Introduction

Ionizing radiation (IR) is a commonly employed treatment modality for many types of human cancers. In clinical practice, radiation is often used in combination with other types of treatments including chemotherapy and surgery [1]. For example, concomitant chemotherapy plus radiation is the standard of care for treating patients with head and neck cancer, lung cancer, rectal cancer, cervical cancer, glioblastomas, and anal cancer [2-6]. It is estimated that nearly 60% of all cancer patients receive radiation as a component of their treatment [7]. This widespread use and the need for new, targeted therapies to avoid systemic toxicity associated with standard chemotherapy has prompted researchers to explore new ways to utilize radiation for cancer therapy [8-14]. One of the unique aspects of using IR for treating tumors is that it can be delivered to a focused tissue volume using either external beams or internal sources such as brachytherapy seeds. This allows for the deposition of high cumulative radiation doses at the tumor site while sparing the normal surrounding tissues. IR can achieve cell killing through interaction with water molecules and subsequent formation of hydroxyl radicals that lead to DNA strand breaks [15]. These DNA strand breaks are cytotoxic primarily in rapidly dividing cells such as cancerous neoplasms. IR has been shown to elicit phenotypic changes in both normal and cancerous cells that survive an insult [16, 17].

Over the past 15 years, there has been increasing interest in molecules that are expressed on the surface of tumor cells following irradiation. The usefulness of these neoantigens for imaging and therapeutic applications lies in the fact that they are differentially expressed on the surface of irradiated tumor cells to a greater extent than on normal tissues. Drug delivery vehicles or imaging agents conjugated to ligands that recognize and interact with the neoantigens can help to improve tumor-specific targeting and reduce systemic toxicity with cancer drugs. This article provides a review of the molecules that have been reported to be expressed on the surface of tumor cells in response to IR either in vivo or in vitro. Additionally, we provide a discussion of some of the methods used in the identification of these antigens and applications for their use in drug delivery and imaging.

Keywords: Radiation-induced neoantigens, targeted therapy, nanoparticles, ionizing radiation
specific tumor targeting and reduce systemic toxicity with cancer drugs. Conceptually, the IR can be described as “marking” the tumor for further destruction or imaging. This is a role that suits radiation well because of the tissue penetration and precise delivery to the perceived tumor volume that can be achieved.

Tumors are heterogeneous masses consisting of networks of microvessels and cancer cells. Both endothelial cells and cancer cells comprising tumors have been shown to respond to IR by increasing the surface expression of various molecules [18-22]. However, most of the reports to date have focused on targeting tumor endothelial neoantigens [8, 9, 12, 23]. An advantage of targeting endothelial antigens lies in improved penetration into tumors, improved drug bioavailability, and direct access from the intravascular space [24]. In addition, studies have shown that some molecules like α2β3 integrin are expressed following exposure to IR on many types of tumor vasculature [23]. This suggests that therapeutic or imaging schemes targeting such endothelial molecules would be widely applicable to most, if not all, solid tumors treated with radiation. Figure 1 illustrates a proposed model that explains how irradiation of tumor cells leads to the overexpression of neoantigens in tumor endothelial cells which can then be targeted for drug delivery. The incident IR induces cellular signaling pathways which lead to the upregulation of antigen production and expression on the cell surface. Nanoparticles encapsulating chemotherapeutic drugs conjugated with antigen-specific ligands can then be delivered to the tumor intravascularly.

While antigens that are upregulated at the surface of tumor endothelial cells offer excellent prospects for tumor therapy, other studies have explored the surface expression of molecules directly on cancer cells following irradiation [21, 25-28]. In addition to inducing surface expression of antigens, IR may further improve the delivery of agents - especially bulky ones like liposomes or nanoparticles - to tumor cells because it transiently increases the permeability of the vasculature [29]. There are currently a number of approaches being studied to improve delivery of nanotherapeutics directly to tumors and facilitate tumor penetration through normalization of the vasculature and the interstitial matrix [30-33].

A wide variety of methods that exploit radiation induced neoantigens for imaging or cancer therapeutics have been reported in the literature. Site-directed drug delivery is one of the more commonly investigated applications. Immunoliposomes encapsulating combretastatin were used to target E-selectin on irradiated mouse mammary tumors and melanomas [13, 14]. Others have used drug-encapsulated nanoparticles conjugated to antigen-specific ligands for targeting irradiated tumors [12, 23, 34]. The effect of IR on tumor cell immunogenicity has also been a topic of interest. IR has been shown to increase surface expression of molecular targets which promotes tumor cell killing by endogenous cytotoxic cells [16, 18, 35]. In mouse models it has been shown that irradiation of cutaneous melanoma prior to resection resulted in a dramatic reduction in lung metastasis after systemic challenge with untreated melanoma cells [36]. This evidence indicates that IR may play an important role in enhancing the immunogenicity of cellular tumor vaccines [26, 37]. More recently, work on the identification of a molecule TIP-1 that translocates to the surface prior to cell death has led to an interest in developing post-irradiation tumor imaging modalities and may be useful in predicting tumor responsiveness to radiation [38].

The goal of this article is to provide a review of the molecules that have been reported to be expressed on the surface of tumor cells in response to IR either in vivo or in vitro (Table 1). Additionally, we provide a discussion of some of the methods used in the identification of these antigens and applications for their use in drug delivery and imaging. New strategies for targeting cancers by way of radiation induced antigens have the potential to be very useful in the fields of drug delivery, immunotherapy, and imaging.

**Cell adhesion molecules**

Cell adhesion molecules mediate cell-cell and cell-matrix interactions and are expressed widely in human tissue. They possess many biologically important roles and are involved in tumor development and invasion, inflammatory responses, migration and differentiation, embryogenesis, cell growth, coagulation, wound repair and tissue integrity [39-42]. There are a number of different families of adhesion molecules including integrins, selectins, cadherins,
Radiation induced neoantigens

Table 1. Table of radiation-induced molecules in endothelial or tumor cell lines

<table>
<thead>
<tr>
<th>Antigen</th>
<th>Cell Line/Tissue Type</th>
<th>Model</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>P-selectin</td>
<td>Endothelium of tumors:</td>
<td>In vivo: murine, rat</td>
<td>[20]</td>
</tr>
<tr>
<td></td>
<td>C6 glioma cell line</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>MCa4 mammary carcinoma cell line</td>
<td></td>
<td>[20]</td>
</tr>
<tr>
<td></td>
<td>Lewis lung carcinoma cell line</td>
<td></td>
<td>[20, 44]</td>
</tr>
<tr>
<td></td>
<td>WiDR colon carcinoma cell line</td>
<td></td>
<td>[20]</td>
</tr>
<tr>
<td></td>
<td>MM48 and MM46 murine mammary cell line</td>
<td></td>
<td>[45, 46]</td>
</tr>
<tr>
<td></td>
<td>HUVEC cell line</td>
<td>In vitro</td>
<td>[43]</td>
</tr>
<tr>
<td></td>
<td>Oral Mucosa</td>
<td>In vivo: human</td>
<td>[51]</td>
</tr>
<tr>
<td></td>
<td>HMEC-1 cell line</td>
<td>In vitro</td>
<td>[50]</td>
</tr>
<tr>
<td>E-selectin (LECAM2)</td>
<td></td>
<td>In vitro</td>
<td>[14]</td>
</tr>
<tr>
<td></td>
<td>HUVEC cell line</td>
<td></td>
<td>[27, 53]</td>
</tr>
<tr>
<td></td>
<td>Oral Mucosa</td>
<td></td>
<td>[16]</td>
</tr>
<tr>
<td>ICAM-1 (CD54)</td>
<td>UCI 101, T-222, human ovarian carcinoma cell line</td>
<td>In vitro</td>
<td>[27, 53]</td>
</tr>
<tr>
<td></td>
<td>Caco-2, HCT116, WiDr, SW1463, SW403, SW480 T84, COLO</td>
<td>In vitro</td>
<td>[16]</td>
</tr>
<tr>
<td></td>
<td>205 colon carcinoma cell lines</td>
<td></td>
<td>[16]</td>
</tr>
<tr>
<td></td>
<td>A549, SW900, NCI-H464 lung carcinoma cell lines</td>
<td>In vivo: human</td>
<td>[53]</td>
</tr>
<tr>
<td></td>
<td>DU145, PC-3 prostate cell lines</td>
<td>In vitro</td>
<td>[54]</td>
</tr>
<tr>
<td></td>
<td>Cells from freshly isolated Human ovarian tumors</td>
<td>In vivo: human</td>
<td>[53]</td>
</tr>
<tr>
<td></td>
<td>CaSi, SiHa, HT-3 human cervical carcinoma cell line</td>
<td>In vitro</td>
<td>[54]</td>
</tr>
<tr>
<td></td>
<td>HUVEC cell line</td>
<td></td>
<td>[11, 47, 50, 52]</td>
</tr>
<tr>
<td></td>
<td>Oral Mucosa</td>
<td>In vivo: human</td>
<td>[50]</td>
</tr>
<tr>
<td></td>
<td>ET-1 Ewing tumor cell line</td>
<td>In vitro</td>
<td>[51]</td>
</tr>
</tbody>
</table>

Figure 1. Ionizing radiation leads to the upregulation of surface molecules in tumor endothelial cells that can be targeted for drug delivery. A schematic illustrating a generalized mechanism by which irradiation can be used to deliver tumor targeted therapies. 1) Ionizing radiation is incident upon the tumor from an external source. 2) Radiation induces the upregulation of the neoantigen and its subsequent translocation to the cell surface. 3) Intravascular nanoparticles conjugated with a targeting moiety bind to the irradiated tumor endothelial cell surface. 4) The nanoparticle is endocytosed and the drug contents are released to the surrounding tumor cells.
### Radiation induced neoantigens

<table>
<thead>
<tr>
<th>Name</th>
<th>Reference(s)</th>
</tr>
</thead>
<tbody>
<tr>
<td>ARP-1, ARK-RS multiple myeloma cell line</td>
<td>In vitro [56]</td>
</tr>
<tr>
<td>Cells from isolated human primary multiple myeloma tumors</td>
<td>In vitro [57]</td>
</tr>
<tr>
<td>NCAM (CD56)</td>
<td>In vitro and in vivo: murine [56]</td>
</tr>
<tr>
<td>V-CAM</td>
<td>In vitro [48]</td>
</tr>
<tr>
<td>Endothelium of tumors:</td>
<td></td>
</tr>
<tr>
<td>B16/OVA melanoma cell lines</td>
<td>In vivo: murine [34]</td>
</tr>
<tr>
<td>PECAM-1 (CD31)</td>
<td>In vitro [17]</td>
</tr>
<tr>
<td>αβ Integrin</td>
<td></td>
</tr>
<tr>
<td>Endothelium of tumors:</td>
<td></td>
</tr>
<tr>
<td>GL261 murine glioma</td>
<td>In vivo: murine [23]</td>
</tr>
<tr>
<td>Lewis lung carcinoma cell line</td>
<td>In vivo: murine [23]</td>
</tr>
<tr>
<td>B16FO and B16F10 murine melanoma cell lines</td>
<td>In vitro [9, 13]</td>
</tr>
<tr>
<td>αβ Integrin</td>
<td>In vitro [58]</td>
</tr>
<tr>
<td>U87MG, LN-18, LN-229 human malignant glioma cell lines</td>
<td>In vitro [60]</td>
</tr>
<tr>
<td>B1 Integrin</td>
<td>In vitro [19]</td>
</tr>
<tr>
<td>A549, SKMES1 lung carcinoma cell lines</td>
<td>In vitro [25]</td>
</tr>
<tr>
<td>Colo-320 colon carcinoma cell line</td>
<td>In vitro [25]</td>
</tr>
<tr>
<td>E-cadherin (CD324)</td>
<td>In vitro [61]</td>
</tr>
<tr>
<td>A549 lung carcinoma cell lines</td>
<td>In vitro [61]</td>
</tr>
<tr>
<td>T-SCC squamous cell carcinoma lines</td>
<td>In vitro and in vivo: murine [64]</td>
</tr>
<tr>
<td>MHC Class I</td>
<td>In vitro and in vivo: murine [16, 67]</td>
</tr>
<tr>
<td>carcinoma cell lines</td>
<td></td>
</tr>
<tr>
<td>A549, NCI-H23, NCI-H647 lung carcinoma cell line</td>
<td>In vitro [16]</td>
</tr>
<tr>
<td>MKN45 gastric adenocarcinoma cell line</td>
<td>In vitro [21]</td>
</tr>
<tr>
<td>UC101, UC107, SKOV-3, T-222 ovarian carcinoma cell lines</td>
<td>In vitro [27, 53]</td>
</tr>
<tr>
<td>Cells from freshly isolated human ovarian tumors</td>
<td>In vivo: human [53]</td>
</tr>
<tr>
<td>SiHa, HT-3, CaSkI cervical carcinoma cell line</td>
<td>In vitro [54, 55]</td>
</tr>
<tr>
<td>ARP-1, ARK-RS multiple myeloma cell lines</td>
<td>In vitro [57]</td>
</tr>
<tr>
<td>MeJuSo, B16-F10 melanoma cell line</td>
<td>In vitro [66, 67]</td>
</tr>
<tr>
<td>Cells from isolated human primary multiple myeloma tumors</td>
<td>In vitro [57]</td>
</tr>
<tr>
<td>GL261 glioma cell line</td>
<td>In vitro [68]</td>
</tr>
<tr>
<td>CD80 (B7.1)</td>
<td></td>
</tr>
<tr>
<td>P815 mastocytoma cell line</td>
<td>In vitro [26]</td>
</tr>
<tr>
<td>mhAT3f, mhAT2 hepatoma cell line</td>
<td>In vitro [26]</td>
</tr>
<tr>
<td>NIH3T3 fibroblast cell line</td>
<td>In vitro [26]</td>
</tr>
<tr>
<td>AK7 mesothelioma cell line</td>
<td>In vitro [26]</td>
</tr>
<tr>
<td>CD20</td>
<td></td>
</tr>
<tr>
<td>IM9, IM9/Bcl2, Ramos neoplastic B-cell lines</td>
<td>In vivo [77]</td>
</tr>
<tr>
<td>GM1310B lymphoblastic cell line</td>
<td>In vivo [77]</td>
</tr>
<tr>
<td>Raji, Daudi Burkitt lymphoma cell lines</td>
<td>In vitro [78]</td>
</tr>
<tr>
<td>NKG2D Ligands (MICA, MICB, ULBP1,2,3)</td>
<td>In vitro [35]</td>
</tr>
<tr>
<td>Fas (CD95/Apo-1)</td>
<td></td>
</tr>
<tr>
<td>MCF-7, ZR-75.1, U2-OS breast carcinoma cell line</td>
<td>In vitro [28]</td>
</tr>
<tr>
<td>U87 glioma cell line</td>
<td>In vitro [81]</td>
</tr>
<tr>
<td>Cells from isolated human gliomas</td>
<td>In vitro [81]</td>
</tr>
<tr>
<td>Biopsy tissue from human head and neck lymphoma</td>
<td>In vivo: human [82]</td>
</tr>
<tr>
<td>MC38 murine colon adenocarcinoma</td>
<td>In vitro [18]</td>
</tr>
<tr>
<td>FasL</td>
<td></td>
</tr>
<tr>
<td>C15 nasopharyngeal carcinoma cell line</td>
<td>In vivo: murine [83]</td>
</tr>
<tr>
<td>NOS2, HOS osteosarcoma cell lines</td>
<td>In vitro [86]</td>
</tr>
<tr>
<td>MCF7 breast carcinoma cell line (TRAIL-R2 only)</td>
<td>In vitro [85]</td>
</tr>
<tr>
<td>Hep-2R human larynx squamous cell carcinoma line</td>
<td>In vitro [90]</td>
</tr>
<tr>
<td>GRP78 (BIP)</td>
<td></td>
</tr>
<tr>
<td>HUVECs grown in coculture with GL261 glioma cell line</td>
<td>In vitro [12]</td>
</tr>
<tr>
<td>Endothelium of tumors:</td>
<td></td>
</tr>
<tr>
<td>GL261 murine glioma</td>
<td>In vivo: murine [12]</td>
</tr>
<tr>
<td>MDA-MB-231 breast carcinoma cell line</td>
<td>In vitro [12]</td>
</tr>
<tr>
<td>Calreticulin</td>
<td></td>
</tr>
<tr>
<td>CT26 colon carcinoma cell line</td>
<td>In vitro [103]</td>
</tr>
<tr>
<td>B16F0 melanoma cell line</td>
<td>In vitro [36]</td>
</tr>
<tr>
<td>ERp57 (grp58/PDIA3)</td>
<td></td>
</tr>
<tr>
<td>CT26 colon carcinoma cell line</td>
<td>In vitro [105]</td>
</tr>
<tr>
<td>MKN45 gastric adenocarcinoma cell line</td>
<td>In vitro [16, 21]</td>
</tr>
<tr>
<td>Caco-2, HCT 116, WiDr, HT-29, LS 174T, SW1463, SW480, T84, LoVo, COLO 205 colon carcinoma cell lines</td>
<td>In vitro [16]</td>
</tr>
<tr>
<td>A549, SK-LU-1, SW900, NCI-H23, GL-1 lung carcinoma cell lines</td>
<td>In vitro [16, 109]</td>
</tr>
<tr>
<td>22Rv1, LNCAP prostate carcinoma cell line</td>
<td>In vitro [16]</td>
</tr>
<tr>
<td>TIP-1 (TAX1BP3)</td>
<td></td>
</tr>
<tr>
<td>H460, LLC lung carcinoma cell line</td>
<td>In vitro and in vivo: murine [38, 113]</td>
</tr>
</tbody>
</table>
Radiation induced neoantigens

the immunoglobulin superfamily, and cell surface proteoglycans. The effects of IR on the expression of these molecules varies depending on the individual adhesion molecule and the irradiated cell line/tissue [39].

**P-Selectin**

P-selectin is a molecule that is involved in the adhesion and activation of circulating inflammatory cells and platelet aggregation [22]. Hallahan et al. studied P-selectin expression in irradiated tumor blood vessels and found that prior to irradiation P-selectin is located within the endothelium. After irradiation however, P-selectin was translocated to the lumen of the blood vessels. Its expression on the endothelial cell surface was transient, and platelet aggregation studies showed resolution by 48 hours after irradiation [43]. The surface expression of P-selectin following irradiation was observed in all neoplasms studied and in both rat and mouse species [20, 23]. The effect was only observed in irradiated tissues, suggesting preferential expression after IR treatment. Other studies have shown increased expression of P-selectin on human umbilical vascular endothelial cells (HUVECs) in vitro [43]. These observations have led to the use of single chain variable fragment (ScFv) sequences for targeted visualization of radiation induced P-selectin in lung tumors [44]. P-selectin has also been used as a targeting molecule for the delivery of drug-filled nanocapsules to tumors following irradiation leading to prolonged survival and reduction of metastasis in mouse models [45, 46].

**E-Selectin**

E-selectin is another molecule which is upregulated at both the mRNA and the protein level in endothelial cells in response to IR [47-50]. E-selectin expression in HUVECs begins to increase at 2 hours following irradiation and peaks at 4-6 hours [47]. The elevated levels of surface expression of E-selectin persist on endothelial cells for several days in treated murine lung tissue [49]. In human dermal microvascular endothelial cells (HDMECs), E-selectin was not shown to be present in untreated controls, but expression levels were increased after treatment with both 5 and 10 Gy [48]. In the oral mucosa of head and neck cancer patients treated with total doses of 30-60 Gy, immunohistochemical staining showed a significant increase in expression of E-selectin and may be involved in the development of oral mucositis following irradiation [51]. Recently, Patillo et al. targeted irradiated mouse mammary tumors with anti-E-selectin conjugated immunoliposomes containing the antivascular drug combretastatin disodium phosphate [14]. They found that unirradiated MCa-4 tumor endothelial cells expressed a basal level of E-selectin, while irradiated tumors showed significantly increased E-selectin expression [14]. Tumor treatment in xenografted mice with the targeted liposomes plus fractionated radiation led to a significant delay in tumor growth compared to fractionated radiation or targeted liposomes alone [14].

**ICAM-1**

Another molecule of interest is ICAM-1 which is a member of the Ig superfamily. It is constitutively expressed on the surface of various cells, including macrophages, leukocytes and endothelial cells and is involved in leukocyte trafficking, vascular integrity, and inflammation [22, 52]. Increased surface expression of ICAM-1 was found on tumor cells from freshly isolated human ovarian tumors and ovarian cancer cell lines following high dose irradiation [27, 53]. Cervical cancer cell lines were also found to exhibit upregulated surface ICAM-1 after treatment with IR [54]. Santin et al. later showed that combining retinoic acid treatment with high dose gamma irradiation induces an additive effect and leads to increased surface expression of ICAM-1 over either treatment alone [55]. The results indicate that the combination of these two treatments may provide a powerful antiproliferative effect. A study of human colon, lung and prostate cancer cell lines showed upregulation of ICAM-1 at the surface and indicated that it may be involved in enhanced T-cell binding to tumor targets and enhanced co-stimulatory signaling with T-cells [16]. Both ICAM-1 and NCAM expression were increased in an Ewing tumor cell line following irradiation [56]. Additionally, ICAM-1 upregulation was seen in multiple myeloma cell lines as well as myeloma primary tumors after irradiation [57]. The upregulation persisted in multiple myeloma cells for 6 days. This indicates that ICAM-1 may be a useful target for tumor therapeutics or imaging applications and may be involved in increasing the immunogenicity of the cells [57].

Endothelial cell expression of ICAM-1 has been
another topic of interest. It has been shown that ICAM-1 is increased on the surface of HDMECs, HUVECs, and human microvascular endothelial cells (HMEC) after treatment with IR which suggests that it participates in the radiation-induced inflammatory reaction of endothelium [17, 47, 48, 50, 52]. There have been no reports of attempts for radiation-guided targeting of ICAM-1 for therapeutic purposes to date. However, Kiani et al. studied the upregulation of ICAM-1 by quantifying the delivery of micro-spheres coated with anti-ICAM-1 antibodies to tumor cells both in vivo and in vitro [11]. They found enhanced microsphere adhesion to HUVECs under in vitro flow conditions. In an in vivo rat cranial window model in which they irradiated cerebral tissue, they observed a 13-fold increase in adhesion of the anti-ICAM micro-spheres compared to control indicating that ICAM-1 may be a suitable target for drug delivery [11].

V-CAM

V-CAM is a molecule that has been shown in multiple reports to exhibit no increase in surface expression following IR in HUVECs, HDMECs, and oral mucosa [17, 47, 51]. In contrast, a study by Heckman et al. found that V-CAM up-regulation was increased on an mRNA and protein expression level in HDMECs [48]. The elevated protein expression persisted up to 24 hours after irradiation. Lugade et al. further showed V-CAM upregulation in tumor vasculature of B16/OVA melanomas xenografted into mice thighs after treatment with either 3 or 15 Gy. They related this upregulation following irradiation to the creation of a tumor microenvironment that selects for increased infiltration and retention of T cells [34]. To our knowledge, V-CAM has never been targeted in vivo for drug delivery or imaging applications.

Integrins

Integrins are heterodimeric transmembrane molecules that are comprised of an α and β subunit and are subdivided into subfamilies based upon the β subunit [39]. Each subunit contributes to ligand specificity and contains ligand binding sites. Integrins serve as receptors for many proteins including Ig superfamily adhesion molecules and extracellular matrix proteins like the collagen, fibronectin, vitronectin, and fibrinogen receptors [9]. The conserved peptide sequence arginine-glycine-aspartic acid (RGD) is a major recognition site for several integrins and is commonly found in ligands that bind to integrins including α2β3. Onoda et al. found a transient increase in the expression of α2β3 in B16 melanoma cells after low-dose gamma irradiation. This increased expression resulted in enhanced adhesion to fibronectin in vitro and increased induction of metastases in vivo suggesting the association between upregulation of integrins in cells surviving radiation and increased malignant potential [58]. To assess the targeting of α2β3 in irradiated tumors, B16FO tumor bearing mice were treated with integrin-specific peptides. Increased binding was demonstrated within tumor blood vessels after irradiation as compared to controls [9]. In a seminal paper from 2003, Hallahan et al. reported the first demonstration of radiation-guided drug delivery to tumor blood vessels. They showed a radiation-induced accumulation of α2β3 within the lumen of the microvasculature of tumors isolated 6 hours after irradiation with no increase in untreated controls [23]. Irradiated tumors were then targeted with radiopharmaceuticals by use of α2β3 ligands conjugated to nanoparticles and liposomes. A significant growth delay was observed in irradiated tumors [23]. More recently, RGD conjugated liposomes incorporating the anti-angiogenic drug combretastatin were used to target α2β3 in irradiated mouse melanomas. A single dose of 5 Gy combined with immunoliposome treatment lead to a 5.1-day tumor growth delay compared to untreated controls [13].

Other studies have shown a radiation-induced upregulation of various α and β integrins in lung cancer cell lines and colon cancer cell lines in vitro which may have implications for the induction of malignant phenotypes following IR [19, 25]. β3 integrin and its associated signaling pathways have been explored as a potential therapeutic target for breast cancer [59]. αβ3 integrin mRNA expression was shown to be upregulated in response to IR in 3 glioma cell lines which lead to increased migration and invasiveness [60]. These data suggest that improved therapeutic efficacy of radiotherapy in malignant gliomas may be aided by inhibition of migration and invasion.

Cadherins

The cadherins are molecules involved in cell-cell
Radiation induced neoantigens

adhesion and maintenance of epithelial integrity. They are thought to play an important role in the invasiveness and the metastatic potential of cancer cells [61]. E-cadherin is a calcium-dependent cell adhesion molecule. Its reduced expression by tumor cells may be related to the invasive potential of some cancers [62, 63]. Akimoto et al. demonstrated increased expression of E-cadherin and α-catenin after fractionated irradiation of the A549 human lung cancer cell line which led to a reduction in invasive capacity in the irradiated cells [61]. A study by Ebara et al. on the effects of IR on the surface expression of E-cadherin found it to be increased in thyroid gland squamous cell carcinoma cells both in vivo and in vitro [64]. The authors suggest that this upregulation may decrease the metastatic propensity through inhibition of detachment of the tumor cells from the primary site [64].

Immunoregulatory molecules

The discovery that immunomodulatory molecules are upregulated post-IR on the surface of certain tumor cells has led to research involving tumor cell vaccines and potentiation of cytotoxic T lymphocyte (CTL) mediated tumor killing [37]. Studies have demonstrated that an enhancement in the immune system's response to tumor antigens can be achieved using various cancer vaccines targeting tumor associated antigens [65].

MHC Class I

MHC Class I molecules are important for presentation of intracellular antigens to class I-restricted T cells. These molecules have been shown to be upregulated on tumor cells following irradiation in a variety of cell lines including multiple myeloma, melanoma, colon, lung, ovarian, and gastric tumor cell lines [16, 21, 27, 57, 66, 67]. In tumor cells, MHC Class I molecules play a role in antigen presentation of tumor molecule epitopes to circulating T cells, and their upregulation has been shown to enhance the effectiveness of immunotherapy in high grade gliomas treated with IR [68]. Conversely, through the downregulation of MHC molecules, tumor cells may evade immune recognition and elimination [69]. Reits et al. showed that immunotherapy was successful in eradicating murine colon adenocarcinoma through CTL mediated killing only when preceded by radiotherapy of the tumor tissue [67].

Three schemes through which IR leads to the upregulation of MHC class I molecules were described: early effect, repair response, and late effect [67]. The early effect increase of MHC Class I occurs in the first 0-4 hours after irradiation and involves damage and degradation of the existing cellular proteins with subsequent presentation of the degradation products via MHC class I. The repair response effect occurs when irradiation forms free radicals through water radiolysis which leads to oxidation of protein amino acids. This modification targets affected proteins for degradation and subsequent upregulation of MHC Class I. Finally, the late effect involves mTOR pathway activation, increased protein synthesis, and an increase of the intercellular peptide pool for subsequent presentation [67]. These findings may help to explain the abscopal effect which is a phenomenon in which IR can inhibit distant tumors outside of the region where local IR is absorbed [70].

CD80

CD80 is a co-stimulatory molecule that is essential for activating T lymphocytes. Recognition of MHC class I peptide presentation without CD80 co-stimulation leads to T cell anergy. Typically, CD80 is found on activated antigen presenting cells like B-cells and macrophages. However, Morel et al. showed that IR can induce CD80 neoexpression in various murine tumor cells including hepatomas and mastocytomas, which prior to irradiation constitutively exhibited no CD80 expression [26]. These findings may further explain the irradiation-enhanced immunogenicity of many tumor cells, which may present the tumor antigen and then directly provide the costimulatory signal to the lymphocyte, provided that MHC Class I molecules and CD80 molecules are expressed on the same cell. Galiximab is a monoclonal antibody to CD80 which has been demonstrated to have antitumor activity against various B-cell lymphoma models in preclinical trials [71]. However, the role of radiation in enhancing the effects of anti-CD80 therapy has yet to be fully explored.

CD20

A majority of B-cell lymphomas express the CD20 surface antigen which has been a mo-
The cell undergoes early or delayed cell death [79]. One molecule that is implicated in cellular apoptosis following irradiation is Fas (also called APO-1 or CD95). Fas is a cell-surface death receptor expressed in a variety of tissues and interaction with its ligand, FasL, plays a role in the control of the immune system as well as in triggering apoptosis [80]. Chakraborty et al. showed directly that upregulation of Fas on murine colon adenocarcinoma cells after irradiation lead to enhanced cytotoxic T lymphocyte activity. These results hold implications for further work exploring a combination of immunotherapy and radiation therapy [18]. Sheard et al. showed that after irradiation, Fas was upregulated at the surface in p53-wild type breast carcinoma cells in vitro in a dose-dependent manner [28]. This effect was observed at doses at and below 10 Gy. However, p53 null and mutant cell lines failed to up-regulate Fas levels and halt their cell cycles in the G1 phase indicating that p53 status is an important determinant of radiation induction of Fas. Another study by Ishikawa et al. showed that Fas expression levels increased in a dose dependent manner following irradiation of human malignant glioma cells and the U87 glioma cell line [81]. This observation was accompanied by enhanced cell-mediated cytotoxicity of the glioma cells in vitro which suggests that local irradiation combined with immunotherapy may be effective in the management of malignant gliomas. In another study, Ogawa et al. demonstrated that tumor tissue from patients with malignant lymphomas showed differentially increased Fas expression before and after IR treatment [82]. The Fas-positive lymphomas showed high radiosensitivity. However, radioresistant squamous cell tumors showed negative or only slight elevations in Fas levels which suggests that Fas expression may be useful as a marker of radiation efficacy in some tumor types [82].

FasL

FasL is another molecule that has been studied in tumor cells following treatment with IR. It is known to be the ligand to the Fas receptor, and is an essential determinant of apoptosis. Abdulkarim et al. demonstrated strong surface co-expression of FasL and Fas on nasopharyngeal carcinoma cells treated with radiation in vivo in which a large percentage of cells underwent apoptosis [83]. These results led them to conclude that these molecules can play a major role in the anti-tumor effect of IR at therapeutic
The death receptor pathways that are involved during radiation-induced early and delayed cell death were studied by Luce et al. in a number of cell lines [79]. Following radiation, they observed early upregulation of Fas, TRAIL-R2 (DR5), and TNF-R2 which was associated with sensitization of the cells to apoptosis. However, late expression of FasL, TRAIL, and TNF-α was associated with mitotic catastrophe and delayed cell death.

**TRAIL**

The TRAIL receptors TRAIL-R1 (DR4) and TRAIL-R2 possess an intercellular death domain which is capable of activating the extrinsic pathway for apoptosis after TRAIL-induced receptor trimerization [84]. TRAIL is non-toxic when delivered intravascularly in vivo and preferentially kills neoplastically transformed cells over normal cells [85]. A study by Chinnaian et al. showed that TRAIL-R2 could be upregulated in the MCF7 breast carcinoma cell line with radiation and that the addition of TRAIL to treatment regimens led to large increases in apoptotic cell death [85]. Tumors xenografted into the hindlimbs of nude mice showed growth regression when treated with combined IR and TRAIL. More recently, these receptors were studied in human osteosarcoma cells after treatment with IR. It was found that both TRAIL receptors (DR4 and DR5) were upregulated at the cell surface [86]. Combined treatment with IR and TRAIL synergistically enhanced apoptosis and decreased cell viability in osteosarcoma cell lines. These two studies provide support for combining IR with TRAIL to improve tumor responsiveness and eradication.

Another therapeutic scheme that has gained interest recently involves the use of monoclonal antibodies targeted to TRAIL-R2 [87, 88]. A number of studies have shown the beneficial effects of combining radiation with anti-TRAIL-R2 treatment both in vitro and in vivo [89, 90]. Nagane et al. demonstrated significantly suppressed growth of subcutaneous glioma xenografts leading to complete regression in tumor-burden mice following treatment with the direct agonist anti-TRAIL-R2 mAb KMTR2 [87].

**Phosphatidyserine**

One of the most widely exploited molecules of apoptosis is the cell membrane phospholipid phosphatidylserine (PS). While it is normally confined to the cytosolic side of the membrane, induction of apoptosis leads to rapid externalization of PS. This occurs as a result of disruption of the enzymes responsible for maintaining cell membrane phospholipid asymmetry [91]. This ‘eat me’ signal helps macrophages recognize and eliminate dying cells. PS exposure is now understood to be a ubiquitous phenomenon of apoptosis that is independent of cell type and cell death inducing trigger [92]. It is no surprise then, that radiation-induced apoptosis of tumor cells leads to the expression of PS. Molecules that bind to extracellular PS with high affinity, such as Annexin V, are commonly used to target PS as an indicator of cells undergoing apoptosis [91]. PS has previously been evaluated clinically in follicular lymphoma, head and neck squamous cell carcinoma, breast cancer, and lung cancer as a marker of apoptosis and has been correlated to ultimate prognosis and treatment response [93-97]. A more in depth discussion of PS and its applications is provided below in the tumor imaging section.

**Miscellaneous molecules**

**GRP78**

There are a variety of molecules that have been shown to be upregulated in response to IR that do not fit neatly into the previously discussed categories. One of them is glucose regulated protein-78 or GRP78 (also known as BiP) which is an endoplasmic reticulum (ER) chaperone that is involved in suppression of stress-induced apoptosis [98]. On the cell surface, it can function in the transduction of extracellular stimuli to intracellular signals in the promotion of tumorigenesis [99, 100]. From a therapeutic viewpoint, GRP78 is a practical target because it is expressed at elevated levels in tumors as compared to normal tissues. Recently, Passarella et al. showed that GRP78 expression is increased in tumor endothelium after treatment with IR [12]. In co-culture experiments in which tumor cells are grown in two-tiered dishes containing HUVECs, they found that the expression of GRP78 only occurred when HUVECs and tumor cells were irradiated in co-culture together. This implies that some signaling is taking place between tumor and endothelial cells that leads to the upregulation of GRP78 in response to IR. Attempts to target GRP78 for cancer therapy have generally shown encouraging results [12,
Radiation induced neoantigens

101, 102]. Passarella et al. used peptide-conjugated nanoparticles to deliver paclitaxel to breast carcinoma tumors grown in nude mice and observed a significant delay in tumor tripling time for tumors treated with combination therapy of IR and the targeted nanoparticle [12]. Similarly, Katanasaka et al. implemented peptide-modified liposomes targeting GRP78 for the delivery of doxorubicin to colon carcinoma tumor-bearing mice and demonstrated suppressed tumor growth and prolonged survival [102].

Calreticulin

Calreticulin is another molecule that is primarily found in the ER, but is translocated to the cell surface following ER stress. It serves as a signal that allows dying cells to be recognized and ingested by dendritic cells and its expression therefore leads to increased cancer cell immunogenic apoptosis [103, 104]. Perez et al. showed that translocation of calreticulin to the cell surface is increased in response to irradiation of B16F0 melanoma cells in vitro [36]. Irradiation of subcutaneous melanomas in mice prior to resection resulted in a large reduction in systemic metastases when challenged with unirradiated melanoma cells. The results confirm that calreticulin is involved in increasing the immunogenicity of melanoma cells which are targeted for removal by dendritic cells. Calreticulin translocation is reported to be controlled by another ER stress protein ERP57/GRP58 and they are translocated to the surface together in response to IR and a number of other cytotoxic insults [105]. For therapeutic applications, induction of calreticulin by pro-apoptotic peptides has been shown to enhance the immunogenicity of tumor cells and ultimately the antitumor efficacy of these peptides [106].

CEA

Carcinoembryonic antigen (CEA) is a marker that is often used to assay for liver metastasis from colon cancers and is also used to track the response of primary colon cancer to treatment. It is one of the most widely used tumor markers worldwide and is a glycoprotein that is actually classified in the Ig superfamily [107]. While CEA is reported to be prevalent in a variety of human malignancies, a number of studies have shown the induction of CEA in tumor cells following irradiation [108]. Hareyama et al. were the first to show that the expression of CEA was increased on the surface membrane of irradiated gastric adenocarcinoma cells and that this expression was regulated at the gene expression level [21]. Matsumoto et al. showed upregulated CEA expression on the surface of GLL-1 human lung cancer cells [109]. The peak expression was observed 4 days after irradiation and combination treatment with interferonγ showed additional effect. Garnett et al. demonstrated surface increases of CEA in colon, lung and prostate tumor cell lines and that this increased expression led to improved killing by CEA-specific CTLs [16]. CEA is a target that has been shown to improve responsiveness to immune-mediated killing, and may be a useful marker for other targeted therapeutic modalities including radioimmunotherapy and drug encapsulated nanoparticles [110, 111].

TIP-1

PDZ domain proteins play roles in acting as molecular scaffolds and in maintenance of cell polarity and cell signaling [112]. Recently, TIP-1 which is an atypical PDZ domain protein in that it functions as a negative regulator of PDZ-based scaffolding was found to be translocated to the cell surface following IR in a dose dependent manner [38, 112]. Targeting TIP-1 was found to be a useful assessment of tumor response to IR due to the discovery that TIP-1 is expressed by dying tumor cells prior to apoptosis. TIP-1 was also recently shown to be an effective target for drug delivery. Peptide-conjugated paclitaxel encapsulated, doxorubicin encapsulated, and platinum nanoparticles were found to aggregate at the site of irradiated tumors leading to improved bioavailability of the drug and significant tumor growth delay when compared to controls [113-115].

Strategies for identifying and targeting inducible antigens

Surface protein analysis is commonly performed by flow cytometry using primary and fluorescently-labeled secondary antibodies [9, 21, 68, 116]. The use of propidium iodide is typically employed as a second stain to identify dead cells that have a ruptured membrane.
because propidium iodide is a membrane impermeant DNA stain that will provide a fluorescent signal in non-viable cells. This is particularly important when the protein under investigation is expressed in large quantities in internal cellular compartments where a ruptured membrane would result in false positive staining. Other useful techniques for identifying and quantifying surface proteins include membrane fraction processing followed by western blotting, and plasma membrane biotinylation in which neutravidin coated beads or gel are used to trap surface proteins conjugated with biotin. The proteins are then removed from the beads or column and are electrophoresed. Staining methods are also widely used for surface antigen detection including immunohistochemical and immunocytochemical techniques. To characterize the nature of overexpression of the antigen, mRNA levels are often detected by northern blotting. Increased mRNA expression indicates the upregulation is, at least in part, through transcriptional mechanisms. Unchanged mRNA levels from pre-to-post-treatment, given an increase in protein surface expression, may indicate either translocation of the protein from one cellular compartment to the surface, or a change in translational regulation.

Typically, a radiation-induced antigen is identified through one of the previously mentioned methods before it is targeted for drug delivery, often through the use of monoclonal antibodies. However, the use of specific antibodies for antigen discovery requires at least some a priori knowledge of what to look for before it can be found. A different option involves using phage display to determine ligands that bind to the tumor cells [117]. This can be performed without prior knowledge of any particular molecular target. The usefulness of this technique lies in the ability to screen a massive number of phages (up to $10^{10}$ variants) each which encode a different peptide sequence. The enormous variety is achieved by the introduction of a DNA library into the phages. The recombinant DNA encodes the peptide sequences which are subsequently produced and expressed on the protein capsid. The phages which express a variety of peptide sequences can be delivered intravenously, often in a mouse, for in vivo targeting studies [118]. Phages that bind to normal tissues are discarded, while phages that bind only to the tumor are harvested and the DNA can be amplified within bacteria infected with the phage. Serial rounds of “biopanning” can be performed which serve to enrich the phages homing specifically to the tumor. Phage DNA can then be sequenced to determine the amino acid sequence of peptides on the capsid which can then be reproduced for use as targeting moieties.

There have been a number of studies employing tumor targeting peptides for imaging, drug delivery, or as potential drugs themselves [117, 119]. Phage display has also proven to be useful for identifying peptides that target radiation-induced antigens [120]. Hallahan et al. utilized a phage-displayed peptide RGDGSSV to target $\alpha_{2\beta_3}$ integrin in irradiated tumors [23]. Later, phage display was used to discover a set of peptides that were capable of targeting irradiated gliomas [12]. One of the most promising peptides, GIRLRG, was found to bind to GRP-78. Subsequent studies showed that GIRLRG could be conjugated to paclitaxel encapsulated nanoparticles. Breast tumor (MDA-MB-231) and glioma (GL261) cell line xenografts in nude mice were irradiated and then treated with the peptide-nanoparticle conjugate and showed a significant increase in tumor tripling time as illustrated in Figure 2 [12]. In 2008, Han et al. published an article in which they discovered a peptide (HVGGSSV) that displayed potential as an in vivo molecular imaging probe [118]. In a subsequent paper, they identified the target for HVGGSSV as TIP-1 through a separate phage display experiment where a T7 phage-displayed human lung tumor cDNA library was screened against beads coated with the HVGGSSV peptide [38]. After 5 rounds of screening, they isolated the phages that bound to the beads, and subsequently sequenced the phage clones which encoded the amino acid sequence of TIP-1 [38]. The peptide was then shown to be capable of discerning between responding and non-responding tumors because it binds to TIP-1 which is expressed prior to cell death [38]. One of the most recent examples in the literature of peptides being employed for targeting irradiated tumors is by Hariri et al. where they conjugated the HVGGSSV peptide to a nanoparticle containing paclitaxel and studied the tumor targeting abilities [113]. They found improvement in tumor growth delay in vivo with a combination of IR and targeted nanoparticle. These results indicate that short peptide sequences can be used in a similar fashion to monoclonal antibodies for targeting radiation-induced antigens and that
they can be developed using positive and negative screening techniques to improve tumor site specificity.

**Radiation guided-cancer therapy**

Once a radiation-induced antigen is identified, the next step is to exploit its presence on tumor cells by utilizing it for targeted delivery of a therapeutic entity. This can be done in a variety of ways as evidenced by the articles cited in the previous sections. The most common ways of targeting tumor antigens is through the use of antibodies, affibodies, or peptides linked to drug encapsulated nanoparticles [12, 23, 111, 113, 121, 122]. Nanoparticles are advantageous for drug delivery because they offer tumor-specific accumulation when linked to a targeting moiety, reduced non-specific toxicity, and improved tumor killing efficacy [121]. Of the molecules listed in Table 1, many are excellent candidates for targeted therapeutics and some have already been shown to be useful for such purposes. Recently, new developments in nanoparticle drug delivery have been reported. Lowery et al. demonstrated that liposome-encapsulated doxorubicin nanoparticles can be delivered selectively to irradiated hind-limb tumors in mice when conjugated with the HVGGSSV peptide [115]. Tumor treatment with the targeted liposomes containing doxorubicin in combination with radiation showed moderate tumor growth suppression when compared to other treatment schemes including radiation alone, and free doxorubicin plus radiation [115].
Magnetic iron oxide nanoparticles have also been shown to be useful for combined cancer imaging and drug delivery [123]. These multifunctional nanoparticles have been dubbed "nanotheranostics" and can be conjugated with antibodies, aptamers, peptides, or other molecules like folate to improve tumor targeting capacity [124, 125]. They are useful as MRI contrast enhancement agents, and have been employed for drug delivery either as a carrier for externally conjugated drugs, or through pH-dependent drug release of encapsulated anticancer agents like doxorubicin [125-127]. In a preclinical in vivo study, iron oxide nanoparticles conjugated with antibodies targeting mutant EGFR receptors were shown to impart a significant survival benefit in glioblastoma xenografted mice when delivered via MRI-guided convection enhanced delivery [128]. Other metal oxides like zinc oxide nanoparticles are also currently being studied due to their cancer cell cytotoxic properties through reactive oxide species production and their relative biocompatibility [129]. There is great potential for targeting radiation-inducible antigens with new nanoparticle technologies or with the more well-studied drug delivery vehicles such as albumin-bound nanoparticles that have shown success in pre-clinical and clinical trials [130].

Another potential application that has not been the focus of much research to date is radiation-guided radioimmunotherapy. The idea is that IR from an external source can be focused on a defined tumor volume to elicit the expression of neoantigens which can then be targeted with radiolabeled antibodies. An advantage to the use of radioimmunoconjugates is that cytotoxic ionizing photons are minimally attenuated by tumor tissue which circumvents the problems of drug distribution to regions of tumors with poor blood supply or drugs that cannot penetrate tumors readily [131]. Internal emitters are therefore capable of irradiating the entire neoplasm even when the biodistribution is nonhomogeneous. Radioimmunoconjugates have been implemented thus far primarily in the treatment of hematologic malignancies [74, 76, 132]. This approach, however, has been limited by suboptimal target-to-nontarget ratio and an inability to deliver sufficient radiation doses to tumors selectively [133]. Pre-targeted radioimmunotherapy is a technique that separates the targeting moiety from the subsequently administered therapeutic radioisotope. Conceptually, sequential administration allows for maximal antibody targeting to occur prior to delivery of the therapeutic radionuclide which is bound to a small molecule to enhance its tumor uptake and rapid excretion [133]. If the radionuclide molecule has higher penetration, clearance, and diffusion rates than the targeting antibody, more rapid tumor radionuclide localization and excretion can occur which minimizes nonspecific radiation to normal tissues. Two main schemes have been proposed for pre-targeted radioimmunotherapy. In one, a tumor-specific antibody tagged with streptavidin is administered followed by a biotin-linked isotope which then binds to the antibody through a biotin-streptavidin interaction [133, 134]. In the other scheme, a molecularly engineered antibody fragment with dual antigen specificity is administered which can bind to both the subsequently delivered isotope and the targeted tumor antigen [133, 134]. A number of clinical studies targeting hematologic malignancies as well as solid tumors like gliomas, small cell lung cancer, medullary thyroid cancer, and colon cancer have been reported with varying levels of success [135-139]. A controlled open non-randomized study of the effects of adjuvant pre-targeted radioimmunotherapy in high grade glioma patients found a median disease-free survival of 28 months versus 8 months in the control group [140]. In another study, the investigators used a bispecific antibody with $^{131}$I in the treatment of medullary thyroid cancer. However, the results of the phase I/II clinical trial indicated that of 17 treated patients, minor tumor response was seen in only 5 and pain relief in 4 patients [136].

**Tumor imaging with radiation induced antigens**

In addition to targeted therapeutic applications of radiation-induced antigens, there is also the potential to create new imaging modalities based on the selective expression of these antigens on the tumor cell surface. Current noninvasive imaging methods for staging and cancer delineation include primarily 18-fluorodeoxyglucose (18-FDG) positron emission tomography (PET), magnetic resonance imaging (MRI) and computed tomography (CT). FDG-PET requires a significant conglomeration of hypermetabolic tissue in order to yield clinically detectable disease. MRI and CT require significant tissue and density contrasts, respectively, at a
Radiation induced neoantigens

given disease site in order to be observable. As a result, many smaller lesions as well as cellular groups of less active malignant histologies are routinely missed on FDG-PET, MRI and CT. The ultimate result of this imaging insensitivity is inferior disease staging which may lead to treatment with a less aggressive or inappropriate modality. It is postulated that neoantigen-based non-invasive imaging modalities would have the potential to increase sensitivity and specificity for the detection of smaller tumor sites. The neoantigen based radiotracer would localize and respond directly to the neoplastic-specific expression of cellular markers instead of the more variable and less reliable metabolic activity level or tissue and density contrasts [141, 142]. Furthermore, if the quantified level of neoantigen expression in response to radiation administration could be correlated with therapeutic resistance or ultimate prognosis, then there would exist the potential to quickly adapt treatment strategy much earlier in the treatment course than currently possible with existing imaging technology [143].

Historically, the development of neoantigen based imaging or molecular imaging has centered around the primary imaging modalities of PET, single photon emission computed tomography (SPECT), contrast enhanced MRI and contrast enhanced ultrasound. PET and SPECT have the inherent advantage of unlimited depth penetration and conjugation with a wide range of molecular agents [141]. PET provides higher spatial resolution than SPECT, though the resolution of both modalities is inferior to CT and MRI. Additionally, PET requires significant capital equipment investment with a regionally located cyclotron for the generation of the radiotracers. MRI has the distinct advantage of higher resolution and tissue contrast, and does not subject the patient to IR. In order to further improve MR imaging and tumor delineation, a molecular agent is generally coupled with an MRI contrast such as iron nanoparticles or gadolinium. This same strategy may also be employed with ultrasound. Molecular agents may be conjugated with various forms of ultrasound contrast, such as microbubbles or liposomes, in order to provide tumor specific enhancement of the ultrasound images. A limitation of this approach is reduced tissue penetration of the ultrasound contrast-agent assembly due to its relatively large size [141].

The specific technique of employing radiation-induced neoantigens into new imaging modalities is still in its infancy. While many of the previously mentioned antigens (Table 1) have been incorporated to some extent into molecular imaging research, very little of this research has involved the use of radiation as a trigger for antigen expression prior to targeted imaging. As mentioned previously, specific advantages of such a radiation-triggered molecular imaging approach would include the potential to quickly identify radio-resistant tumor variants and efficiently adapt treatment strategies [143]. Given a small number of studies involving radiation-triggered molecular imaging, this section will also explore basic molecular imaging research and techniques involving the previously mentioned radiation-induced neoantigens.

Perhaps the most successful example of a radiation-induced molecular imaging strategy to date involves the targeted delivery of iron platinum (FePt) nanoparticles to tumors implanted in mouse hindlimbs [114]. The paramagnetic FePt nanoparticles possess similar characteristics to the iron oxide nanoparticles previously discussed, and can therefore function as MRI contrast agents. In the study by Hariri et al. the nanoparticles were conjugated with the HVGGSSV peptide giving them the capacity to bind to TIP-1 which is induced by radiation at the tumor cell surface prior to initiation of apoptosis and cell death [38, 118]. The results showed that irradiated tumors possessed much higher levels of the targeted nanoparticle as compared to nanoparticles conjugated with a scrambled peptide [114]. Further, significantly higher levels of the targeted nanoparticle were observed in tumors treated with 3 Gy versus untreated tumors [114]. This study indicates that paramagnetic nanoparticles have potential application for radiation-guided targeting and imaging of cancers.

Other strategies have focused on conjugating radiotracers to RGD to assess for tumor angiogenesis [144, 145]. The RGD peptide binds to the αvβ3 integrin, which is known to be overexpressed on the activated endothelial cells of tumor neovascularure and during tumor spread [146, 147]. Numerous studies have evaluated both PET (18F and 64Cu) and SPECT (99mTc) imaging for the evaluation of tumor integrin expression, angiogenesis and metastasis [148, 149]. Out of these various modalities, 18F PET appears to be the most promising given the high tumor-to-background ratios [146, 150-152].
phase I trial of RGD conjugated to a PET radioligand in breast cancer patients was recently shown to be safe, metabolically stable, and retained in tumor tissues. In addition, it was able to detect metastatic breast cancer lesions by PET in most anatomic sites [153].

Another antigen that has been used as a molecular imaging target of tumor vascular changes is P-selectin. Hariri et al. explored the in vivo expression of P-selectin in Lewis Lung Carcinoma (LLC) tumors implanted in the hind limbs of mice in response to IR with Cy7 and 111In-DTPA labeled ScFv antibodies to P-selectin [44]. They found significantly increased tumor-specific binding of the labeled ScFv antibodies to irradiated tumors as compared to unirradiated tumors.

Closely related to P-selectin is the cellular adhesion molecule, E-selectin, which also plays a role in the inflammatory response. Funovics et al. conjugated a fluorescent peptide-magnetic nanoparticle to an E-selectin-binding peptide to evaluate the ability of this peptide imaging construct to localize preferentially to LLC as part of an in vivo mouse model [154]. They successfully demonstrated the utility of this approach and also found that E-selectin was expressed in both tumor and endothelial cells. While there has been limited follow-up work on E-selectin imaging in malignancies, there have been several research efforts to develop a clinical molecular imaging technique for benign conditions based on E-selectin. These imaging modalities are primarily based on an 111In labeled monoclonal antibody to E-selectin for use in the scintigraphic evaluation of arthritis and other inflammatory conditions [155, 156].

Two other cellular adhesion targets with molecular imaging developments and significance are ICAM-1 and V-CAM. As mentioned above, these antigens have demonstrated increased expression in tumors in response to radiation. However, in spite of the possible applications of these antigens in tumor imaging, there has been limited cancer related molecular imaging research. Instead, the antibodies to these targets have been integrated into 18F PET, photoacoustic imaging with gold nanorods, and contrast enhanced ultrasound for the in vivo evaluation of coronary artery disease, cardiac transplant rejection, atherosclerosis, and other benign inflammatory diseases [157-162].

Given the existing molecular imaging techniques for benign diseases, these antigen targets are ripe for further exploration within the malignant disease imaging realm.

In addition to cellular adhesion targets, markers of apoptosis hold special relevance to malignant tumor molecular imaging. The most thoroughly explored marker of apoptosis is PS, which translocates to the cell surface early in apoptosis in response to therapy. The protein annexin V, which binds selectively to PS, has been exploited through PET, SPECT and MR imaging modalities to non-invasively identify areas of apoptosis as an indicator of tumor response [91, 163, 164]. A recent in vivo study of lymphoma tumors within the hind limbs of mice using 99mTc-6-hydrazinonicotinic annexin V (99mTc-HYNIC-annexin V) imaging found that radiotracer uptake, as an indicator of apoptosis, significantly increased with radiation dose and correlated with terminal deoxynucleotidyl transferase-mediated deoxyuridine triphosphate nick end-labeling (TUNEL) assay quantifications of apoptotic cells [93]. A similar study was performed by Van de Wiele et al. in human patients that successfully correlated 99mTc-annexin V uptake in head and neck carcinomas with TUNEL assessed apoptosis in the tumor samples after surgical resection [165].

99Tc-Annexin V scintigraphy and SPECT has also been extensively evaluated as a non-invasive indicator of likely treatment response in a variety of tumor sites. Belhocine et al. used 99mTc-Annexin V scintigraphy before and within 3 days after the administration of chemotherapy in lung cancer, breast cancer, and lymphoma patients. They found that overall survival and progression-free survival were significantly correlated with the Annexin V radiotracer uptake after therapy in lung cancer and lymphomas [166]. Similarly, studies by Haas et al. on follicular lymphoma patients and Kartachova et al. on lymphoma, leukemia, non-small cell lung carcinoma and head and neck squamous cell carcinoma patients (HNSCC) found that ultimate tumor response was significantly correlated with annexin V based 99mTc radiotracer uptake following chemotherapy or radiation [94, 167]. In addition to correlation with treatment response, 99mTc-annexin V has also been shown to display radiation-dose-dependent uptake within parotid glands receiving radiation as part of HNSCC treatment plans [95].
Acknowledgements

This work was supported in part by the Emory University School of Medicine Department of Radiation Oncology Medical Student Scholars Program (Corso), the Department of Defense Lung Cancer Research Program Grant W81XWH-10-1-0605 (Ali), start-up funds from Emory University (Diaz), and the Department of Defense Breast Cancer Research Program Grant W81XWH-06-1-0788 (Diaz). We thank Sam Spratt for his artwork.

Abbreviations: CTL: Cytotoxic T Lymphocyte; HDMEC: Human dermal microvascular endothelial cell; HMEC: Human dermal microvascular endothelial cell; IR: Ionizing Radiation; RGD: Arginine-glycine-aspartic acid.

Please address correspondence to: Dr. Roberto Diaz, Department of Radiation Oncology, Emory University School of Medicine, and Winship Cancer Institute of Emory University, Suite A1300, 1365 Clifton Road, NE, Atlanta, GA 30322. Phone: 404-778-3669. Fax: 404-778-1750. E-mail: roberto.diaz@emory.edu

References

[14] Pattillo CB, Venegas B, Donelson FJ, Del Valle L,
Radiation induced neoantigens


[41] Buckley CD, Rainger GE, Bradfield PF, Nash GB
Radiation induced neoantigens


Radiation induced neoantigens

52: 5104-5109.


Radiation induced neoantigens


Radiation induced neoantigens


[138] Forero A, Weiden PL, Vose JM, Knox SJ, LoBuglio AF, Hankins J, Goris ML, Piccozi VJ, Aoworothy DB, Breit HB, Sims RB, Ghalie RG, Shen S and Meredith RF. Phase 1 trial of a novel anti-CD20 fusion protein in pretargeted...


Radiation induced neoantigens


