A long and winding road to the RNA world

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Compared to many contributors, I am a relative late-comer to the RNA field, even an interloper if you will. However, my perception is that this field welcomes researchers who enter by way of relevant discoveries. In my burgeoning studies of RNA, I have benefited from the generosity of great scientists who have been willing to take time out of their busy schedules to contemplate my results and provide their insight. Such greats as Joan Steitz, Christine Guthrie, Bill Marzluff, and Jo Ann Wise (to name just a few) have become not only role models but also informal advisors and/or collaborators. Perhaps it greased my entry into the field to share a first name with Dr. Anita Hopper, one of the giants in the field of RNA biology, but I have found the RNA world to be more supportive and accepting than many of the other fields I have flirted with over the years. This attitude of support, acceptance, and intellectual generosity has always been reflected in the RNA journal, which publishes impactful work in the field of RNA biology with no nod to the politics that are pervasive at many other journals. For this reason, I have aspired to publish my own work in RNA from the early days of the journal. Admittedly I am a bit of a slow-starter because I finally achieved this goal in 2014 just in time to become part of the first 20 years of the journal.

My interest in RNA biology and awareness of the RNA journal arose while I was a post-doctoral fellow in Pamela Silver’s laboratory in the mid-1990s. While my own research at the time examined mechanisms for protein import into the nucleus, many of my lab mates were focused on understanding the function of rather elusive RNA binding proteins. I was a bystander to the investigations that stemmed from interest in the Nuclear poly(A) binding (Nab) proteins by Maury Swanson when he was characterizing S. cerevisiae hnRNP proteins in the Dreyfuss laboratory. Maury’s efforts identified a large number of important RNA binding proteins, so he had a plethora of proteins to characterize. After an initial report, studies of Nab2 waned, likely because conditional alleles of this essential gene were surprisingly elusive to obtain. While in the Silver laboratory, I tried to interest many rotons and fellow post-doctoral fellows in the mutant allele that had inexplicably (to this day) emerged from my screen. Apparently my sales pitch was poor as no one was interested. Thus, when I established my own laboratory at Emory University, this nab2 mutant moved with the other −80°C freezer stocks and took up residence in Atlanta.

For a number of years, this nab2 mutant remained in the freezer and seemed destined to stay there until a determined graduate student, Deanna Green, told me that she had chosen to rotate in my laboratory because of my passion in talking about RNA biology in a first year graduate course. She dug into the freezer and our studies of Nab2 began. Initial experiments were very standard as some rudimentary characterization of the protein was required. In my mind, these studies did not merit a submission to RNA but at least we were moving in the right direction. Our work proceeded, characterizing the role of Nab2 using yeast genetics and cell biology as well as structural biology approaches that were carried out in collaboration with another key scientific and life mentor, Dr. Murray Stewart.

In 2006, a critical grant review changed the direction of our research and paved the way for what would become our first RNA-worthy manuscript. While reviewers of the grant

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praised the mechanistic experiments proposed to decipher the function of Nab2, they expressed significant concern that the studies might not translate to human cells where no Nab2 ortholog had been described. This summary statement pushed us to draw on our recent structural studies to identify a putative human ortholog of Nab2, the aptly named zinc finger Cys3His #14 (ZC3H14) protein. A talented long-term member of the lab, Sara Leung, characterized splice variants of human ZC3H14 and localized the protein to nuclear speckles, work that was published in *Gene* as mechanistic insight required for a publication in *RNA* was lacking. These studies sparked an interest in tissue-specific functions of this protein. While budding yeast is an optimal organism for mechanistic studies of conserved pathways, it is sub-optimal for the study of tissue-specific functions. Drawing on local expertise, we initiated a collaboration with Dr. Ken Moberg (Emory University, Department of Cell Biology) to study the *Drosophila* ZC3H14/Nab2 protein, which we termed dNab2, a rather mundane name for the fly community.

Studies of dNab2 carried out by a talented graduate student, ChangHui Pak, who was either brave or naïve enough to take on this project, quickly revealed that the gene is essential in flies as in yeast and hypomorphic mutants show pleiotropic phenotypes affecting a number of different tissues. We were literally scratching our heads trying to decide what tissue we should study, when a phone call came in that provided clear and exciting direction. Colleagues at the Max Planck Institute of Molecular Genetics in Berlin led by Andreas Kuss were studying inherited forms of intellectual disability (previously termed mental retardation). In two separate families, they had linked the occurrence of intellectual disability to mutation of ZC3H14. This finding led us to focus on understanding the neuronal function of ZC3H14/dNab2. In a large collaborative effort, we reported the identification of the human disease and creation of the *Drosophila* model of the disease. However, in my mind, a burning question remained. Could we actually demonstrate that the human protein and fly protein carry out the same function? This important question was tackled by members of Ken Moberg’s laboratory including a post-doctoral fellow, Seth Kelly. Using fly genetics, Seth demonstrated that transgenic expression of human ZC3H14 specifically in neurons, could rescue many phenotypes of dNab2 zygotic mutant flies. His studies included functional assays such as locomotion as well as molecular assays examining poly(A) tail length. This work was very important as it not only validated the use of the fly model but also included the first evidence that human ZC3H14 was critical for proper control of poly(A) tail length, a function previously only demonstrated in budding yeast and flies. This, I thought, might be my chance to publish important and relevant work in *RNA*.

We worked hard to put together a polished manuscript, pushed the button for the submission, and held our collective breath. We were delighted when reviews were returned with minor comments and the Editor stated that we could merely make the changes with no need for the manuscript to be returned to the reviewers. Finally, with the publication of Kelly and Pak et al. in 2014, I had arrived in the RNA world!

While publication in *RNA* was a major achievement for me, this work highlights a major area of research for the future that is pervasive in the world of RNA biology. There are many examples where mutation of a gene encoding an RNA binding protein that is ubiquitously expressed, such as ZC3H14, leads to tissue-specific disease. While many of these diseases impact neurological function, this is not always the case. A striking example is mutation of the nuclear poly(A) binding protein, PABPN1, which is a nuclear-localized polyadenosine RNA binding protein that shares many properties with ZC3H14. Mutation of *PABPN1* leads to a specific form of muscular dystrophy, oculopharyngeal muscular dystrophy (OPMD). Thus, major challenges for the future include understanding the basic functions of key factors such as RNA binding proteins and then integrating this information into the context of specific tissues impacted in human disease. With these challenges ahead, there will be many exciting studies to pursue and publish as the *RNA* journal enters its next 20 years.