Transcriptional attenuation controls macrolide inducible efflux and resistance in Streptococcus pneumoniae and in other gram-positive bacteria containing mef/mel (msr(D)) elements

Scott Chancey, Emory University
Xianhe Bai, Emory University
Nikhil Kumar, University of Maryland
Elliott F. Drabek, University of Maryland
Sean C. Daugherty, University of Maryland
Thomas Colon, University of Maryland
Sandra Ott, University of Maryland
Naomi Sengamalay, University of Maryland
Lisa Sadzewicz, University of Maryland
Luke J. Tallon, University of Maryland

Only first 10 authors above; see publication for full author list.

Journal Title: PLoS ONE
Volume: Volume 10, Number 2
Publisher: Public Library of Science | 2015-02-19, Pages e0116254-e0116254
Type of Work: Article | Final Publisher PDF
Publisher DOI: 10.1371/journal.pone.0116254
Permanent URL: https://pid.emory.edu/ark:/25593/pg1b4

Final published version: http://dx.doi.org/10.1371/journal.pone.0116254

Copyright information:
This is an open access article, free of all copyright, and may be freely reproduced, distributed, transmitted, modified, built upon, or otherwise used by anyone for any lawful purpose.
This is an Open Access article distributed under the terms of the Creative Commons Universal : Public Domain Dedication License (http://creativecommons.org/publicdomain/zero/1.0/), which permits making multiple copies, distribution of derivative works, distribution, public display, and publicly performance, provided the original work is properly cited.

Accessed November 12, 2017 2:24 PM EST
Transcriptional Attenuation Controls Macrolide Inducible Efflux and Resistance in *Streptococcus pneumoniae* and in Other Gram-Positive Bacteria Containing *mef/mel* *(msr(D))* Elements

Scott T. Chancey¹², Xianhe Bai¹², Nikhil Kumar³, Elliott F. Drabek³, Sean C. Daugherty³, Thomas Colon³, Sandra Ott³, Naomi Sengamalay³, Lisa Sadzewicz³, Luke J. Tallon³, Claire M. Fraser³⁴, Hervé Tettelin³⁵, David S. Stephens¹²*

¹¹ Division of Infectious Diseases, Department of Medicine, Emory University School of Medicine, Atlanta, Georgia, United States of America, ² Laboratories of Microbial Pathogenesis, Department of Veterans Affairs Medical Center, Atlanta, Georgia, United States of America, ³ Institute for Genome Sciences, University of Maryland School of Medicine, Baltimore, Maryland, United States of America, ⁴ Department of Medicine, University of Maryland School of Medicine, Baltimore, Maryland, United States of America, ⁵ Department of Microbiology and Immunology, University of Maryland School of Medicine, Baltimore, Maryland, United States of America

*dstep01@emory.edu*

**Abstract**

Macrolide resistance, emerging in *Streptococcus pneumoniae* and other Gram-positive bacteria, is increasingly due to efflux pumps encoded by *mef/mel(msr)* operons found on discrete mobile genetic elements. The regulation of *mef/mel(msr)* in these elements is not well understood. We identified the *mef(E)/mel* transcriptional start, localized the *mef(E)/mel* promoter, and demonstrated attenuation of transcription as a mechanism of regulation of macrolide-inducible *mef*-mediated macrolide resistance in *S. pneumoniae*. The *mef(E)/mel* transcriptional start site was a guanine 327 bp upstream of *mef(E)*. Consensus pneumococcal promoter -10 (5'–TATACT-3') and -35 (5'-TTGAAC-3') boxes separated by 17 bp were identified 7 bp upstream of the start site. Analysis of the predicted secondary structure of the 327 5' region identified four pairs of inverted repeats R1-R8 predicted to fold into stem-loops, a small leader peptide [MTASMRLR, (Mef(E)L)] required for macrolide induction and a Rho-independent transcription terminator. RNA-seq analyses provided confirmation of transcriptional attenuation. In addition, expression of *mef(E)L* was also influenced by *mef(E)L*-dependent mRNA stability. The regulatory region 5' of *mef(E)* was highly conserved in other *mef/mel(msr*)-containing elements including *Tn1207.1* and the *5612Iq* complex in pneumococci and *Tn1207.3* in Group A streptococci, indicating a regulatory mechanism common to a wide variety of Gram-positive bacteria containing *mef/mel(msr*) elements.
Introduction

Macrolides are characterized by 14- to 16-membered lactone rings with attached amino sugars which confer an affinity for bacterial ribosomes. Binding of the 23S ribosomal subunit by a macrolide molecule effectively blocks de novo synthesis of proteins to inhibit bacterial proliferation. Macrolides are used to treat a wide range of bacterial infections and are particularly important for treatment of infections caused by Gram-positive bacteria. These include, among others, soft tissue infections often caused by *Staphylococcus aureus* and *Streptococcus pyogenes*, and invasive diseases caused by *Streptococcus pneumoniae*. Macrolides are often the drug of choice for empirical treatment of upper respiratory tract infections which are most often caused by *S. pneumoniae*. Unfortunately, resistance to macrolides in Gram-positive bacteria, especially in streptococci, complicates the treatment of these diseases. The two major mechanisms of macrolide resistance in these organisms are target modification and active efflux. RNA methylases, such as, Erm(A), Erm(B) and Erm(C) modify specific nucleotides in the 23S rRNA and block macrolide binding. Erm-type methylases confer the MLSB phenotype: high-level resistance to macrolides, lincosamides and streptogramin B. The M phenotype, conferred by macrolide efflux, is generally characterized by lower levels of resistance to 14- and 15-membered macrolides and sensitivity to 16-membered macrolides, lincosamides and streptogramin B.

Macrolide efflux was first described in *Staphylococcus epidermidis* and was attributed to *msr* (A), an ATP-binding subunit ABC-transporter lacking a membrane-binding domain or an obvious membrane bound partner [1]. Subsequently, the *mef* (macrolide efflux) family of genes encoding major facilitator superfamily-type (MFS) efflux pumps were identified in *S. pyogenes* [2] and *S. pneumoniae* [3]. Interestingly, *mef* genes are invariably found upstream and are co-transcribed with a homologue of *msr*(A), named *msr*(D) or *mel*, suggesting a coordinated function between the MFS and the ABC-transporter as a dual efflux pump [4]. Homologues of *mef* and *msr*(D) have been identified in a wide range of Gram-positive bacteria. In *S. pneumoniae* efflux is encoded by one of four *mef* alleles: *mef*(A), *mef*(E), *mef*(O) or *mef*(I). The most common in pneumococci is *mef*(E) which is associated with the *msr*(D) homolog *mel* and is located on the 5.5 kb mobile element Mega (Macrolide efflux genetic assembly) [3]. *Mef*(A) is more commonly associated with group A streptococci (GAS) and is usually carried on phage-related elements. In the pneumococcus, *mef*(A) is carried on the degenerative phage element Tn1207.1 [5]. *mef*(I) was reported in pneumococcal isolates from Spain and is located on the 5612IQ complex, so named because of the presence of the chloramphenicol resistance-encoding *catQ* gene [6].

Macrolide resistance mediated by rRNA methylation or by efflux is inducible by 14- and 15-membered macrolides, such as erythromycin and azithromycin, respectively (reviewed in [7]). It is well established that inducible expression of *erm* genes is controlled by attenuation, usually at the level of translation. Transcripts of inducible *erm* genes are characterized by the presence of one or two small (8-20 amino acids) leader peptides encoded 5’ of the *erm* structural gene. Macrolide-bound ribosomes pause at a programmed stall site on the leader peptide disrupting the attenuator and allowing formation of the anti-attenuator. We have previously demonstrated that, unlike most *erm* genes, *mef*(E) and *mel* are induced at the level of transcription of the genes as an operon (*mef*(E)/*mel*), however the mechanism of induction has not been elucidated [8]. We have also shown that *mef*(E)/*mel* is induced weakly by the human and mouse antimicrobial peptides LL-37 and CRAMP-38, respectively [8,9]. This has not been shown for *erm* genes. The differences in expression of *mef*(E)/*mel* and *erm* genes suggest dissimilar mechanisms control these macrolide resistance genes. The goal of this study was to characterize the mechanism controlling macrolide induction of transcription of *mef*(E)/*mel* by
identifying its promoter and searching for cis-acting regulatory sequences in the mef(E)/mel transcript. In addition, phylogenetically analyses of mef(E)/mel and the macrolide efflux systems in other pneumococcal and Gram-positive bacteria.

Materials and Methods

Ethics statement

All isolates used in the study were received from previously existing pneumococcal strain collections. Ethics approval was obtained from the Institutional Review Board (IRB) of the institutions who collected the biological specimens from the human subjects: Emory University, Centers for Disease Control and Prevention or Georgia Department of Human Resources. Ethics approval was not required for this study because all data were anonymized.

Bacterial strains

*Streptococcus pneumoniae* GA17457 is an erythromycin-resistant, Mega-containing, serotype 19A strain originally isolated from the blood of a patient in the Atlanta metropolitan area in 2001 [3, 4]. GA17457 was selected from 147 isolates, including 115 macrolide resistant isolates, most of which were identified through population-based surveillance or carriage studies conducted by the Georgia Emerging Infections Program (GEIP) (S2 Table) [30]. England14–9 is an erythromycin resistant, mef(A)-containing serotype 14 strain isolated from a patient in the United Kingdom in 1993 [31] (S2 Table). In addition ten strains were isolated from the nasopharynx of healthy children in the Atlanta metropolitan area [32]. Draft genome sequences were deposited into the National Center for Biotechnology (NCBI) Whole Genome Shotgun database (WGS). Isolate metadata and the NCBI accession number for each genome is provided in S2 Table.

Construction of the mef(E)-lacZ reporter strain XZ7042 from GA17457 has been previously described [8,9]. The control strain XZ7049 was generated by insertion of the promoterless lacZ of pPP2 [33] into bgaA of GA17457. The Genbank WGS master accession number for GA17457 draft genome sequence is AILS00000000 (S2 Table).

Construction of mutations in the regulatory region of the mef(E)-lacZ reporter

To generate mutations in the mef(E)/mel 5′ regulatory region governing mef(E)-lacZ expression, plasmid pXZ7032 [9] was used as a template for inverse PCR to introduce point mutations and deletions. For example, the putative mef(E)/mel promoter was deleted by amplification of pXZ7032 with overlapping primers 471–777F and 741–777R (S1 Table). The product was DpnI digested and self-ligated to generate plasmid pXB05 which was transformed into GA17457 and selected on 3 μg ml⁻¹ tetracycline to generate the XZ7042 mef(E)-deletion derivative XB05 (Table 1).

Construction of mutations in the regulatory region of the native mef(E)/mel operon

To assess the influence of the mef(E)/mel 5′ regulatory region on mef(E)/mel-mediated macrolide resistance, mutations were introduced into the region in the native locus of GA17457 by allele replacement. Briefly, the kanamycin resistance gene apha-3 was fused by overlap extension PCR immediately upstream the 1.1 kb region of Mega upstream of mef(E) and cloned into the EcoRI site of pSF151 [34] to create plasmid pXB101. GA17457 was transformed with pXB101 and transformants were selected on 400 μg ml⁻¹ kanamycin creating XB29 (Table 1) containing...
the kanamycin-resistance marker immediately upstream of wild type Mega. This inserted the kanamycin resistance marker 780 bp upstream of the mef(E)/mel transcriptional start site. Because of the distance between the desired mutation location and the selectable marker, recombination events resulting in kanamycin resistance, but without the desired mutation were possible. Therefore, the nucleotide sequence of the entire 1.1 kb region upstream of mef(E) of the resulting mutants were determined to confirm the mutations and that no other changes in the regulatory region occurred. Strain XB29 was selected as the positive control strain for erythromycin susceptibility assays.

To generate mef(E) regulatory mutants, plasmid pXB101 was used as template for inverse PCR using primers designed to create deletions or introduce point mutations (S1 Table). Inverse PCR products were DpnI digested, ligated and transformed into GA17457 as above. Selection for kanamycin resistance, screening for promoter deletion by PCR and verification of the deletion produced the promoter deletion mutant XB30. XB30 was complemented by restoration of the promoter sequence by transformation of XB30 with a PCR product containing the wild type promoter and selected on 1.0 \( \mu \)g ml\(^{-1}\) erythromycin. The sequence of the entire 1.1 kb mef(E) regulatory region and the putative promoter deletion were verified by Sanger DNA sequencing.

### Susceptibility assays

The minimum inhibitory concentration (MIC) of erythromycin was determined by Etest as per manufacturer recommendations (AB bioMerieux, Solna, Sweden). To determine the effects of erythromycin induction on resistance, strains were cultured overnight at 37°C in 5% CO\(_2\) on TSA blood agar plates with or without 10\(^{-1}\) MIC concentration of erythromycin representing inducing and non-inducing conditions, respectively. Cell suspensions from the overnight plates were swabbed onto cation-adjusted Mueller-Hinton agar plus 5% horse blood. Plates were

---

**Table 1. Strains used in this study.**

<table>
<thead>
<tr>
<th>Strain</th>
<th>Description</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>GA17457</td>
<td>Serotype 19A, ST199 invasive isolate with Mega. Ery(^R)</td>
<td>[8,9]</td>
</tr>
<tr>
<td>England(^{14,9})</td>
<td>Serotype 14, ST19 invasive isolate with Tn1207.1. Ery(^R)</td>
<td>[31]</td>
</tr>
<tr>
<td>XZ7042</td>
<td>GA17457 bgaA::mef(E)::lacZ reporter strain; Ery(^R)</td>
<td>[8,9]</td>
</tr>
<tr>
<td>XZ7049</td>
<td>GA17457 bgaA::lacZ (promoterless) Mega. Ery(^R)</td>
<td>[8]</td>
</tr>
<tr>
<td>XZ8009</td>
<td>GA17457 mef(E)::aphA3. Ery(^R)</td>
<td>[9]</td>
</tr>
<tr>
<td>XB05</td>
<td>XZ7042 bgaA::mef(E)(-41)-(-5)::lacZ</td>
<td>This study</td>
</tr>
<tr>
<td>XB29</td>
<td>GA17457 aphA3-Mega wild type control strain</td>
<td>This study</td>
</tr>
<tr>
<td>XB30</td>
<td>GA17457 aphA3-Mega(+)::甲方(-41)-(-5)</td>
<td>This study</td>
</tr>
<tr>
<td>XB30R</td>
<td>XB30 aphA3-Mega deletion restored</td>
<td>This study</td>
</tr>
<tr>
<td>XB16</td>
<td>XZ7042 bgaA::mef(E)(+1)-(-2)::lacZ</td>
<td>This study</td>
</tr>
<tr>
<td>XB34</td>
<td>GA17457 aphA3-Mega(+)::甲方(-41)-(-5)</td>
<td>This study</td>
</tr>
<tr>
<td>XB03</td>
<td>XZ7042 bgaA::mef(E)(+2)-(-2)::lacZ</td>
<td>This study</td>
</tr>
<tr>
<td>XB32</td>
<td>GA17457 aphA3-Mega(+)::甲方(-2)-(-2)</td>
<td>This study</td>
</tr>
<tr>
<td>XB14</td>
<td>XZ7042 bgaA::mef(E)(+)-(-2)-(-2)</td>
<td>This study</td>
</tr>
<tr>
<td>XB45</td>
<td>GA17457 aphA3-Mega(+)::甲方(-2)-(-2)</td>
<td>This study</td>
</tr>
<tr>
<td>XB12</td>
<td>XZ7042 bgaA::mef(E)(+)+(-2)-(-2)</td>
<td>This study</td>
</tr>
<tr>
<td>XB13</td>
<td>XZ7042 bgaA::mef(E)(+)+(-2)-(-2)</td>
<td>This study</td>
</tr>
<tr>
<td>XB36</td>
<td>GA17457 aphA3-Mega(+)::甲方(-2)-(-2)</td>
<td>This study</td>
</tr>
<tr>
<td>XB37</td>
<td>GA17457 aphA3-Mega(+)::甲方(-2)-(-2)</td>
<td>This study</td>
</tr>
</tbody>
</table>

---

doi:10.1371/journal.pone.0116254.t001
overlaid with Etest strips and incubated overnight at 37°C in 5% CO₂. Erythromycin MIC is reported as µg ml⁻¹. Values are the averages of the results of at least three independent experiments. Bold font indicates a greater than 2-fold reduction in erythromycin resistance compared to the similarly treated control XB29. Underlined values indicate at least 3-fold difference compared to the control. ND, not determined

5’ RACE
The 5’ end of the mef(E)/mel transcript from GA17457 was mapped by 5’ Rapid Amplification of cDNA ends (5’ RACE) as has been previously described [35]. Briefly, total RNA was extracted from GA17457 grown in 10⁻¹ MIC erythromycin (12 µg ml⁻¹) using the Qiagen RNAeasy Mini Kit (Qiagen Sciences, Germantown, MD). RNA was reverse transcribed into cDNA using Avian Myeloblastosis Virus (AMV) reverse transcriptase (New England Biolabs, Ipswich, MA) and target-specific primers SC10, SC143 and SC199 designed to produce overlapping fragments covering the entire 1.1 kb between the start of the Mega element and the mef (E) start codon (S1 Table). Reverse transcription reactions were incubated at 55°C for 1 hr and terminated at 65°C for 10 min. Poly-A tails were added to the 5’ end of cDNAs by incubation with dATP and terminal transferase at 37°C for 20 min. Tailing reactions were terminated at 70°C for 10 min. dA-tailed cDNA was amplified with primer RACE-TT (5’ GTTCAGCG-CAGGTTCTTTTTTTTTTTTTT 3’) and the appropriate target-specific primers SC10, SC143 or SC199 (S1 Table). Final amplification was conducted with forward primer RACE-T-F and a nested primer for each reaction (SC10-NEST, SC143-NEST, or SC199-NEST) (S1 Table). Final amplification products were purified and the nucleotide sequence was determined.

mRNA secondary structure prediction
The secondary structure in RNAFold (ViennaRNA Package 2.0 [11–13]) to
Analyses were performed on the full length e 327 bp region from the promoter to the mef(E) start codon and on systematically smaller fragments thereof. In silico, 5’ and 3’ nested deletions and other fragments were tested to account for the effects of 5’ folding during transcription, 5’ to 3’ degradation, and ribosomal interference.

β-galactosidase assays
To measure expression of mef(E)-lacZ, cultures of XZ7042 and mutant derivatives were grown to mid-log phase (OD₆₀₀ ~0.3–0.4), divided and exposed to concentrations of erythromycin equal to one-tenth the non-induced MIC of each strain, as determined by Etest (described above). Each subculture was harvested one hour after induction for assessment of β-galactosidase activity. Cultures for time course expression assays were treated as above with cells harvested from continuous cultures at the point of induction and at 30 min intervals thereafter. Expression was determined by standard β-galactosidase protocols [36]. Data was expressed as mean Miller Units ± standard error of the mean (SEM). All experiments were performed in duplicate.

RNA-Seq
The erythromycin (ERY) minimum inhibitory concentrations were determined by microdilution and Etest. Induced MIC was determined in the same manner after overnight growth of the isolates on TSA+ sheep blood agar amended with 10⁻¹ the erythromycin MIC of each strain. Mid-log cultures of GA17457 were divided equally into four aliquots and each parallel culture was treated with erythromycin (1.2 µg ml⁻¹), LL-37 (200 µg ml⁻¹), spiramycin (0.1 µg
ml⁻¹) or left untreated. Cells were harvested after a one hour incubation and total RNA was extracted, enriched for mRNA, reverse transcribed and sequenced. Strains were induced at mid-log growth phase (OD₆₀₀) as indicated and cells were harvested after one hour incubation. Total RNA from bacterial samples in each condition was extracted using the QIAGEN RNeasy Mini Kit (Qiagen Sciences, Germantown, MD) followed by DNase treatment in solution and rRNA depletion with the Epicentre Ribo-Zero kit (Epicentre, Madison, WI). Strand-specific RNA-Seq cDNA libraries were constructed and sequenced on the Illumina HiSeq2000 platform; two libraries were combined per channel. The data discussed in this publication have been deposited in NCBI’s Gene Expression Omnibus [37] and are accessible through GEO Series accession number GSE54176 (http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE54176).

Results
Identification of the transcriptional start site of the mef(E)/mel operon
The serotype 19A invasive pneumococcal isolate GA17457 expresses inducible mef(E)/mel-mediated macrolide resistance (Table 1) [8,9]. The start site of the erythromycin-induced mef(E)/mel transcript from GA17457 was mapped by 5' RACE. Unless otherwise stated, all isolates and mutants were induced by one hour exposure to erythromycin at a concentration equal to 10⁻¹ minimum inhibitory concentration (MIC). The 5' RACE reactions produced an approximately 0.4 kb 5' RACE PCR product. The nucleotide sequence of the 5' terminus correlated to a guanine residue located 327 bp upstream of mef(E) and was tagged with a run of 8–10 consecutive thymine residues which is a hallmark of 5' RACE (Fig. 1A). Consensus-35 (5' -TTGAAC-3') and -10 (5' -TATACT-3') sequences were identified beginning eight base pairs 5' of guanine transcriptional start (+1G) and were identical to previously identified pneumococcal promoters (Fig. 1B) [10]. The -10 site was also a consensus “extended-10” pneumococcal promoter (5' -TTGTGTTATACT-3') (Fig. 1B). A second poly-T tract was noted on the 5' RACE chromatograph (Fig. 1A). The second detected transcript appeared to begin at the cytosine residue (+41C) positioned 287 bases upstream of mef(E) (Fig. 1A). However, no promoter sequences were detected immediately upstream of +41C suggesting that it was not an alternate transcription start site. The mRNA molecule terminating with +41C detected by 5' RACE may be a product of transcript degradation and may reflect biologically relevant endoribonuclease activity (see below).

To show that the predicted promoter was required for expression of mef(E), a 37 bp fragment [Δ-41-(-5)] containing the putative-10 and -35 boxes was deleted in the reporter locus inserted into bgaA of XZ7042 (Fig. 1B, 1C). The resulting mutant, XB05, was tested for non-induced and erythromycin-induced expression of mef(E)-lacZ (Fig. 2). As anticipated, the expression of mef(E)-lacZ in XZ7042 containing the wild type promoter was induced rapidly by erythromycin and reached maximum induction (6.8-fold) after 1 hour. Expression in the promoter deletion mutant XB05 was negligible, regardless of erythromycin induction and was similar to the negative control, a promoterless lacZ, strain XZ7049 (Fig. 2) These data confirm that the promoter was required for transcription of mef(E)-lacZ. Further, site directed mutagenesis of +1G to a thymine (G+1T) in the reporter locus of the GA17457-derived mef(E)-lacZ transcriptional reporter strain XZ7042 resulted in the mutant reporter strain XB16. Under non-inducing conditions β-galactosidase (β-gal) activity of XB16 was not significantly different from XZ7042, however, under erythromycin-inducing conditions, expression by XB16 was 3.9-fold lower, confirming that +1G is required for optimal induction of mef(E)/mel (Table 2).

To confirm that the promoter was also required for macrolide resistance, the native mef(E)/mel locus of GA17457 was introduced by allele replacement using the kanamycin resistance marker
Fig 1. Identification of the promoter for the mef(E)/mel(msr(D)) operon. a) Identification of the operon transcriptional start site in the chromatogram of the nucleotide sequence of 5’RACE PCR products. Poly-thymine runs indicate the 5’ end of a transcript molecule. Numbered arrows indicate the predicted 5’ termini. b) Comparison of the predicted mef(E)/mel promoter sequence with putative promoters of other mef-containing elements mef(A) and mef(I) found in S. pneumoniae. The extended-10 and the-35 promoter sequences are underlined and labeled. The deleted sequence (Δ-41-(-5) of mutations in the native mef(E) locus (XB30) or the mef(E)-lacZ reporter locus (XB05) is indicated; ‘+1’ indicates the transcriptional start site. Shading indicates dissimilar nucleotides compared to the mef(E) promoter region. c) The top panel illustrates the genetic organization of mef-containing elements in S. pneumoniae. The homology of the 3.2 “mef cassette” was extensive from RBS1 through mef(msr(D)) as indicated by shading. Blue arrows, macrolide resistance genes; red arrow, chloramphenicol resistance gene catQ; green open arrows, transposon-related genes; open arrows, unrelated genes. The bottom panel illustrates the series of mutations generated in the 5’ regulatory region of the native mef(E)/mel locus of GA17457 and the reporter locus, a mef(E)-lacZ fusion inserted into bgaA in XZ7042. The annotated red line represents the 327 base 5’ mef(E)/mel regulatory mRNA region with the mef(E)/mel promoter shown as a bent arrow. Each pair of converging, colored solid arrows represents one of four pairs of inverted repeats. Black arrow, mef(E)L. closed arrow heads, distal 17 base inverted repeats (dIR1 and dIR2); open arrow heads indicate 12 base direct repeats (DRs). The descriptive name of each mutation and the resulting native-locus and reporter mutant designations are indicated in the right-hand columns; nc, not created.

doi:10.1371/journal.pone.0116254.g001
The β-galactosidase activity of promoter mutations in the mef(E)-lacZ reporter locus. XZ7042 (squares) is the reporter strain generated by insertion of a mef(E)-lacZ fusion into the pneumococcal β-galactosidase gene bgaA; XZ7049 (diamonds), promoterless lacZ strain; XB05 (circles), strain containing the mef(E) promoter region deletion (Δ-41-(-5)) in the mef(E)-lacZ fusion in the bgaA locus. Dashed lines and open symbols, non-inducing conditions; solid lines and closed symbols, inducing conditions (treated with erythromycin at a concentration equal to 10^{-1} erythromycin MIC). Bars represent the standard error of the mean (SEM) of experiments performed in duplicate.

doi:10.1371/journal.pone.0116254.g002

Table 2. Influence of regulatory mutations on resistance to erythromycin and expression of mef(E).

<table>
<thead>
<tr>
<th>Mutationa</th>
<th>Reporter mutant</th>
<th>mef(E)-lacZ expression (Miller units±SEM)c</th>
<th>Native locus mutant</th>
<th>Ery MIC (µg ml^{-1})e</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>-Ery</td>
<td>+Ery</td>
<td></td>
</tr>
<tr>
<td>none</td>
<td>XZ7042</td>
<td>13.0±2.9</td>
<td>80.0±4.5</td>
<td>XB29</td>
</tr>
<tr>
<td>-controlb</td>
<td>XZ7049</td>
<td>5.1±3.2d</td>
<td>7.6±0.2d</td>
<td>XZ8009</td>
</tr>
<tr>
<td>Δ-41-(-5)</td>
<td>XB05</td>
<td>10.5±0.9</td>
<td>16.4±1.9d</td>
<td>XB30</td>
</tr>
<tr>
<td>Δ-41-(-5) C’</td>
<td>na</td>
<td></td>
<td></td>
<td>XB30R</td>
</tr>
<tr>
<td>G+1T</td>
<td>XB16</td>
<td>6.8±1.3</td>
<td>20.4±0.2d</td>
<td>XB34</td>
</tr>
<tr>
<td>RBS1’</td>
<td>na</td>
<td></td>
<td></td>
<td>XB37</td>
</tr>
<tr>
<td>Δ+162-189</td>
<td>XB14</td>
<td>4.8±3.1d</td>
<td>11.2±2.5d</td>
<td>XB45</td>
</tr>
<tr>
<td>Δ+243-280</td>
<td>XB03</td>
<td>89.0±0.9d</td>
<td>145.7±2.3</td>
<td>XB32</td>
</tr>
<tr>
<td>Δ+54-93</td>
<td>XB12</td>
<td>8.6±3.1d</td>
<td>16.6±2.5</td>
<td>XB36</td>
</tr>
<tr>
<td>Δ+63-80</td>
<td>XB13</td>
<td>62.2±4.4d</td>
<td>64.8±0.9</td>
<td>na</td>
</tr>
<tr>
<td>Δ+19-298</td>
<td>na</td>
<td></td>
<td></td>
<td>XB31</td>
</tr>
<tr>
<td>Δ+63-298</td>
<td>na</td>
<td></td>
<td></td>
<td>XB38</td>
</tr>
</tbody>
</table>

a mutations introduced into the 327 bp regulatory region controlling expression of a mef(E)-lacZ transcriptional fusion inserted into the bgaA locus of XZ7042 or the regulatory region controlling expression of mef(E)/mel in the native Mega locus of GA17457. Numbers are relative to the transcriptional start site (+1), Δ, deletion; C’, mutation complemented by allele replacement; RBS1’, site-direct mutagenesis reverse complementing the mef(E)L ribosomal binding site.

b negative control. XZ7049, promoterless mef(E)-lacZ inserted into bgaA of GA17457; XZ8009, mef(E)/mel deletion in GA17457.

c mean ± standard error of the mean (SEM) β-galactosidase assays performed in duplicate.

d indicates a significant difference from the similarly treated control. Relative expression data were analyzed for statistical significance by the unpaired two-tailed Student’s T-test. Changes in MIC values greater or equal to four were considered significant.

e MIC, minimum inhibitory concentration, determined by Etest. Values are the mean of at least three independent experiments. MICs are given in micrograms per milliliter. Bold font indicates a greater than 2-fold change in erythromycin resistance compared to the similarly treated control XB29. Underlined values indicate at least 3-fold difference compared to the control. ND, not determined.

doi:10.1371/journal.pone.0116254.t002
aphA-3 inserted immediately upstream of Mega, strain XB29. Erythromycin resistance in XB29 containing the aphA-3 and the wild type mef(E)/mel regulatory region was identical to the wild type strain GA17457 (MIC 12 μg ml⁻¹). Induction of mef(E)/mel expression by erythromycin resulted in an erythromycin MIC of 64 μg ml⁻¹ for XB29, similar to the induced resistance in GA17457 (MIC 48 μg ml⁻¹) indicating that the aphA-3 marker did not influence macrolide resistance or induction. A promoter deletion mutant, XB30, containing the Δ-41-(-5) mutation was sensitive to erythromycin (MIC ≤0.094 μg ml⁻¹) under non-inducing and inducing conditions. Restoration of the Δ-41-(-5) deletion in mutant XB30R restored non-induced and induced erythromycin resistance to wild type levels (MIC 8 μg ml⁻¹ and 64 μg ml⁻¹, respectively) (Table 2). Thus, the mef(E)/mel transcript contains a 327 base leader sequence including the promoter and also nucleotides not translated as part of the mef(E) and mel structural genes that be involved in regulation of the operon.

Predictions of the secondary structure of the mef(E)/mel 5’ regulatory mRNA

Having defined the start of the mef(E)/mel transcript and the promoter, we sought to identify cis-acting regulatory sequences in the 327 base regulatory leader sequence that may contribute to inducible expression of the mef(E)/mel operon. Four pairs of proximal inverted repeats were identified and predicted by RNAFold [11–13]) to form four stem-loops (R1/R2, R3/R4, R5/R6 and R7/R8) (Fig. 1C; Fig. 3). The 54 base repeat R1 annealed to the 51 base R2 to produce a stem-loop (R1/R2) containing a 44 bp stem (37.5% G+C), a 21 base terminal loop. The predicted minimal free energy (ΔG) of the R1/R2 stem-loop was -39.5 kcal mol⁻¹ (Fig. 3). Immediately adjacent to R1/R2 was stem-loop R3/R4 containing an eight base pair stem with a ΔG of -8.1 kcal mol⁻¹ (Fig. 3). Overlapping the inverted repeats R3 and R4 was a pair of perfect 12 base direct repeats (5’ UUAUUUAACUAU 3’) separated by four bases (red arrows, Fig. 3).

Stem-loop R5/R6, located 39 bp downstream of repeat R4, had the characteristics of a classic Rho-independent transcriptional terminator [14]; the R5/R6 loop was predicted to be stable (ΔG = -18.0 kcal mol⁻¹), contained a 55.6% G+C-rich 15 bp stem, and was immediately followed by a run of five uracil residues (Fig. 3). The erm(K) transcriptional attenuator has two adjacent Rho-independent transcriptional terminators [15], raising the possibility that last stem-loop R7/R8 in the mef(E)/mel 5’ regulatory mRNA was a second terminator. However, R7/R8 did not have the characteristics of a Rho-independent terminator; the stem was not G+C-rich (14.3%), it was not predicted to be thermodynamically stable (ΔG=5.3 kcal mol⁻¹) and was not followed by a poly-uracil tract.

Identification of a leader peptide mef(E)L required for mef(E)/mel expression

Expression of inducible erm methylase genes occurs when erythromycin-bound ribosomes stall during translation of a small leader peptide encoded upstream of the structural gene on the erm transcript. A search of the 327 base 5’ regulatory sequence of the mef(E)/mel transcript revealed a small (eight codon) open reading frame (hereafter called mef(E)L) located 34 bases from +1G and preceded by a consensus ribosomal binding site (5’ AGGAGG 3’) (hereafter called RBS1) (Fig. 1C; Fig. 3). The peptide sequence of Mef(E)L (MTASRLR) was similar to predicted and confirmed leader peptides of other macrolide resistance determinants (Table 3). Notably, Mef(E)L was homologous to the Bacillus licheniformis erm(K) leader peptide Erm(K)L (MTHSMRLLRFPPTLNQ) and the putative leader peptide Msr(A)L (MTASMRLRQ) encoded upstream of the macrolide-inducible mel(msr(D)) homolog msr(A) from S. epidermidis (Table 3) [1,15]. The similarity of mef(E)L to erm(K) was of note because erm(K) is one of the few erm-
Fig 3. Structure of the mef(E)/mel 5' attenuator structure (transcriptionally inactive). The structure of the 327 nucleotide 5’ mRNA region predicted by RNAFold and visualized with Visualization Applet for RNA (VARNA) [Version 3.8; [43]]. Inverted repeats are shaded with colors consistent with Fig. 1C. The amino acid sequence of the leader peptide Mef(E)L is indicated. The minimum free energy values from each stem-loop predicted by RNAFold analyses of the nucleotides involved in each duplex. Red arrows direct repeats. Dashed arrows, distal inverted repeats; RBS1, mef(E)L ribosomal binding site; RBS2 mef(E) ribosomal binding site.

doi:10.1371/journal.pone.0116254.g003
type methylase genes known to be regulated by transcriptional attenuation [15] and mef(E)/
mel is transcriptionally regulated [4,8]. The minimal leader peptide sequence required for
erythromycin induction of erm(K) has been shown to be MTHSMRLR [15,16], nearly identical
to Mef(E)L suggesting that the mef(E) leader peptide may be the site of ribosome stalling in
the presence of macrolides. Apart from the similarities of the leader peptide, no discernible homol-
gy was noted between the 327 base
mef(E)/mel
50 regulatory region and the 354 base 50 regulatory region of
erm(K) .
To confirm that translation of the predicted leader peptide mef(E)L was required for
mef(E)/mel expression, the native locus
mef(E)L was translationally inactivated by altering the ribo-
somal binding site from AGGAGG (RBS1) to CCTCCT (RBS1') in the mutant XB37 (Fig. 1C;
Table 2). XB37 was sensitive to erythromycin in inducing and non-inducing conditions indi-
cating that mef(E)L expression was required for mef(E)/mel-mediated resistance (Table 2).
These data were consistent with transcriptional attenuation as a mechanism whereby stalled
macrolide-bound ribosomes interfere with stem-loop R1/R2 to allow the formation of an anti-
attenuator.

**Stem-loop R5/R6 is a Rho-independent transcriptional terminator**

The stem-loop structure R5/R6 was present in the predicted attenuator but absent from the
predicted anti-attenuator structure consistent with the conclusion that R5/R6 represents a
Rho-independent transcriptional terminator in non-inducing conditions. To determine if R5/
R6 was required for repression of mef(E)/mel expression in non-inducing conditions a 39 bp
deletion (Δ+243–280) containing repeat R6 and the 5’ end of R7 was introduced into the re-
porter locus of XZ7042 and into the native Mega locus of GA17457 creating XB03 and XB32,
respectively (Fig. 1C). The non-induced expression of mef(E)-lacZ in XB03 (89.0 m.u.) was 6.8-
fold higher than non-induced expression in XZ7042 and was nearly the same as induced ex-
pression in XZ7042 (Table 2). However, expression in the XB03 remained inducible, increasing
1.6-fold to 145.7 m.u. after induction by erythromycin (Table 2). This may suggest an addition-
al mechanism of regulation and induction.

Erythromycin resistance of XB32, the native locus
Δ+243–280 mutant, in non-inducing
conditions was 4-fold higher than in the native locus control XB29 (Table 2). Erythromycin re-
sistance in macrolide inducing conditions exceeded the limit of the Etest assay (MIC >256 μg
ml−1), >4x higher than XB29 (MIC 48 μg ml−1) (Table 2). These data further indicated that

---

**Table 3. Putative leader peptide sequences of inducible macrolide resistance genes.**

<table>
<thead>
<tr>
<th>Structural gene</th>
<th>Peptide sequence</th>
<th>Species</th>
<th>Genbank accession no. (reference)</th>
</tr>
</thead>
<tbody>
<tr>
<td>mef(E)</td>
<td>MTASMRLR</td>
<td>S. pneumoniae</td>
<td>AILS01000012 (This study)</td>
</tr>
<tr>
<td>mef(A)</td>
<td>MTASMRLR</td>
<td>S. pyogenes</td>
<td>AY657002 [38]</td>
</tr>
<tr>
<td>mef(A)</td>
<td>MTASMRLR</td>
<td>S. pneumoniae</td>
<td>AIL00000000 (This study)</td>
</tr>
<tr>
<td>mef(ll)</td>
<td>MTASMRLR</td>
<td>S. pneumoniae</td>
<td>AJ971089 [6]</td>
</tr>
<tr>
<td>mef variant 1</td>
<td>MTASMRLR</td>
<td>S. suis</td>
<td>CP002465 [39]</td>
</tr>
<tr>
<td>mra(A)</td>
<td>MTASMRLK</td>
<td>Staphylococcus epidermidis</td>
<td>X52085 [1]</td>
</tr>
<tr>
<td>mra(C)</td>
<td>MTASMKLRFELLNNN</td>
<td>Enterococcus</td>
<td>ABD51781 [40]</td>
</tr>
<tr>
<td>ermA(D)</td>
<td>MTHAMRLRFPTLNQ</td>
<td>B. licheniformis</td>
<td>M77505 [41]</td>
</tr>
<tr>
<td>ermA(J)</td>
<td>MTHSMLR</td>
<td>B. licheniformis</td>
<td>M29832 [42]</td>
</tr>
<tr>
<td>msr(A)</td>
<td>MTASMRLK</td>
<td>Staphylococcus epidermidis</td>
<td>X52085 [1]</td>
</tr>
<tr>
<td>msr(C)</td>
<td>MTASMKLRFELLNNN</td>
<td>Enterococcus</td>
<td>ABD51781 [40]</td>
</tr>
<tr>
<td>erm(B)</td>
<td>MTHAMRLRFPTLNQ</td>
<td>B. licheniformis</td>
<td>M77505 [41]</td>
</tr>
<tr>
<td>erm(K)</td>
<td>MTHAMRLRFPTLNQ</td>
<td>B. licheniformis</td>
<td>M77505 [41]</td>
</tr>
</tbody>
</table>

*a* underlined sequence represents amino acid residues conserved in the programmed ribosome stall site of erm(K) [16].

---

**doi:**10.1371/journal.pone.0116254.t003
stem-loop R5/R6 was needed for repression of mef(E)/mel transcription, and that R5/R6 functioned as a Rho-independent transcriptional terminator.

**Prediction of the anti-attenuator structure of the mef(E)/mel transcript**

To identify the anti-attenuator structure, additional inverted repeats in the 327 base leader sequence that could be involved in alternate folding of the mef(E)/mel transcript were identified. A pair adenine-uracil rich 18 base repeats separated by 174 bases, predicted not to anneal were located in the attenuator structure (dashed arrows, Fig. 3). The first unit of these distal repeats, dIR1, was located at the 3’ end of R1, six bases downstream of mef(E)L suggesting availability for pairing when a stalled ribosome disrupts the R1/R2 stem-loop. The second distal repeat unit, dIR2 overlapped the Rho-independent transcriptional terminator and included a poly-U tract such that pairing of the distal inverted repeats dIR1 and dIR2 would disrupt the transcriptional terminator and allow transcription to proceed. Under the presumption that ribosome stalling would prevent folding of the transcript from the start through the mef(E)L coding sequence, the 5’ terminal 63 bases were excluded from interacting in the in silico folding of the regulatory region.

The predicted transcriptionally active anti-attenuator structure is shown in Fig. 4. The 5’ terminal 63 bases were excluded from interacting in the in silico folding of the regulatory region under the presumption that ribosome stalling would prevent folding of the transcript from the start through the mef(E)L coding sequence. The leader sequence +60–327 was predicted to fold into a stable (∆G = 67 kcal mol⁻¹) containing a duplex formed by annealing of dIR1 and dIR2 (Fig. 4). Significantly, the putative Rho-independent terminator R5/R6 was resolved, supporting the predicted identity of the anti-attenuator structure (Fig. 4).

To experimentally test the anti-attenuator structure, a deletion of 18 bases corresponding to dIR1 (∆+63–80) was introduced into the reporter locus of XZ7042 creating XB13 (Fig. 1C). Folding analyses predicted deletion of dIR1 would destabilized the R1/R2 stem-loop, result in the annealing of R6 and the novel sequence created between the truncated R1 and the repeat R2 (Fig. 5), and prevent the formation of the R5/R6 terminator. This favored formation of the anti-attenuator structure even under non-inducing conditions (Fig. 5). The model of attenuation due to a single Rho-independent transcriptional terminator would predict this mutant to constitutively expression mef(E)/mel and macrolide resistance (Fig. 5). Indeed, in non-inducing conditions, expression of mef(E)-lacZ was 4.8 times higher in XB13 (62.2 m.u.) than in the wild-type reporter XZ7042 (13.0 m.u.), and was not further induced by erythromycin exposure (64.8 m.u.) (Table 2).

The roles of stem-loop R1/R2 and the leader peptide mef(E)L were further tested by introduction of a second deletion (∆+54–92) into the reporter locus to create the reporter mutant strain XB12 (Fig. 1C). Like ∆+63–80, this mutation deleted dIR1 but extended upstream to disrupt mef(E)L. Folding analyses of the ∆+54–92 deletion predicted that the transcriptional terminator R5/R6 would form in XB12 (Fig. 5) and would form a R1/R2 stem-loop, albeit with reduced stability. The ∆+54–92 deletion also resulted in a truncation of mef(E)L due to substitution of the arginine codon (AGA) in the eighth position with an opal stop codon (UGA) (Fig. 5). The premature termination of the leader peptide was predicted to prevent ribosome stalling and thus render mef(E)/mel expression uninducible.

Indeed, basal expression of β-gal activity in the ∆+54–92 reporter mutant XB12 was not significantly different from the negative controls XZ7049 or XB05 and, under inducing conditions expression was 4.8-fold lower than wild-type (16.6 m.u. and 80.0 m.u., respectively) indicating a nearly complete loss of induction by macrolides (Table 2). Consistent with this result, the ∆+54–92 native locus mutant XB36 (Fig. 1C) was susceptible to erythromycin regardless of prior
erythromycin induction (MICs 0.125 μg ml⁻¹) (Table 2). These data confirm the role of stem-loop R1/R2, and the leader peptide mef(E)L, and support the theory that induction of mef(E)/mel is due to ribosome stalling at mef(E)L in the presence of inducing macrolides.

Stem-loop R3/R4 is also required for the regulation of mef(E)/mel expression

Stem-loop R3/R4 was formed by imperfect annealing of R3 and R4 repeats (Fig. 3). R3/R4 contained a pair of 12 base direct repeats (Fig. 3, red arrows) and could function as a cis-acting operator, influencing mef(E)/mel expression through interaction with trans-acting regulatory factors. To determine if R3/R4 influenced mef(E)/mel expression, 28 bp containing the direct repeats (Δ+162–189) was deleted from the mef(E)-lacZ reporter locus to create reporter strain XB14 (Fig. 1C). Folding analyses predicted the terminator would form even without erythromycin induction (Fig. 6). β-gal activity expressed in XB14 grown with or without erythromycin induction was markedly reduced and was similar to the negative controls XZ7049 or XB05, demonstrating that mef(E)-lacZ was not expressed in an R3/R4 mutant (Table 2). Likewise, XB45 containing the Δ+162–189 deletion in Mega was susceptible to erythromycin with or
without induction with erythromycin (MICs 0.75 μg ml⁻¹) (Table 2). These data confirmed the requirement of stem-loop R3/R4 in the regulation of mef(E)/mel expression.

Mef(E)/Mel regulation differs from the attenuation model of erm methylases

The observation that the R5/R6 terminator deletion mutants remained inducible to macrolides suggested the presence of unidentified secondary structures or an additional level of regulation.
Deletion of the 5’ leader sequence, or disruption of the 5’ secondary structures, leads to constitutive expression or *erm* methylases such as *erm*(A), *erm*(B) and *erm*(C) [17,18]. To assess additional levels of regulation, bases +19 to +298 were deleted from the native *mef(E)/mel* locus and erythromycin resistance was determined. Erythromycin resistance of non-induced Δ+19–298 mutant, XB31 (Fig. 1C), was not different from the non-induced control strain XB29 (MIC, 12 μg ml⁻¹) (Table 2). However, resistance was not induced significantly by erythromycin induction (MIC, 16 μg ml⁻¹) which represented a significant change from the induced control (MIC, 64 μg ml⁻¹) (Table 2). This result was also not consistent with the simple attenuation model observed with *erm* methylases.

**Fig 6. Stem-loop R3/R4 is required for mef(E)/mel expression.** The predicted secondary structures of the Δ+162–189 mef(E)/mel regulatory region in (a) non-inducing and (b) inducing conditions. Disruption of R3/R4 by mutation was predicted to not disrupt the R5/R6 transcriptional terminator in inducing or non-inducing conditions, indicating that mutants with the Δ+162–189 deletion were uninducible. Annotations are consistent with those described in the Fig. 1C legend. Inducing conditions include growth with exposure to erythromycin at a concentration equal to 10⁻¹ the erythromycin MIC.

doi:10.1371/journal.pone.0116254.g006
Ribosomal stalling protects nascent transcripts from degradation by ribonucleases [19]. The deletion of XB31 removed mef(E)L, and thus ribosome stalling was not possible, suggesting that the XB31 mef(E)/mel transcript, though not attenuated, was unstable and quickly degraded, preventing over-expression of the efflux pump. To test this hypothesis, mutant XB38 was generated by deletion of bases +63–298. The mutation was identical to that found in XB31 except that the leader peptide and the putative ribosome stall site remained (Fig. 1C). The non-induced resistance of XB38 to erythromycin (MIC, 32 μg ml⁻¹) was 2.7-fold higher than the control XB29, and at least 4-fold higher after exposure to erythromycin (MIC, >264 μg ml⁻¹) (Table 2). These data indicated that the mef(E)/mel transcript was protected from degradation by ribosomes stalled during translation of the leader peptide. Further, 5’ RACE detected transcripts that begin with the +41C (Fig. 1A), located in the third codon of mef(E)L, or five codons from its 3’ end (see above). This was consistent with the reported endonuclease cleavage site of the erm(C) leader peptide erm(C)L which was shown to be protected by the stalled ribosome [19]. The data suggest that mef(E)L translating ribosomes stall on the last codon of mef(E)L and protect the transcript from further degradation.

Visualization of mef(E)/mel attenuation by RNA-seq

RNA-Seq whole transcriptome analyses were utilized to further determine if the mef(E)/mel transcript was terminated prematurely in non-inducing conditions. Fig. 7 shows the reads mapping to the mef(E)/mel operon viewed using the Integrative Genomics Viewer (IGV) (Broad Institute, Cambridge, MA). The tracks represent expression on the forward strand (reverse strand expression data not shown) under each experimental treatment. As expected, the mef(E)/mel operon was not expressed in untreated samples or in samples treated with the non-inducing macrolide spiramycin (Fig. 7). LL-37, a weak inducer of mef(E)/mel [9], also did not induce mef(E)/mel under these conditions. In the erythromycin-induced sample, mef(E) and mel were expressed as a single operon (Fig. 7). Interestingly, in the non-induced samples, a low level of expression was observed originating downstream of the transcriptional start site and terminating after approximately 250 bases. In the erythromycin-induced sample,
transcription began at the +1G and continued until the end of the mef(E)/mel operon (Fig. 7).
Similar expression patterns for mefE/mel expression (non-induced or induced with erythromycin) were observed in whole transcriptome analyses of three other Mega-containing invasive pneumococcal isolates, GA17545 (AFGA00000000.1), GA02254 (AIKI00000000.1) and GA41565 (AGPO00000000.1) (data not shown). RNA-Seq demonstrated transcriptional attenuation of mef(E)/mel in non-inducing conditions and transcriptional activation in the presence of the inducing macrolide.

Phylogenetic analyses of mef-containing mobile elements
As part of a comparative genomic study of the evolution and dissemination of antimicrobial resistance in a geographically defined population of S. pneumoniae, the genomes of 147 pneumococcal isolates, including 115 macrolide resistant invasive pneumococcal strains, were sequenced. Isolate metadata and the NCBI accession number for each genome are provided in S2 Table. Mega carrying mef(E) was present in 102 macrolide resistant isolates, including 20 that also contained erm(B) carried on Tn2010. Strain England14–9 (accession no. NZ_AILI00000000 contained mef(A)/msr(D) located on the mobile element Tn1207.1. All other isolates contained either erm(B) alone (n = 11) or 23S ribosomal mutations (n = 1). Type I Mega (5.5 kb) was found in 61 isolates, 49 isolates had the type 2 Mega (5.4 kb) [3] and a single isolate contained a 112 bp duplication of the 5′ leader sequence of mel that has not been previously described (data not shown). Of the 102 Mega sequences, 77 were identical and the remaining contained no more than three single nucleotide polymorphisms (SNPs). The exception was the original Mega sequence (AJ274302), which contained a 16 bp insertion that has been described in S. viridians (EF042094) and many mef-containing elements from non-pneumococcal streptococci.

Mef(E)/Mel-mediated resistance to erythromycin varied from 1 μg ml\(^{-1}\) to >64 μg ml\(^{-1}\) in the 102 Mega strains. However, phylogenetic analyses of the Mega sequence revealed no changes within the mef(E) or mel structural genes that correlated with the strain to strain variation in macrolide resistance, suggesting that variable resistance levels were due to differential expression of mef(E)/mel. Deletions in the mef(E)/mel 5′ regulatory region have not been reported and none were detected in the 102 Mega-containing isolates examined in this study.

Alignment of Mega with pneumococcal elements containing mef(A) (Tn1207.1) and mef(I) (5216IQ complex) using Clustal Omega (version 1.2.1) [20] revealed a 3.2 kb region of homology, “the mef cassette” including the 5′ regulatory region of mef(E) and mel (Fig. 1A). The mef cassette contained the -10 consensus sequence at its left junction and was inserted in all of the elements just downstream of the -35 box of the mef/mel(msr(D)) operons resulting in evolutionary divergence of the -10 and -35 promoter sequences (Fig. 1A). The promoters displayed some nucleotide similarity (approximately 65% identity), but the 327 bp 5′ regions of the mef(A) and mef(I) elements were 96% and 97% identical, respectively, to that found in mef(E) (Fig. 1A).

These data suggest an evolutionary history of inter- and intra-species horizontal transfer of the 3.2 Kb mef cassette.

A consensus attenuator structure was generated by Clustal Omega alignment of the 5′ regulatory mRNA region of these other mef-containing elements and prediction of consensus secondary structures performed by RNAfold (ViennaRNA Package 2.0 [11]). The mef-containing elements included the pneumococcal mef(I)- and mef(A)-containing elements (5216IQ complex and Tn1207.1, respectively), and elements from S. pyogenes, Clostridium kluvyerii, C. perfringens and Group G streptococci. The consensus structure was not significantly different from the mef(E)/mel attenuator structure (Fig. 3).
Further, the sequences of the homologous Rho-independent terminator from these elements were analyzed. The sequence for the predicted terminators of Mega, and Mega carried on Tn\textsubscript{2009} and Tn\textsubscript{2010} were identical. Other elements displayed one to four SNPs in the predicted terminator (Fig. 8). The \textit{mef}(A) elements and the \textit{mef}(I) element from the \textit{C. kluyverii} phage element contained SNP1 and SNP2, and also displayed SNP3 and/or SNP4 at nucleotides that were predicted to base pair in the \textit{mef}(E) terminator (Fig. 8). Both SNP3 and SNP4 were adenine substitutions of uracil, thus disrupting an A-T base pair and thus were predicted to result in decreased stability of the structures compared to Mega (Fig. 8).

The overall sequence conservation of the predicted Rho-independent transcriptional terminators suggested similar mechanisms of regulation the \textit{mef} and \textit{mel}(\textit{msr}(D)) genes in each of these elements. In support of this observation, the erythromycin MIC of strain England14

<table>
<thead>
<tr>
<th>Element \textsuperscript{a}</th>
<th>mef \textsuperscript{b}</th>
<th>Stem-loop sequence</th>
<th>ΔG (kcal mol\textsuperscript{-1}) \textsuperscript{c}</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mega</td>
<td>E</td>
<td>AUGCAAGACAAAGCCACAUUGUGGGGUUGGCGUCGCAU</td>
<td>-18.5</td>
</tr>
<tr>
<td>Tn2009</td>
<td>E</td>
<td>AUGCAAGACAAAGCCACAUUGUGGGGUUGGCGUCGCAU</td>
<td>-18.9</td>
</tr>
<tr>
<td>Tn2010</td>
<td>E</td>
<td>AUGCAAGACAAAGCCACAUUGUGGGGUUGGCGUCGCAU</td>
<td>-18.5</td>
</tr>
<tr>
<td>5216IQ</td>
<td>I</td>
<td>AUGCAAGACAAAGCCACAUUGUGGAUAUUGGCGUCGCAU</td>
<td>-18.9</td>
</tr>
<tr>
<td>Sdys\textsubscript{eq}</td>
<td>I</td>
<td>AUGCAAGACAAAGCCACAUUGUGGAUAUUGGCGUCGCAU</td>
<td>-18.9</td>
</tr>
<tr>
<td>Tn1207.1</td>
<td>A</td>
<td>AUGCAAGACAAAGCCACAUUGUGGAUAUUGGCGUCGCAU</td>
<td>-13.9</td>
</tr>
<tr>
<td>Tn1207.3</td>
<td>A</td>
<td>AUGCAAGACAAAGCCACAUUGUGGAUAUUGGCGUCGCAU</td>
<td>-13.9</td>
</tr>
<tr>
<td>Cklu Φ</td>
<td>I</td>
<td>AUGCAAGACAAAGCCACAUUGUGGAUAUUGGCGUCGCAU</td>
<td>-13.9</td>
</tr>
<tr>
<td>Φ10394</td>
<td>A</td>
<td>AUGCAAGACAAAGCCACAUUGUGGAUAUUGGCGUCGCAU</td>
<td>-13.9</td>
</tr>
<tr>
<td>Cper_IFI</td>
<td>A</td>
<td>AUGCAAGACAAAGCCACAUUGUGGAUAUUGGCGUCGCAU</td>
<td>-13.9</td>
</tr>
<tr>
<td>Φm46.1 like</td>
<td>A</td>
<td>AUGCAAGACAAAGCCACAUUGUGGAUAUUGGCGUCGCAU</td>
<td>-13.9</td>
</tr>
</tbody>
</table>

Fig 8. Comparison of the Rho-independent terminator in the Mega element in \textit{S. pneumoniae} with predicted terminators from mobile elements found in other Gram-positive bacteria species. The nucleotide sequence of the stem-loop portion (not including the poly-uracil tract) of the predicted Rho-independent transcriptional terminators are aligned and single nucleotide polymorphisms are shaded gray. The predicted free energy (ΔG) for each terminator is indicated The Rho-independent terminator structure from the Mega element is shown below in dot-bracket notation. Abbr., Mega (macrolide efflux genetic assembly), \textit{S. pneumoniae} GA17457; Tn2009, \textit{S. pneumoniae} GA08825 Tn916-like element (Genbank accession no. AILK01000006); Tn2010, \textit{S. pneumoniae} GA47628 Tn916-like element (accession no. AILC01000010); 5612IQ, \textit{S. pneumoniae} 5612IQ complex (accession no. AJ971089); Sdys\_eq, \textit{S. dysgalactiae} subspecies \textit{equismilis} G51 mef(A) element (accession no. AM168138); Tn1207.1, \textit{S. pneumoniae} England\textsuperscript{14–1} phage-related element (accession no. AIL10100002); Tn1207.3, \textit{S. pyogenes} 2812A phage-related element (accession no. AY657002); CkluΦ, \textit{Clostridium kluvyeri} DSM 555 phage-related element (accession no. CP000673); ΦMGAS10394, \textit{S. pyogenes} MGAS10394 phage-related element (accession no. CP000003); Cper\_IFI, \textit{C. perfringens} IFI mef(A) element (accession no. EU535439); \textit{S. pyogenes} MB56Spyo045 Φm46.1-like (accession no. JF501521); ΦM46.1, \textit{S. pyogenes} Φm46.1 (accession no. FM864213).

doi:10.1371/journal.pone.0116254.g008
containing \(mef(A)\) on Tn1207.1, was induced 8-fold by subinhibitory concentrations (1.0 \(\mu g\) \(ml^{-1}\)) of erythromycin, increasing from 20 \(\mu g\) \(ml^{-1}\) in the absence of erythromycin to 320 \(\mu g\) \(ml^{-1}\) when induced. These data suggest that transcriptional attenuation is the mechanism of repression/induction of efflux-mediated macrolide resistance in Gram-positive bacteria regardless of the \(mef\) allele and its associated mobile element.

**Discussion**

In this study the mechanism of repression and induction of the macrolide efflux operon \(mef(E)/mel\) encoded on the Mega mobile element in \(S.\ pneumoniae\) and other Gram-positive bacteria was defined. Like \(erm(B)\), \(mef(E)/mel\) is induced by the 14- and 15-membered macrolides such as erythromycin and azithromycin, but not by 16-membered macrolides such as spiramycin and midecamycin [21]. This suggested that the two-systems were controlled by similar mechanisms, that is, attenuation.

Attenuation of inducible \(erm\) genes typically occurs at the level of translation (reviewed in [7]) and is characterized by the presence of one or two small (8–20 amino acids) leader peptides located on the \(erm\) transcript 5’ of the start codon encoding the structural Erm protein. These leader peptides toggle the folding of the \(erm\) transcript between two alternate secondary structures; the attenuator that blocks expression of \(erm\), and the anti-attenuator that promotes expression. Ribosomes bound by an inducing macrolide pause at a programmed stall site on the leader peptide(s) and disrupt the attenuator, allowing formation of the anti-attenuator. Constitutively expressed \(erm\) genes typically have mutations in their 5’ regulatory regions which prevent formation of the attenuator structure [21]. In some \(erm\) macrolide-inducible systems, attenuation occurs at the level of transcription. This is similar to translation attenuation with the distinction that transcriptional attenuators form Rho-independent transcriptional terminators in non-inducing conditions leading to premature termination of transcription of the structural \(erm\) gene. Transcriptional attenuation has been observed for \(erm(D)\)-encoded methylases of the \(erm(D)\) class, which includes \(erm(D), erm(K)\) and \(erm(J)\) found in \(Bacillus\) species [15,16,22]. We have previously found that expression of \(mef(E)/mel\) is controlled at the level of transcription [4,8]. Thus, the hypothesis that \(mef(E)/mel\) was transcriptionally attenuated was explored.

The transcriptional start (+1) of the \(mef(E)/mel\) operon was mapped by 5’ RACE to a guanine nucleotide located 327 bp upstream of the \(mef(E)\) start codon. A consensus pneumococcal promoter sequence was identified upstream that included a pneumococcal extended-10 box and a-35 box. Deletion of the putative promoter abolished expression of a \(mef(E)-lacZ\) transcriptional fusion and rendered the mutant susceptible to erythromycin. Restoration of the mutated promoter to the wild type restored resistance to wild type levels confirming the location of promoter.

Analyses of the 327 bp 5’ regulatory region of \(mef(E)\) downstream of the promoter revealed a feature common to attenuators involved in the regulation of \(erm\)-type methylases, including a small \(orf\), \(mef(E)L\), encoding a small peptide (MTASMLRL) (Table 2) preceded by a consensus ribosomal binding site (RBS1). The molecular signals dictating programmed ribosome stalling involve sequence-dependent interaction between the nascent peptide, the peptidyl transferase center in the exit tunnel and the inducing macrolide molecule [23]. Macrolide-induced stalling at sites located on the leader peptides prevents the attenuator from forming in favor of the anti-attenuator to promote expression of the Erm methylase. The amino acid sequence of \(Mef(E)L\) was very similar to the leader peptide identified in other inducible macrolide systems, including \(Msr(A)L\) (MTASMLRLK) encoded upstream of \(msr(A)\) \(Staphylococcus\) and the transcriptionally attenuated \(erm(K)\) (MTHAMRLRFPLLNQ) and \(erm(D)\)
(MTHSMRLR) genes [15,16]. Significantly, the stall site on the \(\text{erm}(K)\) leader peptide \(\text{erm}(K)L\) is MRLR and the minimal Erm(K)L sequence that will allow induction of \(\text{erm}(K)\) is MTHAMRLR. These are nearly identical to the predicted sequence of the Mef(E)L peptide. We demonstrated that \(\text{mef}(E)L\) leader peptide was required for induced transcription of the \(\text{mef}(E)/\text{mel}\) operon and for efflux-mediated macrolide resistance. Translational inactivation by of \(\text{mef}(E)L\) ribosomal binding site by site directed mutagenesis and truncation of the protein resulted in macrolide susceptibility regardless of induction, consistent with the requirement for ribosomal stalling and anti-attenuation.

Transcriptional attenuators require one or more Rho-independent terminators 5’ of the induced structural gene. Stem-loop R5/R6 (Fig. 6) was identified as the terminator of the \(\text{mef}(E)/\text{mel}\) attenuator. It has a G+C rich stem, was predicted to be stable, and was immediately followed by a run of five uracil residues. Deletion of the structure significantly increased basal-level expression and resistance to erythromycin. Stem-loop R3/R4 was also required for the regulation of \(\text{mef}(E)/\text{mel}\) expression and we identified a pair of distal 17 base perfect inverted repeats in the \(\text{mef}(E)\) 5’ regulatory region separated by 174 nucleotides. The location of the inverted repeats within critical structures of the attenuator suggested involvement in the attenuation mechanism. Deletion of the first inverted repeat sequence, IR1, resulted in high-level expression, but abolished induction by erythromycin confirming a role in \(\text{mef}(E)/\text{mel}\) regulation.

The RNA-Seq data provided additional clarity and strong evidence for a transcriptional attenuation model and the induction of \(\text{mef}(E)/\text{mel}\) by anti-attenuation of transcription in the presence of inducing macrolides. In untreated and non-macrolide antibiotic-treated samples, the sequence reads mapped precisely to the transcriptional start site and terminated at the 3’ end of the Rho-independent terminator. In the macrolide-induced sample, the reads mapped to the start site and continued through \(\text{mef}(E)\) and \(\text{mel}\) structural genes. Our observations do suggest that control of \(\text{mef}(E)/\text{mel}\) is influenced by regulatory mechanisms in addition to transcriptional attenuation. We have previously reported that the antimicrobial peptide LL-37 induces \(\text{mef}(E)\) expression and increased resistance to erythromycin and that the mechanism of induction appeared to be distinct from that of macrolides [9]. Also, in some attenuator deletion mutants expression and resistance were still inducible by erythromycin indicating an additional level of control of the \(\text{mef}(E)/\text{mel}\) promoter. Further, multiple mechanisms have been reported controlling \(\text{erm}\) gene expression. In addition to attenuation, erythromycin-induced ribosomal stalling increases expression of \(\text{erm}\) genes by protecting transcripts from 5’-to-3’ nucleolytic degradation [24–26]. In preliminary experiments we have observed that in \(\text{mef}(E)/\text{mel}\) attenuator-deficient mutants, erythromycin resistance was reduced when \(\text{mef}(E)L\) was deleted as compared to when \(\text{mef}(E)L\) and the ribosome stall site were present. This suggests that erythromycin-mediated ribosome stalling may increase \(\text{mef}(E)/\text{mel}\) mRNA stability in a similar manner as reported for several \(\text{erm}\) genes. Another regulatory mechanism has recently been reported for \(\text{erm}(C)\) that involved macrolide-induced ribosomal frameshifting [27]. Frameshifting during translation of the leader peptide \(\text{erm}(C)L\) requires an telithromycin-bound ribosome and an \(\text{erm}(C)L\)”shift-prone” sequence which is essentially a run of four or more uracil or adenine residues. The ribosomal shift promotes translation of \(\text{erm}(C)\) by a mechanism that has yet to be determined. We have previously demonstrated that telithromycin induced \(\text{mef}(E)/\text{mel}\) expression and increased resistance to non-ketolide antibiotics [8]. The other potential regulatory mechanisms will be explored in future work.

There are other important differences between \(\text{erm}\) methylase and \(\text{mef}(E)/\text{mel}\) regulation suggesting that the systems are not completely analogous. We have shown previously that induction of \(\text{mef}(E)/\text{mel}\) by macrolides is dependent upon the identity of the amino sugar side chains of the macrolide ring [21]. Additionally, constitutively resistant (cMLS\(_B\) phenotype)
clinical isolates of many species have been identified and are readily selected in the laboratory setting ([17,28,29]. To date, there have been no reports of constitutively expressed mef-mediated macrolide efflux in clinical isolates or generated in vitro. This may suggest a severe fitness cost to unregulated expression of mef(E)/mel in contrast to the erm methylases.

The comparisons of the 5′ region of mef(E)/mel of Mega to mef-containing elements from pneumococci and other Gram-positive species revealed conservation of the 327 bp regulatory region and the predicted secondary attenuator structures. These data suggest that mef(E), mef(A) and mef(I) genes and the adjacent msr genes in pneumococci, other streptococci and other Gram-positive species, are likely responsive to macrolide induction and regulation by similar transcriptional attenuation. We verified that mef(A)/msr(D) carried on Tn1207.1 was induced by erythromycin. Thus, mef(E)/mel in *S. pneumoniae* is a model for inducible mef-mediated expression in a variety of Gram-positive pathogens.

**Supporting Information**

S1 Table. Nucleotide primers used in this study. (DOCX)

S2 Table. *Streptococcus pneumoniae* isolates sequenced in this study. a strain collection. Abbr., GA EIP, Georgia Emerging Infections Program; PMEN Pneumococcal Molecular Epidemiology Network; EIP, Emerging Infections Program (Centers for Disease Control, Atlanta, GA); Jain, Shabnam Jain (Emory University, b Locale, geographic location of isolation c Source, biological source of isolation d ST, multilocus sequence type e CC, clonal complex (XLSX)

**Acknowledgments**

We are grateful to Monica M. Farley, Wendy Baughman, Emily Crispell, and the Georgia Emerging Infections Program for providing the invasive pneumococcal isolates from Georgia for genome sequencing. We are also grateful to Leslie McGee, the Pneumococcal Molecular Epidemiology Network and the Centers for Disease Control Streptococcus Laboratory for providing invasive isolates and Pekka Nouri and Shabnam Jain for providing nasopharyngeal isolates.

**Author Contributions**

Conceived and designed the experiments: STC DSS. Performed the experiments: STC XB NK NS LS LJT TC SO. Analyzed the data: STC SD EFD HT DSS. Wrote the paper: STC HT DSS. Supervised sequencing and analysis activities: CMF.

**References**


18. de Vries LE, Christensen A, Agenso Y (2012) The diversity of inducible and constitutively expressed erm(C) genes and association to different replicon types in staphylococci plasmids. Mob Genet Elements 2: 72–80. PMID: 22934240


21. Chancey ST, Zhou X, Stephens DS. The Macrolide Efflux System Encoded by mef(E)/mef(m) in Streptococcus pneumoniae Is Induced by 14- and 15-membered, but Not 16-membered Macrolides; 2008 June 8–12; Reykjavik, Iceland.


27. Gupta P, Kannan K, Mankin AS, Vazquez-Laslop N (2013) Regulation of gene expression by macro-
PMID: 24239289

lates of streptococci and enterococci cross-resistant to erythromycin and lincomycin. J Antimicrob Che-
mother 43: 559–562. PMID: 10350387

tance patterns in macrolide-resistant *Streptococcus pyogenes* harbouring *erm(A)*. J Antimicrob Che-

in invasive pneumococcal disease. Journal of the Medical Association of Georgia 91: 20–23. PMID:
12436698

brial-Resistant Clones of *Streptococcus pneumoniae* Defined by the Pneumococcal Molecular Epidemi-

32. Sharma D, Baughman W, Holst A, Thomas S, Jackson D, et al. (2013) Pneumococcal carriage and in-
vasive disease in children before introduction of the 13-valent conjugate vaccine: comparison with the
0b013e3182788fdd PMID: 23060290


34. Dunny GM, Lee LN, LeBlanc DJ (1991) Improved electroporation and cloning vector system for Gram+

35. Frohman MA (1993) Rapid amplification of complementary DNA ends for generation of full-length com-


bridization array data repository. Nucleic Acids Res 30: 207–210. PMID: 11752295


genes from enterococci isolated from food. Syst Appl Microbiol 30: 1–7. PMID: 16563685


42. Gryczan T, Israeli-Reches M, Del Bue M, Dubnau D (1984) DNA sequence and regulation of *ermD*, a
macrolide-lincosamide-streptogramin B resistance element from *Bacillus licheniformis*. Mol Gen Genet
194: 349–356. PMID: 6429477

43. Darty K, Denise A, Ponty Y (2009) VARNAX Interactive drawing and editing of the RNA secondary struc-