Hidden Epidemic of Macrolide-resistant Pneumococci

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Community-acquired respiratory tract infections (RTIs) account for a substantial proportion of outpatient antimicrobial drug prescriptions worldwide. Concern over the emergence of multidrug resistance in pneumococci has largely been focused on penicillin-resistant *Streptococcus pneumoniae*. Macrolide antimicrobial drugs have been widely used to empirically treat community-acquired RTIs because of their efficacy in treating both common and atypical respiratory pathogens, including *S. pneumoniae*. However, increased macrolide use has been associated with a global increase in pneumococcal resistance, which is leading to concern over the continued clinical efficacy of the macrolides to treat community-acquired RTIs. We provide an overview of macrolide-resistant *S. pneumoniae* and assess the impact of this resistance on the empiric treatment of community-acquired RTIs.

### Macrolide Resistance

**Mechanisms of Resistance**

Macrolides are microbiostatic agents that reversibly bind to the 23S ribosomal RNA in the 50S subunit of ribosomes and block protein synthesis (8). Two main macrolide resistance mechanisms have been identified in...
pneumococci: active efflux of the drug from the cell and target-site modification (8). Energy-dependent efflux of macrolides from target cells by a cell membrane transporter has been associated with the presence of \textit{mef} genes. Recent work by Lannelli et al. (9) has implicated a second gene, \textit{mat(A)}, that encoded 2 ATP-binding domains, as a component of \textit{mef}-mediated macrolide resistance in pneumococci. Irrespective of the identity of the gene responsible for macrolide efflux, \textit{mef(A)}-positive \textit{S. pneumoniae} strains displaying this macrolide efflux phenotype (termed the M phenotype) are resistant to 14- and 15-membered ring macrolides (but not lincosamides or streptogramins) and generally display a low level of in vitro resistance to these antimicrobial agents. However, data from surveillance studies suggest that erythromycin MICs for \textit{mef(A)}-positive isolates may be increasing. MICs were 1–16 µg/mL for \textit{mef(A)}-positive \textit{S. pneumoniae} isolates collected from 1994 to 1995 (10), while results from a more recent study demonstrated an erythromycin MIC of 1 to >256 µg/mL (11).

The second major mechanism of macrolide resistance in streptococci, target-site modification, is predominantly encoded by the \textit{erm(B)} gene, resulting in methylation of an adenine residue on the 23S rRNA by a methylase enzyme. This methylation blocks the binding of macrolide-lincosamide-streptogramin B antimicrobial drugs. Strains with the macrolide-lincosamide-streptogramin B phenotype generally show higher levels of in vitro resistance to macrolides compared to strains with the M phenotype (10).

Other target-site modifications occur rarely in clinical isolates of \textit{S. pneumoniae}. These modifications include mutations that involve domain V of the 23S rRNA and genes encoding riboproteins L4 and L22 (12). Such mutations can confer resistance to macrolide-lincosamide-streptogramin B antimicrobial drugs and are associated with variable levels of in vitro resistance. Although the global prevalence of pneumococcal strains with macrolide resistance conferred by ribosomal gene mutations remains low (<2%), a study of macrolide resistance mechanisms among \textit{S. pneumoniae} isolated in Canada from 1997 to 2003 indicated that the rate of resistance due to mechanisms other than efflux or ribosomal methylation increased from 1% in 1997 to 10% in 2003 (13).

**Macrolide Resistance Trends**

A number of industry-sponsored global surveillance studies, such as the Alexander Project (GlaxoSmithKline) and PROTEKT (Prospective Resistant Organism Tracking and Epidemiology for the Ketolide Telithromycin) (sanofi-aventis), have been designed to define and monitor the prevalence and distribution of antimicrobial resistance among respiratory pathogens, detect new patterns of resistance, provide early warning of emerging resistance, and evaluate the effects of interventions aimed at reducing antimicrobial resistance. Results from the Alexander Project indicate that in 1996 and 1997 the global rate of pneumococcal macrolide resistance was 16.5%–21.9% (14); by 1998–2000, the resistance rate had increased to 24.6% (15). Data reported after completion of the first year of the PROTEKT study (1999–2000 respiratory season) confirmed this high global incidence (31.0%) of pneumococcal macrolide resistance (7), with similar overall levels of resistance among isolates collected as part of the PROTEKT US study (31.0% in 2000–2001 and 27.9% in 2001–2002) (16,17). The slight reduction in macrolide resistance among pneumococcal isolates collected as part of the PROTEKT US study from 2001 to 2002 may be a consequence of the February 2000 introduction of the 7-valent pneumococcal conjugate vaccine (18).

However, both macrolide resistance rates and resistance mechanisms may vary considerably depending on location. Macrolide resistance rates for isolates collected during the PROTEKT US study from 2001 to 2002 vary according to region; at a state level, the highest prevalence of pneumococcal macrolide resistance was recorded in Louisiana (48.2%) and the lowest in Vermont (15.2%) (11). Similarly, while \textit{mef(A)} was the most prevalent pneumococcal macrolide resistance genotype identified in the United States overall (68.7% of genotyped isolates), the relative prevalence varied by state and ranged from 40% in Delaware to 85% in Georgia (11). While \textit{erm(B)} was the second most prevalent genotype overall (16.8%), isolates possessing both the \textit{erm(B)} and \textit{mef(A)} genotype (12.2%) were more prevalent in 11 states than those harboring \textit{erm(B)} alone. A recent analysis of PROTEKT US 2002–2003 data by Farrell et al. (19) indicates an increase in the prevalence of macrolide-resistant isolates containing both \textit{erm(B)} and \textit{mef(A)} from 9.7% in 2000–2001 to 16.4% in 2002–2003. Most (99.2%) of these \textit{erm(B)} + \textit{mef(A)}-positive isolates were resistant to ≥2 classes of antimicrobial drugs. Analysis of erythromycin MIC data for all macrolide-resistant isolates collected in 2000–2001 indicated that the MIC$_{90}$ (MIC at which 90% of isolates were inhibited) varied according to resistance genotype (16 µg/mL for \textit{mef(A)}-positive isolates vs. >256 µg/mL for \textit{erm(B)}-positive isolates and those harboring both the \textit{erm(B)} and \textit{mef(A)} gene) (11).

**Factors Contributing to Development and Spread of Macrolide Resistance**

Inappropriate use of antimicrobial drugs is among the most important factors associated with the emergence and spread of pneumococcal macrolide resistance. Inappropriate use may include using antimicrobial drugs to treat noninfectious or self-limiting infections, using agents with a spectrum of activity that either does not cover the
appropriate causative pathogen(s) or which has too broad a spectrum of activity, and inappropriate dose or duration of treatment (20).

Other risk factors for carriage or infection with resistant pneumococcal strains include age (patients particularly at risk include those <2 or >65 years of age), history of macrolide use, and the presence of severe underlying disease (21). Analyses of data from national and international surveillance studies have suggested a link between increased use of macrolides and increased rates of pneumococcal resistance (22). In Portugal, the emergence of macrolide-resistant \textit{S. pneumoniae} strains from 1994 to 2002 correlated with the use of azithromycin during the same period (23). Several studies have shown that macrolide administration is associated with increased nasopharyngeal carriage of resistant strains of \textit{S. pneumoniae} in children (24); the clonal dissemination of macrolide-resistant pneumococcal strains in crowded environments (e.g., daycare centers, hospitals, jails, long-term care facilities) is also thought to be a major factor contributing to the spread of resistance.

**Clinical Implications of Macrolide Resistance**

Surveillance studies have shown that a substantial percentage of pneumococci are now macrolide resistant. Despite this rising rate of in vitro resistance, some researchers and clinicians have questioned whether resistance to macrolides is clinically relevant given the high concentrations achieved in respiratory tissues such as the epithelial lining fluid. Although macrolide levels in epithelial lining fluid have been reported to exceed the levels achieved in serum, the relevance of the fluid levels has been questioned (25), and sufficiently high macrolide blood levels remain essential to cure bacteremic pneumococcal pneumonia. Moreover, clinically achievable serum, epithelial lining fluid, and middle-ear fluid concentrations of azithromycin were insufficient to eradicate macrolide-resistant \textit{S. pneumoniae}, irrespective of the resistance mechanism (26).

Prospective clinical studies have provided conflicting evidence for an association between discordant antimicrobial therapy (i.e., use of an agent to which the causative pathogen displayed in vitro resistance) and treatment outcome. For example, while results from 1 study of patients with community-acquired pneumonia and bacteremia due to bacteremic pneumococcal infection demonstrated an association between increased death rates and discordant antimicrobial drug therapy (27), no such association was observed in a different study (28) of patients with pneumococcal community-acquired pneumonia. However, the conclusions that can be drawn from such studies may be limited by factors such as small sample size, differences in patient inclusion or exclusion criteria (e.g., recent antimicrobial drug use), use of relatively insensitive measures of treatment outcome (e.g., death), and use of single or multiple antimicrobial drugs (many hospitalized patients receive combination antimicrobial therapy, thus limiting the opportunities to study the effects of discordant treatment on clinical outcomes); in the studies cited above, none of the cases of discordant therapy involved monotherapy with a macrolide.

In acute otitis media, tympanocentesis performed before and after drug therapy has been used in several studies to determine the clinical relevance of antimicrobial resistance. Using this method, Dagan et al. (29) showed that microbiologic failure (correlated with clinical failure) was associated with pneumococcal macrolide resistance among patients treated with azithromycin; treatment of 6 of 6 patients with high-level macrolide resistance failed microbiologically. Furthermore, analysis of data from a pediatric medical center in the United States (30) noted that the rising incidence of antimicrobial-resistant pneumococci corresponded to an increase in supplicative complications of acute otitis media and appeared to contribute to more aggressive infections that required surgical intervention.

In recent years, several reports have described clinical and microbiologic treatment failures that have occurred in hospitalized patients infected with macrolide-resistant pneumococci (31). Among these cases of treatment failure, 2 deaths occurred. In both cases, the previously healthy patients (a 28 year-old man and a 49-year-old woman) received monotherapy with intravenous azithromycin for pneumonia. The clinical status of both patients deteriorated while they were receiving azithromycin, and macrolide-resistant \textit{S. pneumoniae} were isolated from blood and pleural fluid cultures taken while these patients were receiving medication.

A matched case-control study of hospitalized patients with bacteremia conducted by Lonks et al. (32) identified 86 patients with isolates of \textit{S. pneumoniae} that were fully or intermediate resistant to macrolides and 141 controls who had macrolide-susceptible pneumococcal infection. When patients with meningitis were excluded from the analysis, 18 (24%) of 76 patients were taking a macrolide at the time of bacteremia compared to none of the controls (p<0.0001). Moreover, 5 (24%) of the 21 bacteremic patients infected with pneumococci expressing the M phenotype were taking a macrolide (compared with none of the 40 matched control patients; p<0.0016). These data show that breakthrough bacteremia and treatment failure occurred only in those patients infected with a macrolide-resistant pneumococcus; no incidences of breakthrough bacteremia were seen in those infected with a macrolide-susceptible pneumococcus. Similarly, a study of all pneumococcal bacteremias from a hospital in Belgium (33)
showed that 4 (12%) of 33 patients with a macrolide-resistant pneumococcus were taking a macrolide when blood cultures were obtained, i.e., they had breakthrough bacteremia; in contrast, none of the 103 patients with macrolide-susceptible pneumococci was taking a macrolide.

The overall incidence of treatment failure caused by macrolide-resistant pneumococci cannot be estimated from the case reports and observational studies published to date. These reports of treatment failure likely only represent the tip of the iceberg, as most case studies published to date have only captured treatment failures that resulted in breakthrough bacteremia. These published studies underreport the magnitude of treatment failures because nonbacteremic pneumococcal pneumonia is 3–5 times more common than bacteremic pneumonia. In addition, these treatment failures resulted in hospitalization, which is more expensive than outpatient therapy. Most macrolides are prescribed as part of empiric treatment regimens for ambulatory patients in the outpatient setting; microbiologic cultures are not usually obtained from these patients, and antimicrobial susceptibility testing is rarely performed (even if treatment failure occurs).

**Macrolide Resistance and Treatment Guidelines**

In the United States, guidelines for the treatment of community-acquired RTIs have been established by a number of groups, including the American Thoracic Society, the Infectious Diseases Society of America, the Centers for Disease Control and Prevention (CDC), and the Sinus and Allergy Health Partnership. The clinical relevance of macrolide-resistant *S. pneumoniae* has been addressed in updates to these groups’ guidelines for the treatment of community-acquired pneumonia (34,35) and in a report published by the Drug-Resistant *Streptococcus pneumoniae* Therapeutic Working Group convened by CDC (36). The consensus among these guidelines is that empiric therapy should be stratified based on likely cause, treatment setting (inpatient versus outpatient), and the risk for pneumococcal antimicrobial resistance. In general, all 3 guidelines recommend that monotherapy with macrolides should be restricted to specific patient subgroups (i.e., those with no coexisting cardiopulmonary disease and no risk factors for infection with drug-resistant *S. pneumoniae* [e.g., recent antimicrobial drug use]). For outpatients with risk factors for drug-resistant *S. pneumoniae*, current recommended treatment options include combination therapy with a β-lactam (such as high-dose amoxicillin or high-dose amoxicillin-clavulanate) plus a macrolide or an antipneumococcal fluoroquinolone (34,35). The increased use of fluoroquinolones has been associated with the emergence and spread of resistance to these agents (37), and local clonal dissemination of *S. pneumoniae* strains with very high-level resistance to penicillin has been reported in the United States (38). Although the prevalence of these resistant isolates remains low, such findings emphasize the necessity for local resistance patterns to be considered when prescribing empiric antimicrobial drug therapy for patients with community-acquired RTIs.

The Sinus and Allergy Health Partnership guidelines for the treatment of acute microbial rhinosinusitis also highlight the need to consider the increasing prevalence of pneumococcal resistance when making treatment choices, with patients divided into categories dependent on their recent exposure to antimicrobial drugs (39). Similarly, a recent American Thoracic Society statement on the management of acute microbial exacerbations of chronic obstructive pulmonary disease emphasizes the need to consider local resistance patterns when prescribing antimicrobial drugs (40).

**Conclusions**

National and international surveillance studies demonstrate a high global prevalence of in vitro macrolide resistance among pneumococcal isolates obtained from patients with community-acquired RTIs. In recent years, a number of studies have clearly linked in vitro macrolide resistance to microbiologic and clinical treatment failure, indicating that macrolide resistance is an emerging problem. As pneumococcal community-acquired RTIs (particularly community-acquired pneumonia) are a leading cause of illness and death worldwide, appropriate empiric antimicrobial therapy should be used to treat these infections. Recent updates to a number of treatment guidelines have reflected this changing situation by emphasizing the need for clinicians to consider local antimicrobial resistance patterns and risk factors for infection with drug-resistant pathogens when prescribing empiric antimicrobial therapy.

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References


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