Genetic Interactions between the *Drosophila* Tumor Suppressor Gene *ept* and the *stat92E* Transcription Factor

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Abstract

**Background:** Tumor Susceptibility Gene-101 (TSG101) promotes the endocytic degradation of transmembrane proteins and is implicated as a mutational target in cancer, yet the effect of TSG101 loss on cell proliferation in vertebrates is uncertain. By contrast, *Drosophila* epithelial tissues lacking the TSG101 ortholog erupted (*ept*) develop as enlarged undifferentiated tumors, indicating that the gene can have anti-growth properties in a simple metazoan. A full understanding of pathways deregulated by loss of *Drosophila ept* will aid in understanding potential links between mammalian TSG101 and growth control.

**Principal Findings:** We have taken a genetic approach to the identification of pathways required for excess growth of *Drosophila* eye-antennal imaginal discs lacking *ept*. We find that this phenotype is very sensitive to the genetic dose of *stat92E*, the transcriptional effector of the Jak-Stat signaling pathway, and that this pathway undergoes strong activation in *ept* mutant cells. Genetic evidence indicates that *stat92E* contributes to cell cycle deregulation and excess cell size phenotypes that are observed among *ept* mutant cells. In addition, autonomous Stat92E hyper-activation is associated with altered tissue architecture in *ept* tumors and an effect on expression of the apical polarity determinant crumbs.

**Conclusions:** These findings identify *ept* as a cell-autonomous inhibitor of the Jak-Stat pathway and suggest that excess Jak-Stat signaling makes a significant contribution to proliferative and tissue architectural phenotypes that occur in *ept* mutant tissues.

Introduction

The *Drosophila* gene *erupted* (*ept*) encodes an ortholog of human Tumor Susceptibility Gene-101 (TSG101) and yeast Vps23p, which function as part of the ESCRT (endosomal sorting complex required for transport)-I complex to sort proteins to the multivesicular body for ultimate lysosomal degradation [1–3]. Mutations in the *ept/tsg101* gene (referred to hereafter as *ept*) or the ESCRT-II subunit gene *vps25* block endocytic degradation of certain transmembrane proteins and lead to dramatic overgrowth of imaginal discs [4–7], a set of polarized epithelial tissues that grow during larval stages and go on to form the majority of adult structures. This imaginal disc overgrowth occurs in part by a non-cell autonomous process: *ept* and *vps25* mutant cells themselves undergo very high rates of apoptosis, but can potently induce growth of surrounding genetically normal tissue due to overproduction of secreted factors like Unpaired (Upd) [4–6], a secreted cytokine-like mitogen that activates Jak-STAT signaling via the receptor Dorsal (Dome) [8]. Because the death of *ept* and *vps25* mutant cells eventually limits Upd production, the phenotypic outcome of clonal loss of either gene is limited to enlargement of the affected organ. However, both *ept* and *vps25* also display a conditional, cell-autonomous tumor suppressor activity: if the death of mutant cells is prevented, either by expressing of anti-apoptotic genes or by removing competing normal cells from the disc, they overgrow into large, disorganized tumors that kill the host animal [4–6]. Thus *ept* and *vps25* behave both as non-cell autonomous growth suppressors, and as conditional, cell-autonomous neoplastic tumor suppressors.

The vertebrate *ept* gene homolog, TSG101, was first identified based on its ability to inhibit cell transformation [9] and by its apparent mutational inactivation in some cancers [10]. Later studies were unable to confirm key elements of these data [e.g. 11,12–14], and subsequent analysis of mice carrying a targeted deletion of the TSG101 gene found no effect on cancer incidence or progression but did reveal a requirement for embryonic viability [15,16]. More recent work showing that the TSG101 gene is overexpressed in some human cancers [17–19] and that transgenic overexpression of TSG101 in the mouse mammary gland mildly increases the frequency of breast carcinoma [17] tends to support the idea that excess TSG101 promotes, rather than inhibits, cell survival and proliferation. Since signaling via the EGF receptor,
which is sorted to the lysosome via an ESCRT-dependent pathway in mammalian cells [20–23], is elevated in cells that overexpress TSG101 [17], excessively high levels of TSG101 may be capable of acting in a dominant-negative manner to reduce endocytic degradation of its normal targets thereby enhancing cell proliferation. A similar model has been proposed to explain a block in trafficking induced by overexpression of Vps18, a component of the mammalian Class C Vps/HOPS complex that interacts physically with TSG101 [24]. However, because genetic ablation of murine TSG101 does not produce hyper-proliferative phenotypes in vivo, the growth regulatory role of mammalian TSG101 remains uncertain.

The somewhat discordant views of the growth regulatory properties of TSG101-like proteins provided by mammalian and insect models suggest either that TSG101 is required for the endocytic degradation of a distinct set of receptors and membrane-associated proteins during vertebrate and invertebrate development, or that similar proteins enter the ESCRT pathway in each organism but other factors render Drosophila epithelia more susceptible to transformation by reduced ESCRT function. In order to lay the foundation for comparative analysis of vertebrate and invertebrate TSG101 developmental phenotypes, it is necessary to establish how "ept" mutations affect pathways that control cell proliferation and tissue growth. Here we show that the autonomous growth of ept mutant eye-antennal tumors is dependent on the stat92E gene, which encodes the sole fly member of the Signal Transducer and Activator of Transcription (STAT) family of mammalian transcription factors that are well known for their ability to promote tissue growth in Drosophila [25–27] and mammals [reviewed in 28]. Removing a single copy of stat92E gene significantly reduces the overgrowth of eye-imaginal discs composed entirely of ept cells. At a cellular level, this suppression by stat92E heterozygosity correlates with restoration of more normal G1/S cell cycle phasing in mutant discs and a partial reversion of an ept enlarged-cell phenotype. These effects on the proliferative properties of ept cells are accompanied by an unexpected ameliorating effect of stat92E heterozygosity on the epithelial architecture of ept mutant tissues. We find that lowering stat92E gene dosage reduces expression of the epithelial polarity factor Crumbs (Crb) in ept mutant eye discs. As excess Crb is sufficient to disrupt the architecture and polarity of epithelial tissues [29,30], stat92E may thus contribute to ept tissue architectural phenotypes by affecting Crb expression (as we note above, an allele of crb failed to obviously alter the ept tumor phenotype; MMG and KHM, unpub.). To some degree the genetic evidence of a role for stat92E in ept phenotypes is not surprising; the excess Upd produced by ept mutant cells is known to drive Stat92E-dependent proliferation in immediately surrounding cells. However it has not been determined whether excess Stat activation contributes to the growth and polarity phenotypes of ept mutant cells themselves. We find that multiple sensors of Jak-Stat activity detect strong activation of the Jak-Stat pathway within mutant cells and that this phenomenon is coincident with accumulation of the Upd receptor Dome in intracellular puncta in ept mutant cells that co-stain with the endosomal protein Hrs [31]. Stat92E hyperactivity thus correlates with an autonomous effect of ept alleles on Dome localization and levels. In sum, these data indicate that ept is a cell-autonomous inhibitor of Jak-Stat signaling in the larval eye disc, and that elevated Stat activity contributes to deregulated cell cycling, excessive cell size, and defects in tissue organization observed in ept mutant tumors. These data provide a link between a TSG101-like protein and a pathway that controls cell proliferation and tissue architecture in a simple metazoan.

Results

Genetic requirement for stat92E in ept tumors

Eye-antennal discs were engineered to lack ept function by the eyFLP,Minute (Min) technique. In this configuration, mitotic recombination produces Min/Min cells that die and ept mutant cells that populate the disc and structures derived from it. These discs (hereafter referred to as "ept(M/3)") overgrow into large unstructured tumors [5] (Fig. 1, compare A vs. B), indicating that loss of the gene can bypass normal limits on organ size. To test the genetic requirements of this tumor-like phenotype, we screened a small collection of alleles of signaling, polarity, and growth regulatory genes (stat92E, crb, lgl, Drosophila aPKC, yki, cyclinD, dMyc, stk, and others) for their ability to suppress size and/or architectural phenotypes associated with loss of ept/tsg101. A single copy of the stat92E06346 los-of-function allele [32] was found to significantly reduce the size of ept eye-antennal tumors (Fig. 1C) and cause them to grow as two distinct tissue lobes (yellow arrows in Fig. 1C) rather than the large mass of tissue that otherwise characterizes ept mutant eye-antennal discs. This effect on organ size and architecture is completely penetrant, and is

Figure 1. stat92E promotes growth of ept tumors. Bright-field images of (A) control discs [FRT80B/M(3)], (B) ept mutant discs [ept/M(3)], or ept mutant, stat92E heterozygous discs [ept/M(3),stat92E06346/+] from wandering-stage larvae. Yellow arrows in (C) denote the two lobes of tissue resembling eye discs. (D) Flow cytometric analysis of control (grey fill), ept(M/3) mutant (black line), and ept/M(3),stat92E06346/+ (dotted line) eye-antennal discs shows that reducing stat92E gene dosage partially rescues of cell cycle and cell size (inset) defects in ept/M(3) mutant tissues. Percentages of cells in each stage of the cell cycle are indicated. The FACS data are representative of multiple experiments.

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accompanied by a 15-fold increase in the frequency of pupation (0.4% of ept/3) animals [n = 243] vs. 6% of ept/M(3),stat92E06346/+ animals [n = 186]). To determine the effect of stat92E heterozygosity on the proliferative properties of cells within ept tumors, it was necessary to first establish the effect of ept loss on larval cell division and growth. Flow cytometric analysis indicates that cells within ept mutant tumors are enlarged relative to control cells (inset in Fig. 1D, black line vs grey fill) and show an increase in the percentage of cells in the S- and G2/M-phases of the cell cycle compared to cells in control FRT80B/M(3) discs (Fig. 1D, black line vs. grey fill). At a cellular level, the stat92E06346 allele slightly reduces the size of cells within these tumors while increasing the fraction of G1 cells and decreasing the fraction of G2/M cells (Fig. 1D and inset; dotted line vs. black line). Thus a diploid dose of the stat92E gene is required for the effect of ept loss on G1/S cell cycle phasing and to a somewhat lesser degree for the enlarged size of ept mutant imaginal disc cells. As cells in ept tumors are highly proliferative [5], the former effect suggests that stat92E promotes S-phase entry in ept mutant cells. This conclusion agrees with published data showing that overexpression of the mitogen Upd in the eye disc increases the number of cells in S-phase [25].

Stat92E promotes overgrowth and architectural disorganization of ept tumors

Mutations in ept or ept25 have previously been shown to alter epithelial polarity and tissue architecture [4–6]. The ability of the stat92E06346 allele to restore a more normal morphology to ept mutant eye-antennal discs led us to examine the epithelial organization of these tissues. Co-staining for the basolateral membrane marker Discs large (Dlg) and the apical membrane determinant Crumbs (Crb) reveals that ept mutant eye-antennal discs are composed of compacted sheets of cells (Fig. 2A-A*) that form multilobular, ‘pouched’ structures with their apical surfaces oriented inward toward an internal lumen (examples denoted by yellow asterisks in Fig. 2A*). These structures are very much different from the normal monolayer organization of the peripodial and disc epithelium seen in control discs (Fig. 2E). Using the location ofDlg protein to trace the apical and basal surfaces of these ept/M(3) tissues highlights this phenotype (Fig. 2A*). These architectural changes occur in parallel to a build-up of Crb protein in the cytoplasm of ept mutant cells (Fig. 2A*) due to a requirement for ESCRT complexes in vesicular trafficking of Crb [4,5,30]. Instead of concentrating at the apical surface of the tissue as in a control disc (Fig. 2E), Crb protein in ept/M(3) tumors is found more dispersed throughout the tissue (Fig. 2A*). A high magnification view of one of the ‘pouched’ structures (boxed in Fig. 2A) shows that ept is also required to prevent the spread of Crb from its normal location on the apical membrane onto the Dlg-positive basolateral domain (Fig. 2C-C*; arrowheads mark overlap of anti-Crb and anti-Dlg signals).

Tissue architecture was also analyzed in ept mutant discs also carrying a single copy of the stat92E06346 allele. In this genotype, the multi-lobular ‘pouched’ structures do not form and the main masses of tissue (asterisks in Fig. 2B*) appear fairly solid with their apical, Crb-positive faces oriented outward (compare Fig. 2A* to 2B*). Tracing the apical surface of these tissues emphasizes this architectural change (Fig. 2B*). In addition, Crb aggregates are less abundant and Crb is more concentrated on the outer edge of the epithelium (compare Fig. 2A’ to B’). It is not clear whether these apparent effects on Crb levels and localization are independent, or whether a reduction in Crb levels leads to a less severe mislocalization phenotype. However, the membrane-associated pool of Crb in ept,stat92E/+ mutant cells is still be detected overlapping the Dlg-positive basolateral domain (see arrowhead Fig. 2D-D*). Thus, a full genetic dose of the stat92E gene is required for the disruptive effect of ept loss on epithelial organization, and this appears to correlate with a requirement for stat92E in controlling Crb in ept mutant cells. By contrast, the mislocalization of Crb onto the basolateral membrane of ept mutant cells is either stat92E-independent or less sensitive to a two-fold reduction of stat92E dosage.

In view of the effect of the stat92E genotype on anti-Crb staining phenotypes in ept eye-antennal tumors, we examined the relationship between stat92E gene dosage and crb mRNA levels. In situ hybridization with a crb specific probe detects ubiquitous crb expression in control mosaic eye discs (Fig. 3C) and in ept mosaic eye discs (Fig. 3D). In some cases, small patches of cells that appear to contain high levels of crb mRNA are detected in ept mosaic discs (arrowheads in Fig. 3D), although this effect is difficult to visualize against the high level of crb expressed in surrounding normal cells. Direct analysis of the level of crb transcript levels by quantitative realtime RT-PCR indicates that crb mRNA is induced ~1.7-fold in ept mutant eye-antennal discs relative to control discs, and that this is suppressed by introduction of a single copy of the stat92E06346 allele (Fig. 3A). Although these data indicate that stat92E is upstream of crb transcript levels in ept mutant cells, stat92E may not normally be required for crb expression in normal eye cells since Crb protein levels were unaffected in clones of eye disc cells homozygous for the stat92E06346 allele (data not shown). However, crb transcript levels are markedly reduced in stat92E06346 mutant embryos generated using the germline clone technique [33] (Fig. 3F–G). This link between stat92E and crb in the embryo could be due to indirect effects of stat92E loss on tissues that normally express crb. Yet the data linking stat92E and crb in ept tumors it is also quite consistent with the findings that dome function is required for Crb expression in ovarian follicle cells [34] and that Jak-Stat signaling is both necessary and sufficient for expression of crb in the posterior spiracles [35]. Loss of ept may thus affect Crb by two distinct mechanisms: first by a stat92E-dependent effect on crb mRNA levels, and second by defective endocytosis that prevents Crb protein degradation and sequesters it in the cytoplasm. Although reducing crb gene dosage in the background of ept/M(3) discs did not overly affect tumor size or structure (data not shown), the role of Crb in the ept growth and tissue architectural phenotypes remains unclear.

Activation of Jak-Stat signaling in ept mutant cells

Stat92E has been shown to be required for the non-autonomous pro-growth effect of ept mutant cells on surrounding normal cells [5]. The genetic requirement for stat92E in the overgrowth of ept mutant tumors indicates that the gene is also required for the autonomous growth effects of ept alleles. The mechanism of this effect is not known. ept mutant cells are known to overexpress the upd gene [5], which encodes a secreted cytokine-like ligand protein that binds the Dome receptor and signals through the Jak kinase to the Stat92E transcription factor [reviewed in 36]. As upd overexpression is alone sufficient to drive eye-antennal disc enlargement [25], high levels of extracellular Upd produced by ept cells might be predicted to feed back onto nearby cells (irrespective of their genotype) to elicit the growth and proliferation phenotypes observed in FACS analysis (see Fig. 1). To test the effect of ept alleles on Stat92E signaling, we first used an antibody reported to detect ligand-stimulated tyrosine phosphorylation of Stat92E by Jak kinase (anti-pY-Stat92E) which is necessary for Stat92E activity in vivo [reviewed in 37]. This antibody has been used in other studies of Drosophila Stat92E activity [7,38], including one in which it was used to assess Jak-Stat activity in eyes disc mosaic for an allele of the ESCRT-II
Figure 2. stat92E affects tissue architecture in ept tumors. Confocal images of ept/M(3) (A-A’), and ept/M(3),stat92E06346/+ (B-B’) eye discs stained for Dlg (red), Crb (blue) and DNA (green). The two main tissue lobes in panel B are separated by the esophagus (e), which remained embedded in the tissue mass during dissection. Areas outlined by dashed boxes in (A) and (B) are magnified in (C-C’) and (D-D’) respectively. Asterisks in (A’), denote internal lumens bounded by Dlg-negative apical membrane of ept/M(3) mutant cells. Asterisks in B-B’ denote tissue lobes of ept/M(3),stat92E06346/+ discs. Note the dominant effect of stat92E06346 on the appearance of Crb aggregates (A’ vs B’) and the lack of the ‘inverted’ tissue phenotype in ept/M(3),stat92E06346/+ tumors (A’’ vs B’’). The stat92E06346 allele does not have a dominant effect on the mislocalization of Crb onto the Dlg-positive basolateral membrane domain of ept’’ mutant cells (arrowhead in D-D’). (E) A control FRT80B/M(3) disc stained for Crb (blue), Dlg (red), and DNA (green). The peripodial cell layer (PP) and disc proper (DP) are indicated. Panels A’’ and B’’ are tracings of the apical (green) and basal (white) membranes of the tissues in A’ and B’ respectively.

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component vps25 [7]. In eye-antennal discs composed of patches of normal and ept mutant cells, the anti-pY-Stat92E antibody detects strong accumulation of the pY-Stat92E epitope within clones of ept mutant cells (Fig. 4A–A'). A high magnification view of a clonal boundary confirms this effect (Fig. 4B–B'). This epitope is also elevated in mutant areas of ept disc tumors relative to wild type areas of FRT80B/M(3) control discs (Figs. 4C vs. D). To test the relationship between ept and Jak-Stat activity further, two Stat92E reporter transgenes were placed in the background of ept mutations. The first, 3xGAS-lacZ [39], is located on the same chromosome arm as ept, such that mitotic recombination of a 3xGAS-lacZ-ept transgene produces clones of ept cells carrying two copies of the 3xGAS-lacZ transgene that can be compared to discs carrying control FRT80B clones with two copies of 3xGAS-lacZ. When these cells are imaged under identical optical settings (Fig. 5A–B), the 3xGAS-lacZ reporter is up-regulated in patches of ept eye disc cells (Fig. 5A–A') relative to control FRT80B clones (Fig. 5B–B'). The inset in Fig. 5A shows a small ept clone in the posterior region of the eye disc that stains brightly for β-galactosidase expressed from the 3xGAS-lacZ transgene. This small-clone phenotype is due to increased rates of apoptosis (Figure S1, and see [5]), which make it difficult to recover large ept mutant clones in the eye-antennal disc. To bypass this problem, expression of the second Stat92E reporter, 10xStat92E-GFP [40], was analyzed in two backgrounds in which the death of ept mutant cells was blocked. The first of these was ept/M(3) tumors, in which removal of competing normal cells allows mutant cells to overgrow into large tumors [5]. 10xStat92E-GFP is very strongly expressed in this background relative to its expression in control FRT80B/ M(3) discs composed of normal cells (Fig. 5F–G). The magnitude of the difference is so substantial that the fluorescent signal from the disc in Fig. 5F was only detectable following a doubling of the ‘gain’ setting used to visualize the disc in Fig. 5G. 10xStat92E-GFP was also analyzed in a second background in which the death of ept mutant cells was blocked by simultaneous loss of the H99 chromosomal region, which contains genes required for apoptotic cell death [41]. We first confirmed that loss of the H99 region alone had no effect on expression of 10xStat-GFP (Fig. 5C). By contrast, expression of 10xStat-GFP is very strongly up-regulated in eye-antennal discs carrying clones of ept/H99 cells (Fig. 5D). An interesting pattern of GFP expression was observed in this experiment: some clones of ept/H99 mutant cells located in the eye disc express 10xStat-GFP highly both within clonal boundaries and in surrounding cells (see arrow in 5D' and D''), whereas other nearby eye clones do not (see arrowhead in 5D' and D''). This difference appears to be due to anterior/posterior positioning: clones that activate 10xStat-GFP tend to be located anterior to the presumptive morphogenetic furrow, whereas those that do not tend to be located in posterior regions of the disc. Autonomous and non-autonomous activation of 10xStat-GFP by ept/H99 clones is also observed in the antennal disc (Fig. 5D–E). 10xStat-GFP can be strongly activated both within clones and in cells located 5–10 cell diameters away from the mutant clones (Fig. 5E–E', arrows and white outlines). This evidence of non-autonomous 10xStat-GFP expression fits very well with previous models in which excess Upd produced by ept or vps25 mutant cells is able to activate Stat in surrounding cells [5]. The more novel observation of autonomous 10xStat-GFP activation within ept/H99 mutant clones agrees with the data gathered with the pY-Stat92E and 3xGAS-lacZ reporters. The differences in the readouts provided by these three pathway reporters also suggest that 10xStat-GFP may be a more faithful reporter of Stat92E activity in larval discs than either pY-Stat92E or 3xGAS-lacZ. In aggregate, these observations indicate that the effect of stat92E heterozygosity on ept tumor growth is a reflection of a role for Jak-Stat hyperactivity in the growth, cell cycle, and polarity characteristics of ept mutant cells.

Cell-autonomous effect of ept loss on Dome

The autonomous effect of ept loss of pY-Stat92E suggests that this phenotype is not only an indication of Jak-Stat activation, but may also reveal a requirement for ept in controlling an intracellular step in the Jak-Stat cascade. Consequently, we sought to test whether ept loss effects trafficking of the transmembrane receptor Dome. Dome traffics through the late endosome [34], and loss of the hrs gene, which acts at the step immediately preceding ept [reviewed in 42], blocks Dome trafficking and activation even in the presence of Upd [43]. These observations have led to the proposal that movement of Dome into and through the endosomal system is a significant regulatory step in Jak-Stat signaling [43]. Since loss of ept leads to accumulation of certain apical transmembrane proteins in the late endosome [3], we tested whether ept...
loss might also affect levels or localization of Dome. An antibody specific to the Dome protein [34] detects much higher levels of Dome in ept cells than in surrounding normal cells (Fig. 6A-A'). This Dome appears as puncta that partially co-localize with the endosomal protein Hrs (see arrowheads in Fig. 6). Loss of ept may also have more mild non-autonomous effects on Dome (see cells to the right of the clone in Fig. 5A). Since Hrs-dependent movement of Dome into the late-endosome has been proposed to be required for activation of Stat92E [43], we next tested whether the accumulation of the pY-Stat92E epitope in ept mutant cells was dependent on Dome activity. The Actin>CD2>Gal4 “flip-out” chromosome [44] was used in combination with a UAS-GFP transgene and a transgene carrying a dominant-negative form of dome that lacks the C-terminal tail (UAS-dome<sup>DCYT</sup>) [45] to produce GFP-positive/dome<sup>DCYT</sup>-expressing clones in the background of an ept tumor. ept mutant cells that express the dome<sup>DCYT</sup> allele (GFP-positive area in Fig. 6B-B’') do not display excess anti-pY-Stat92E staining, whereas those that do not express dome<sup>DCYT</sup> (GFP-negative area in Fig. 6B-B’') retain high levels of the pY-Stat92E epitope. From these data, we conclude that ept loss alters Dome localization and levels in eye imaginal disc cells and this correlates with Dome-dependent accumulation of the pY-Stat92E epitope. This data agrees with a proposed model in which Dome must access the Hrs-positive late endosome in order to activate signaling [43]. The trapping of Dome in this ‘activation’ compartment in ept mutants may therefore contribute to high-level activation of the Jak-Stat pathway observed in these cells.

Discussion

We have sought to identify pathways that mediate the cell-autonomous growth suppressor activity of the Drosophila endocytic gene ept, which encodes a homolog of mammalian Tumor Susceptibility Gene-101. We find that ept is required in vivo to restrict cell-autonomous activation of the Jak-Stat pathway, a well-established oncogenic pathway in mammals. This correlates with effects of the Drosophila stat92E gene on G1/S cell cycle control, cell size, and epithelial organization of ept mutant tumors. We also find that trafficking of Dome, which acts upstream of Stat92E, is altered in ept mutant cells. A previous study in the cultured Drosophila hemocyte S2 cell line also identified ept as a negative regulator of Jak-Stat signaling [46], but the extent to which this relationship is conserved in developing tissues in the whole organism and its contribution to ept loss-of-function phenotypes were not addressed. The data presented here suggest that excess Stat92E activity contributes to deregulation of the G1-to-S phase
cell cycle transition and promotes growth of ept mutant cells and tissues. We also find evidence that stat92E promotes accumulation of the crb mRNA in ept mutant cells, and that this correlates with an effect of stat92E gene-dosage on Crb and epithelial architecture in ept mutant tumors. These findings, and those of an accompanying study, are summarized in Figure 7.

Since Upd can stimulate the endocytic uptake of Dome [34], the effect of ept loss on Dome protein could theoretically be a secondary consequence of the fact that these cells express highly elevated levels of Upd. In addition, it has been shown that dome itself is a transcriptional target of Stat92E, as part of a positive feedback loop [23]. Alternatively, the relationship between ept and Dome localization could indicate a direct requirement for ept in Dome endolysosomal trafficking in much the same way that ESCRT mutants block the vesicular movement and lysosomal turnover of the Notch receptor [4–6,47]. The simple observation that Dome can be trapped in an Hrs-positive compartment agrees with other studies that have shown that the Dome receptor also fluxes through the ESCRT endosomal system in imaginal disc and ovarian follicle cells, and that endocytic trafficking of Dome can modulate the output of the downstream Jak-Stat pathway [34,43]. The oncogenic properties of this pathway are well-established in flies [25,27,48–50] and mammals [reviewed in 28], but its pro-growth targets are not fully understood. The data presented here support a role for Stat92E in promoting cell division and to a lesser degree cell size in ept mutant eye disc cells. A genetic screen for suppressors of a large-eye phenotype produced by misexpression of upd identified multiple components of the dpp (TGFβ) pathway [25], suggesting that Jak-Stat promotes growth via dpp signaling. However in the same study, activation of the dpp pathway was insufficient to rescue the small eye size of upd mutants, suggesting that additional pathways are responsible for Upd-dependent organ growth. One of these may involve Cyclin D, the cell cycle regulatory molecule that can promote tissue growth in Drosophila [51,52] and cancer in mammals [reviewed in 53]. Mammalian STAT5 is known to bind and transactivate the CyclinD1 promoter [54], and more recently misexpression of upd has been found to induce cyclin D in the Drosophila eye [27]. These findings offer a potentially more direct link between Jak-Stat and cell proliferation control in imaginal disc cells and provide a basis for further studies of growth pathways activated in ept tumors.

The Drosophila and mammalian forms of TSG101 are quite similar at a primary sequence level (46% identical/61% similar), share the same domain structure [5], and are predicted to have very similar molecular properties. Each has also been shown to function as part of the same conserved complex, ESCRT-I, and to be involved in the same biological process: endocytic trafficking of internalized receptors and membrane proteins. Thus the observed differences in the phenotypes elicited by loss of vertebrate and invertebrate TSG101 are likely to arise either due to 1) differences in the strength of alleles used in each system, 2) differences in the spectrum of proteins routed into the ESCRT pathway in each type of organism. In addition, TSG101 does not appear to be a member of a multi-gene family in vertebrates. Differences in cellular phenotypes produced by invertebrate and vertebrate ept/TSG101 alleles thus may reflect stage- or tissue-specific differences in the spectrum of proteins routed into the ESCRT pathway in each type of organism. In addition to Crb and Notch, Drosophila ept has now been shown to affect localization and levels of the Dome receptor (see model in Fig. 7). Loss of the ESCRT-II subunit and tumor suppressor gene

**Figure 5. Stat92E sensor activity in ept mutant eye-antennal cells.** Eye discs carrying clones of eptmutant cells (A’-A”) or FRT80B control cells (B-B”) marked by the absence of GFP (green) and stained for expression of β-galactosidase (β-gal; red in A’ and B’). Expression of the 10xStat-GFP reporter (green) in eptX1,H99 mosaic discs in which mutant cells are marked by the absence of β-gal (red). Arrowhead marks clone of eptX1,H99 mutant cells in the eye disc that do not activate 10xStat-GFP; arrow marks an example of an antennal clone that activates 10xStat-GFP within the clone and in surrounding wild type cells. Images in (E-E”) are of the antennal region of an eptX1,H99 mosaic disc. Expression of the Stat reporter 10xStat-GFP (green) in a control FRT80B/M(3) eye disc (F) and an eptmutant eye-antennal tumor (G). The disc in (F) was imaged at half the fluorescence intensity relative to the control disc in (G). doi:10.1371/journal.pone.0007083.g005
Figure 6. Dome localization in ept mutant tissue. (A-A') A clone of ept mutant eye disc cells marked by the absence of GFP (green) stained for Dome (blue) and the endocytic marker Hrs (red) shows extensive accumulation of Dome in Hrs-positive structures (yellow arrowhead denotes example of magenta overlap). (B-B') A confocal image of a section of an ept eye-antennal tumor expressing dome (using the eyFLP;Actin->CD2->Gal4, UAS-GFP system) and stained for anti-pY-Stat92E (blue); GFP (green) marks cells that express the dome transgene. The anti-pY-Stat92E epitope is strongly reduced in cells that express dome but not in the patch of GFP-negative, ept mutant cells that do not express dome.

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Figure 7. A hypothetical model of the cell-autonomous effects of ept loss. This model incorporates data from this paper and an accompanying study on the link between ept, DaPKC, Notch, Crb, Psn and Rbf1. The Erupted protein (Ept) is required to traffic Crb, Notch, and Dome into the lysosome. DaPKC may regulate an upstream step in this process by promoting internalization of Notch and Crb. Loss of Ept causes Notch, Crb, and Dome (and perhaps other unidentified receptors) to accumulate in the endosome, from which they drive downstream effects. Stat92E likely has many nuclear targets in ept mutant cells, including crb. Given the links between mammalian Stats and G1 cyclin expression, Stat92E may have the potential to affect expression of a cyclin as well. Presenilin (Psn) acts upstream of Rbf1, perhaps via its role in activating Notch; a similar role for Psn upstream of the Notch-target Upd is untested but would follow logically from the available data. The excess Crb expressed in ept mutant cells is predicted to cycle back into the late endosome (LE), leading to very high levels of vesicular Crb. Overexpressed Crb is known to disrupt tissue architecture (e.g. [30]) and may to do the same in the context of ept tumors.

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vps23 additionally affects trafficking of the Tkv type-I TGFβ receptor [6], and loss of the hsm gene affects a wide spectrum of cell surface receptors [56]. It remains to be determined whether or not the effect of ept on Dome is direct, and whether Dome homology in other species are also affected by alterations in ESCRT-mediated trafficking. However if these proteins require TSG101 to traffic through the ESCRT pathway in a specific subset of mammalian epithelia, it may be that loss of TSG101 function in these tissues will result in growth phenotypes similar to those observed in Drosophila imaginal disc epithelia lacking ept.

Materials and Methods

Genetics

Croses were done at 25°C unless otherwise indicated. ept clones were generated by crossing w;ernetFRT80B/TM6B and yweyFLP;P[m-w];ubGFP,FRT80B. ept,H99 clones were generated by crossing w;ernetFRT80B/TM6B and yweyFLP;P[m-w];ubGFP,FRT80B. ept mutant eye-antennal tumors were generated by crossing w;ernetFRT80B and yweyFLP;P[m-w];yweyFLP;P[m-w];ubGFP,FRT80B. The 3xGAS-lacZ reporter was placed into control or ept mutant backgrounds by crossing 3xGAS-lacZ,FRT80B/TM6B or 3xGAS-lacZ,ernetFRT80B,FRT80B/TM6B males to yweyFLP;P[m-w];ubGFP,FRT80B females. 10xStat-GFP (gift of E. Bach) activity was measured by crossing 10xStat-GFP;ernetFRT80B/TM6B or 10xStat-GFP;ernetFRT80B to yweyFLP;P[m-w];L141,TM6B. ept,stat92E06346/+ animals were obtained by crossing w;ernetFRT80B,stat92E06346/+;ubGFP,FRT80B/TM6B, UAS-dome; ept/TM6B with Domeless-intra (gift of S. Noselli) 1:200; goat anti-rabbit Cy3, goat anti-mouse Cy3, goat anti-human guinea pig Cy3, and goat anti-rat Cy3 (Jackson Laboratories) at each 1:50; JOYO (Molecular Probes) was used at 1:2000.

Molecular biology

Total RNA from FRT80B/M(3), ept/M(3) and ept/TM6B, stat92E06346/+ animals was isolated using TRIzol (Invitrogen) and reverse transcribed (SuperScript II RT/Invitrogen). cactctgcttgtcg-3 and 5'-cgacg-9 transcripts were analyzed by qPCR (SYBR Green 1 Master/Roche). 10xStat-GFP;ernetFRT80B/TM6B animals were obtained by crossing UAS-domeGFP;ernetFRT80B/TM6B and yweyFLP;P[m-w];RpL141,FRT80B/TM6B animals. rp49 was analyzed by qPCR (SYBR Green 1 Master/Roche) with RpL141,FRT80B/TM6B activity was measured by Caspase-3 (blue).

References


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Microscopy & Immunohistochemistry

Immunostaining and confocal microscopy was performed as described previously [38]. Antibodies used: rat anti-Crb-extra (gift of U. Tepass and E. Knust) 1:500; guinea pig anti-Hrs (gift of H. Bellen) 1:1000; rabbit anti-pYStat92E (Cell Signaling) 1:1000; mouse anti-Dig (DSHB) 1:20; rabbit anti-Domeless-intra (gift of S. Noselli) 1:200; goat anti-rabbit Cy3, goat anti-mouse Cy3, goat anti-human guinea pig Cy3, and goat anti-rat Cy3 (Jackson Laboratories) at each 1:50; JOYO (Molecular Probes) was used at 1:2000.

Supporting Information

Figure S1 Rates of death in ept/tsg101 mutant eye-antennal clones. A clone of ept/tsg101 mutant cells in the eye disc marked by the absence of GFP (green) stain brightly for the cleaved, activated form of Caspase-3 (blue). Found at: doi:10.1371/journal.pone.0007083.s001 (0.54 MB TIF)

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Author Contributions

Conceived and designed the experiments: MMG CKB BSR KHMM. Performed the experiments: MMG CKB BSR. Analyzed the data: MMG CKB BSR KHMM. Wrote the paper: MMG CKB KHMM.


