E) HIV-1 Env-mediated cell-cell fusion assay. 293T-DSP1 cells transfected with HXB2 Env were cocultured with target N4X4-DSP2 cells in the absence or presence of 30 μM NF449 or NF340, 0.5 μM ATP, 100 μM oATP, 10 U/ml apyrase, or 7 μM C52L for 2 h at 37°C. Data points are the normalized mean luminescence values and standard deviations from three independent experiments carried out in triplicate. *, P < 0.05; **, P < 0.01; ***, P < 0.001.
ATPase (SERCA) inhibitor thapsigargin. Thapsigargin did not significantly augment HXB2pp or BaL26pp fusion with TZM-bl or CEM-A cells, while ionomycin actually tended to inhibit fusion (Fig. 2A and C). Neither of the above-described conditions, including the presence of ionomycin or thapsigargin, affected cell viability (Fig. 2D). These results imply that elevation of the cytosolic calcium concentration alone does not promote HIV–cell fusion.

We also tested the effects of the above-described agents/inhibitors on HIV-1 Env-mediated cell-cell fusion, using a dual-split-protein assay (38, 39). In this assay, fusion is detected based upon de novo assembly of functional GFP and Renilla luciferase proteins from their respective complementary domains (designated DSP1 and DSP2) expressed separately in fusing partners. A robust luciferase signal was detected after coculture of HEK 293T cells stably expressing DSP1 that were transfected with HXB2 Env and target N4X4–DSP2 cells expressing DSP2 and CD4/CXCR4. NF279 effectively inhibited cell-cell fusion, while another P2X1R antagonist (NF449) and another P2Y11 antagonist (NF340) were less potent (Fig. 2E). Similar to virus-cell fusion, apyrase did not considerably affect the extent of cell-cell fusion, while oATP exhibited a mild deleterious effect. Our results thus show that P2X1R antagonists selectively inhibit HIV-cell and Env-mediated cell-cell fusion. However, NF279 is universally potent in both fusion assays involving different cell types, whereas NF449 exhibits variable potency. The different potencies of these two selective P2X1R antagonists against HIV-1 fusion raise concerns regarding a possible off-target effect of NF279.

Functional P2X receptors are present in epithelial but not lymphoid cell lines. The insensitivity of HIV-1 fusion to exogenous ATP (Fig. 2) prompted us to examine the expression of the P2X1 receptor in TZM-bl, CEM-A, or Jurkat cells by Western blotting. None of these cell lines exhibited a detectable band corresponding to this receptor (Fig. 3A), but a robust P2X1R signal was present after ectopic expression of this receptor or its truncated dominant negative mutant (30) in HEK 293T or TZM-bl cells. The lack of P2X1R expression in epithelial cells is in agreement with data from previous reports (16, 34, 40). The undetectable levels of this receptor in lymphoid cells are somewhat surprising, since recent studies reported P2X1R expression in Jurkat and primary CD4+ T cells (14, 41). It is possible that the subclones of Jurkat and CEM-A cells used in our fusion experiments expressed exceptionally low levels of this receptor.

We next asked if the cells used in the above-described fusion experiments express any functional P2X receptors by testing the ability of ATP to induce calcium signaling. Changes in intracellular calcium concentrations were monitored by imaging of cells loaded with the calcium indicator Fluo4-AM, as described in Materials and Methods. Exogenously added ATP elicited strong calcium signaling in TZM-bl and HEK 293T cells (Fig. 3B and C). Surprisingly, even very high (millimolar) concentrations of ATP did not elicit detectable responses in CEM-A or Jurkat cells (Fig. 3D and E). The lack of calcium signaling indicates that functional P2X1 receptors are not expressed on the lymphoid cell lines used in our experiments and thus argues against the involvement of these channels in HIV-1 fusion. T cells have been reported to respond to very high concentrations of exogenous ATP by increasing the cytosolic calcium levels (41, 42), but these responses likely involve P2X7 channels.

Collectively, the undetectable levels of P2X1R in epithelial and lymphoid cells and the lack of ATP-mediated signaling in lymphoid cells indicate that functional P2X1R is not required for HIV-1 fusion. These findings suggest that NF279 affects a different cellular target(s) at concentrations used to inhibit fusion.

P2X1R antagonists prevent functional engagement of CXCR4 and CCR5 coreceptors by HIV-1 Env. To define the mechanism by which NF279 blocks HIV-1 fusion, we reversibly arrested this process at an intermediate step referred to as a temperature-arrested stage (TAS) (10, 33, 43). This is achieved through a prolonged incubation of viruses with target cells at a temperature that is not permissive to fusion, typically ~20°C; a subsequent temperature increase leads to fast and efficient fusion. At the TAS, a fraction of viruses functionally engages a requisite number of CD4 and coreceptor molecules, as evidenced by the acquired partial resistance of fusion to very high concentrations of inhibitors of receptor and coreceptor binding (10, 33, 43). We therefore created a TAS between HXB2pp or BaL26pp and TZM-bl cells and asked whether NF279 was still capable of blocking fusion after the formation of Env-CD4–coreceptor complexes.

The optimal conditions to create a TAS depend on the virus strain and on the CD4/coreceptor expression levels in target cells and therefore must be empirically optimized. In agreement with our previously reported results (10, 33), optimal TAS conditions for HXB2pp and BaL26pp were 2 to 3 h of preincubation at 20°C and 18°C, respectively (Fig. 4B). The TAS was probed by using concentrations of fusion inhibitors that effectively blocked uninterrupted HXB2pp and BaL26pp fusion at 37°C without the preincubation step (Fig. 4A). We found that, as expected, HIV-1 fusion at the TAS was somewhat more resistant to the CD4 inhibitor BMS-806 than to the CXCR4 or CCR5 antagonists AMD3100 and TAK-779 (Fig. 4C). Consistent with the lack of critical gp41 refolding at the TAS (44, 45), a 6-helix bundle inhibitor, the C52L peptide, abrogated HIV-cell fusion (Fig. 4C). HIV-1 fusion at the TAS remained partially sensitive to NF279, and the extent of inhibition was close to those achieved by AMD3100 and TAK-779 (Fig. 4C). The similar extents of fusion inhibition by specific coreceptor antagonists and NF279 indicate that the latter compound targets the coreceptor binding step of fusion, downstream of CD4 binding.

To further ascertain the exact step of HIV-1 fusion targeted by NF279, we employed an inhibitor substitution strategy, which we have previously implemented to define the mechanism of action of human α-defensin (46). In these experiments, HXB2pp or BaL26pp and target cells were preincubated at their respective subthreshold temperatures in the presence of a fully inhibitory concentration of NF279. At the end of incubation, NF279 was washed away, and a fully inhibitory concentration of CD4 or the coreceptor inhibitor was added prior to induction of fusion by shifting cells to 37°C (Fig. 4D). In this protocol, if NF279 blocks CD4 binding, fusion would remain sensitive to both BMS-806 and coreceptor inhibitors. If, on the other hand, coreceptor binding is blocked by NF279, then fusion is expected to remain sensitive to coreceptor antagonists but not to CD4 inhibitors. As shown in Fig. 4E, HXB2pp and BaL26pp fusion remained partially resistant to BMS-806, similarly to the conventional TAS protocol (Fig. 4C), showing that NF279 did not interfere with the Env–CD4 binding step. However, substitution of NF279 with AMD3100 and TAK-779 inhibited HXB2pp and BaL26pp fusion, respectively, to the levels observed with the 6-helix bundle inhibitor C52L (Fig. 4E).
FIG 3 Functional P2X receptors are present in epithelial but not lymphoid cell lines used in fusion experiments. (A) Western blotting does not detect P2X1 receptor expression in target cells of epithelial or lymphoid origin. (Top) P2X1R expression was examined in lysates of HEK 293T/17 (lane 1), TZM-bl (lane 4), and lymphoid (lanes 7 to 9) cells. As a positive control for P2X1R expression, lysates of HEK 293T/17 and TZM-bl cells transiently transfected with wild-type (WT) P2X1R (lanes 2 and 5) or the P2X1RdelL mutant (lanes 3 and 6) are shown. (Bottom) Loading control using α-tubulin antibodies. (B to E) Elevated intracellular calcium concentrations induced by ATP stimulation were measured in TZM-bl (B), HEK 293T/17 (C), CEM-A (D), and Jurkat-CCR5 (E) cells. Cells were loaded with the calcium indicator Fluo4, and fluorescence changes in complete growth medium at 37°C were visualized by time-lapse microscopy. Cells were imaged for ~1 min to acquire the background signal, after which ATP was added at the indicated concentrations (left arrows). After 5 min of image acquisition, the calcium ionophore A23187 was added (right arrows) as a Fluo4 loading control. The curves show the normalized fluorescence intensities of individual cells from a representative experiment performed in triplicate. N is the number of traces shown in each panel.
These findings clearly demonstrate that NF279 blocks HIV-1 Env interactions with both CXCR4 and CCR5.

NF279 inhibits HIV-1–coreceptor engagement in primary human macrophages. To extend the above-described observations to more physiologically relevant cells, we examined the effects of P2 receptor inhibitors and ATP derivatives on BaL26pp fusion with monocyte-derived human macrophages (MDMs). These experiments were carried out by using methods similar to those described above for TZM-bl cells. Briefly, pseudoviruses were prebound to MDMs in the cold and incubated at 37°C for 2 h. This protocol resulted in a robust BlaM signal. Similarly to TZM-bl cells, NF279, but not ATP, α,β-meATP, or the P2X7R inhibitor A740003, suppressed HIV-1 fusion with MDMs (Fig. 5A). The only difference was that αATP modestly reduced the efficiency of fusion with MDMs. The inhibitory effects of αATP (however modest) and NF279 on HIV-1 fusion with MDMs are consistent with data from a previous study (13).

To address whether NF279 inhibits HIV-1 fusion with macrophages through interfering with Env-coreceptor binding, we examined the efficiency of NF279 inhibition after BaL26 Env forms ternary complexes with CD4 and CCR5 on macrophages. Using a protocol similar to that used to capture a TAS with TZM-bl cells, we observed that BaL26 successfully engaged CD4 and CCR5 on MDM cells at a reduced temperature. The formation of ternary complexes was apparent from the partial resistance of fusion to fully inhibitory doses of compounds targeting the CD4 and CCR5 binding steps of HIV-1 fusion (Fig. 5B). The identical degrees of fusion inhibition by saturating concentrations of NF279 and TAK-779 added at the TAS (Fig. 5B) support the notion that NF279 blocks the CCR5 binding step.

To verify this conclusion, NF279 washout experiments were performed, whereby MDMs were incubated for an extended period of time at a subthreshold temperature for fusion in the presence of NF279 (for the experimental protocol, see the legend of Fig. 4D). At this point, the NF279-imposed block of fusion could be readily reversed by simply washing off the inhibitor prior to raising the temperature (Fig. 5C). When NF279 was replaced with TAK-779, fusion with macrophages was abrogated, further confirming that NF279 inhibits the CCR5 engagement step. Interestingly, substitution of NF279 with BMS-806 partially inhibited BaL26pp fusion with TZM-bl cells and MDMs (Fig. 4E and 5C).

Since we have previously shown that NF279 does not downregulate CD4 expression (24), it is possible that NF279 can also...
The P2X1R inhibitor NF279 is a dual CXCR4/CCR5 antagonist. Functional dissection of the HIV-1 fusion steps targeted by NF279 (Fig. 4) shows that this compound blocks Env-coreceptor interactions but does not show whether this effect is due to binding to coreceptors or to the coreceptor binding site on gp120. We therefore examined the ability of NF279 to interact with CCR5 and CXCR4. Since small-molecule coreceptor antagonists are known to inhibit chemokine-mediated calcium signaling (e.g., see references 31 and 47), we asked if NF279 can also modulate this activity in TZM-bl and CEM-A cells. Elevation of the intracellular calcium concentration was visualized by loading the cells with Fluo4-AM and measuring the increases in cell fluorescence in response to antagonists/agonists. To control for the Fluo4-AM loading efficiency, 2.5 μM A23187 was always added at the end of each experiment to evoke maximal increases in fluorescence, and the agonist-mediated signals were normalized to the signal in the presence of the ionophore.

Both RANTES (CCR5 agonist) and SDF-1α (CXCR4 agonist) elicited robust transient increases of cytosolic calcium concentrations in TZM-bl cells (Fig. 6A and B; see also Movie S1 in the supplemental material). As expected, these responses were abrogated when RANTES or SDF-1α was applied together with the respective antagonists of CCR5 and CXCR4 coreceptors, TAK-779 and AMD3100. Importantly, NF279 abrogated Ca2+ influx mediated by either SDF-1α or RANTES, whereas the P2Y11 agonist NF340 exhibited minor to no effects on cytosolic calcium levels (Fig. 6A and B; see also Movie S2 in the supplemental material). In CEM-A cells, SDF-1α application elicited extremely strong calcium responses at levels similar to those elicited by A23187 (Fig. 6C). AMD3100 fully blocked SDF-1α-mediated elevations of calcium concentrations in these cells, whereas high concentrations of NF279 partially inhibited this response. The antagonistic effect of NF279 was more apparent at lower doses of SDF-1α, whereas elevated calcium levels in CEM-A cells induced by a high dose of the agonist were only marginally inhibited (data not shown). Together, our results show that NF279 acts as a dual CXCR4/CCR5 antagonist in TZM-bl cells. In comparison, the ability of NF279 to antagonize CXCR4 signaling in CEM-A cells is limited, so it appears to act as a partial antagonist. Of note, incomplete inhibition of CXCR4 signaling in CEM-A cells seems to correlate with the lower potencies of NF279 and especially of NF449 against HXB2pp fusion with these cells than with TZM-bl cells (Fig. 1).

Considering that NF279 antagonizes calcium signaling mediated by both CCR5 and CXCR4, we asked if this compound could also interfere with the activities of other chemokine receptors. NF279 blocked Ca2+ influx elicited by the CXCR3/CXCR7 agonist I-TAC (interferon-inducible T cell alpha chemoattractant) in CEM-A cells (Fig. 6D). Of note, the average I-TAC-mediated calcium signal was much weaker than that induced by SDF-1α due to a small fraction of cells responding to I-TAC. Collectively, the above-described results indicate that NF279 interacts with chemokine receptors and not with the gp120 coreceptor binding site. This compound most likely blocks chemokine receptor signaling by binding to these proteins. The alternative possibility that NF279 blocks calcium influx by targeting a downstream step of chemokine receptor signaling appears unlikely considering that this compound (i) blocks HIV-1–coreceptor interactions and (ii) is relatively polar and thus should not permeate the plasma membrane.

interfere with Env-CD4 binding, at least in the case of Bal26. Collectively, the above-described results with MDMs confirm and expand our conclusion that NF279 antagonizes Env-coreceptor interactions.
We next asked if NF279 could also interfere with gp120 signaling. The addition of recombinant monomeric gp120 derived from the X4-tropic IIIB strain to TZM-bl cells resulted in transient increases in cytosolic calcium levels. Although calcium responses were observed in only a fraction of cells, the cells responding to this treatment exhibited profound increases in fluorescence (Fig. 7A). In contrast, no calcium signaling was detected when gp120 was added together with NF279 (Fig. 7B), demonstrating that this compound antagonizes gp120-mediated activation of CXCR4. Our results thus show that NF279 is a promiscuous antagonist of several chemokine receptors that is capable of blocking functional interactions between gp120 and coreceptors.

The determinants of NF279 and TAK-779 inhibitory activities are similar but distinct from those of other CCR5 antagonists. Among a panel of CCR5 mutants examined by different laboratories, a G163R substitution at the interface between the second extracellular loop and the transmembrane domain 4 reduces the affinity of gp120 for CCR5 and thereby attenuates virus fusion/infection (48, 49). Nonetheless, JGR.H11 cells expressing the G163R CCR5 mutant support efficient HIV-1 fusion/infection (49). Since this mutation appears to selectively attenuate the antiviral activity of aplaviroc (48) but only modestly affects the activity of the dual CCR5 and CCR2b antagonist TAK-779 (50, 51), we asked whether the dual-coreceptor antagonist NF279 maintains the ability to block HIV-1 entry through the mutant coreceptor.

Fusion of BaL26pp with TZM-bl cells or with JGR.H11 cells expressing the G163R CCR5 mutant was allowed to proceed in the presence of different CCR5 antagonists. In agreement with data from a previous report (49), BaL26pp fusion with TZM-bl cells was twice as efficient as fusion to JGR.H11 cells (data not shown). Whereas high concentrations of the three CCR5 antagonists Scheering C (Sch-C), AD101, and TAK-779 blocked fusion with TZM-bl cells, only TAK-779 antagonized HIV-1 fusion with cells expressing mutant CCR5 (Fig. 8A). Interestingly, similarly to TAK-779, NF279 blocked BaL26pp fusion with both cell lines. In fact, NF279 was ~3-fold more potent against HIV-1 fusion in cells expressing the G163R CCR5 mutant than in those expressing wild-type CCR5 (Fig. 8B). These results reveal differences in the mechanisms of action of NF279 and specific CCR5 antagonists while highlighting similarities with the dual CCR5 and CCR2b antagonist TAK-779.
DISCUSSION

To undergo fusion, HIV-1 strictly requires CD4 and cognate coreceptors. However, additional host factors and cell signaling may modulate this process (3–12). Specifically, several reports have implicated P2 receptors in HIV-1 entry/fusion (13–15, 24). It has been proposed that Env-CD4/coreceptor engagement provides a signal for ATP release from target cells, which in turn activates purinergic receptors and raises intracellular calcium levels. Exactly how these events aided HIV-1 entry remained unclear. Although plasma membrane depolarization resulting from calcium influx has been envisioned to promote HIV-1 fusion (14), we are unaware of direct experimental evidence linking HIV-1-mediated membrane depolarization to enhanced viral fusion.

Our results strongly argue against the involvement of P2 receptors in HIV-1 fusion. First, P2X1R expression on the cells used in our fusion assays was undetectable, and ATP failed to mediate calcium signaling in T cell lines supporting efficient HIV-1 entry. Second, data from mechanistic studies of P2X1R inhibitors imply that the observed inhibition of HIV-1 entry/fusion by NF279 is due to an off-target effect of high concentrations of this antagonist. Indeed, whereas 10 to 100 μM NF279 or NF449 is required to inhibit HIV-1 fusion with different target cells (13–15, 24), nanomolar concentrations of these compounds have been reported to block P2X1 channels (see reference 35 and references therein).

Several lines of evidence support the conclusion that NF279 (and possibly other P2X1R inhibitors) acts as a dual HIV-1 core-
FIG 8 NF279 and TAK-779, but not Sch-C or AD101, inhibit HIV-1 fusion mediated by the attenuated CCR5 mutant. Virus-cell fusion was initiated as described in Materials and Methods and measured by using a BlaM assay. (A) Bal26pp were allowed to fuse with TZM-bl or GJR.H11 cells in the absence or in the presence of the 5 μM the CCR5 antagonist Sch-C, AD101, or TAK-779 or in the presence of 50 μM NF279. *, P < 0.05; ***, P < 0.001. (B) Dose-response curves for NF279-mediated inhibition of Bal26pp fusion with TZM-bl or GJR.H11 cells. Data points are the mean normalized BlaM activities and standard deviations from two experiments conducted in triplicate. IC_{50}, 50% inhibitory concentration.

P2X1R in HIV-1 fusion (13–15, 24), small interfering RNA (siRNA) knockdown of P2X1R in target cells appears to actually promote HIV-1 Env-mediated cell-cell fusion (14). In contrast, depletion of panxenxin-1, P2Y2 receptor, and Pyk2 kinase in target cells attenuated HIV-1 infection and Env-mediated cell-cell fusion (14). Unfortunately, the possibility that knockdown of these proteins can diminish CD4 and/or coreceptor expression in target cells was not addressed in that study. Additional evidence implicating P2Y2 came from experiments with nonspecific inhibitors of P2Y2 receptors and panexxin-1 and usually involved prolonged incubation (24 to 48 h) with high doses of inhibitors (14). Under our conditions, HIV-1 fusion was not inhibited by the P2Y1 inhibitor NF340 (Fig. 1) or the nonspecific P2Y2 inhibitor kaempferol, which has been employed previously (14) (data not shown).

The lack of a considerable inhibitory effect of the P2Y receptor inhibitors on HIV-1 infection of CD4⁺ T cells was also observed previously (15). However, our results do not rule out the role of purinergic receptor signaling in HIV-1 infection. We surmise that long-term inactivation of P2 receptors or downstream signaling pathways can modulate postentry steps of HIV-1 infection, as documented previously (13, 14).

The mechanism by which conventional coreceptor antagonists block HIV-1 fusion has been extensively studied. It appears that these compounds bind to a conserved hydrophobic pocket and allosterically block the functional engagement of coreceptors by HIV-1 Env (e.g., see references 48 and 58–60). Our data imply that NF279 binds to CXCR4 and CCR5 and prevents the functional engagement of these coreceptors by HIV-1 via an unknown mechanism. Although the dual-coreceptor antagonist TAK-779 binds to the same hydrophobic pocket as that of highly specific CCR5 antagonists, the TAK-779-interacting residues do not seem to fully overlap residues interacting with vicriviroc or Sch-C (60). Several CCR5 residues, including G163, are involved in a hydrogen bond network that appears to be essential for maintaining proper protein conformation (48). Consequently, the G163R mutation markedly reduces the binding of antagonists to mutant CCR5 (48, 61). In fact, aplaviroc-CXCR4 interactions have been mapped to several residues lining the conserved hydrophobic pocket and to G163 (48). The fact that both TAK-779 and NF279, but not other CCR5-specific antagonists, effectively inhibit HIV-1 fusion mediated by the G163R mutant (Fig. 8) highlights the similarities between TAK-779 and NF279. However, NF279 appears too large to fit into the hydrophobic pocket on coreceptors occupied by smaller antagonists (Fig. 9).

This elongated symmetric molecule consists of two arms with three sulfates at each end, which is quite different from the structures and net charges of conventional coreceptor antagonists (Fig. 9). An intriguing possibility is that the NF279 sulfates can compete with the N-terminal tyrosine sulfates on CCR5 and CXCR4 for binding to gp120. Although we currently have no evidence for an NF279-gp120 interaction, the ability of NF279 to block CXCR4 and CCR5 signaling does not rule out its binding to gp120. Competition with coreceptors for gp120 binding is consistent with the ability of NF279 to inhibit HIV-1 fusion with cells expressing attenuated CCR5 carrying the G163R mutation near the second extracellular loop (Fig. 8). The mechanism by which NF279 and its analogs inhibit gp120-coreceptor interactions awaits further investigations.

Purinergic receptor inhibitors appear to lack significant toxicity in vivo and are currently being developed as drug candidates to...
treat rheumatoid arthritis and neuropathic pain (62, 63). However, while the dual-coreceptor-antagonistic activity of NF279 can appear desirable for blocking R5- and X4-tropic HIV-1 infection, the promiscuous inhibition of multiple chemokine receptors by this compound may not be a good trait for a drug candidate.

ACKNOWLEDGMENTS

We thank M. Hoylaert for the P2X1 expression vectors and David Kabat for stimulating discussions and Caleb Mason and other members of the Melikyan laboratory for critical readings of the manuscript.

This work was supported by NIH R01 grants GM108480 and GM054787 to G.B.M. and grant R01 AI058828 to P.S.

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