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Atbf1 Regulates Pubertal Mammary Gland Development Likely by Inhibiting the Pro-Proliferative Function of Estrogen-ER Signaling

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Abstract

ATBF1 is a candidate tumor suppressor that interacts with estrogen receptor (ER) to inhibit the function of estrogen-ER signaling in gene regulation and cell proliferation control in human breast cancer cells. We therefore tested whether Atbf1 and its interaction with ER modulate the development of pubertal mammary gland, where estrogen is the predominant steroid hormone. In an in vitro model of cell differentiation, i.e., MCF10A cells cultured in Matrigel, ATBF1 expression was significantly increased, and knockdown of ATBF1 inhibited acinus formation. During mouse mammary gland development, Atbf1 was expressed at varying levels at different stages, with higher levels during puberty, lower during pregnancy, and the highest during lactation. Knockout of Atbf1 at the onset of puberty enhanced ductal elongation and bifurcation and promoted cell proliferation in both ducts and terminal end buds of pubertal mammary glands. Enhanced cell proliferation primarily occurred in ER-positive cells and was accompanied by increased expression of ER target genes. Furthermore, inactivation of Atbf1 reduced expression of basal cell markers (CK5, CK14 and CD44) but not luminal cell markers. These findings indicate that Atbf1 plays a role in the development of pubertal mammary gland likely by modulating the function of estrogen-ER signaling in luminal cells and by modulating gene expression in basal cells.

In human breast cancer, although ATBF1 is infrequently mutated [4], its genomic locus is deleted in as high as 75% of ductal cancers and 100% of lobular cancers [6,17]. In addition, ATBF1 mRNA expression is often downregulated in human breast cancer, and the downregulation is associated with adverse features of breast cancer such as higher tumor stage and grade, larger tumor volumes, metastasis, and worse patient survival [7]. Interestingly, higher levels of ATBF1 expression were associated with estrogen receptor alpha (ERα, hereafter ER) positivity in breast cancer [7], and ATBF1 and the estrogen-ER signaling appear to form an autoregulatory feedback loop relationship [18,19,20]. On one hand, ATBF1 interacts with ER to inhibit the function of estrogen-ER signaling in gene regulation and cell proliferation control [20]. Proper ER function also appears to require fine-tuned levels of ATBF1, because ER induces ATBF1 transcription but causes ATBF1 protein degradation via the proteasome by inducing the estrogen responsive finger protein (ERF) [18,19].

Postnatal mammary gland development involves a number of different stages such as ductal elongation and bifurcation during puberty, side branching during estrous cycles, and alveologenesis and lactogenesis during pregnancy and lactation [21]. It is highly regulated by reproductive steroids including estrogen, progester-
Mammary epithelial cells, were purchased from the Jackson Lab (MMTV-Cre hereafter) mice, which specifically express the Cre in atbf1 in mammary gland development at least during the puberty. These findings indicate a regulatory role for enhancing the pro-proliferative function of estrogen-ER signaling, mammary gland promoted ductal elongation/bifurcation, likely by gland development. Furthermore, deletion of atbf1 expression varied during cell differentiation and mammary gland development. Moreover, deletion of Atbf1 in mouse mammary gland promoted ductal elongation/bifurcation, likely by enhancing the pro-proliferative function of estrogen-ER signaling, and attenuated the expression of basal cell markers in pubertal mammary gland. These findings indicate a regulatory role for Atbf1 in mammary gland development at least during the puberty.

Materials and Methods

Ethics statement
Mice used in these studies were housed at the Division of Animal Resources (DAR) facility at Emory University and handled by DAR staff. All mice were closely monitored and humanely euthanized. All experimental procedures involving animals were approved by the Institutional Animal Care and Use Committee (IACUC No. 2001337).

Reagents, antibodies, cell lines and mice

The following reagents were purchased from their respective vendors: Matrigel with reduced growth factor (BD Biosciences, San Jose, CA); Cell counting kit-8 (Dojindo Molecular Technologies, Gaithersburg, MD); Hematoxylin and permount mounting solution (Fisher Scientific, Waltham, MA); Immunohistochemical (IHC) staining reagents (Dako, Carpinteria, CA); Alexa Fluor Dye (Invitrogen, Carlsbad, CA); Fluoro-gel with TES buffer (Electron Microscopy Sciences, Hatfield, PA); β-casein antibody (Santa Cruz, Santa Cruz, CA); Ki67 antibody (Fisher Scientific); ERα antibody (Santa Cruz); CK5 antibody (Covance Research Products, Princeton, NJ); Human CK18 antibody (Dako); Mouse CK18 antibody (GeneTex, Irvine, CA). Generation of ATBF1 antibody, negative siRNA and ATBF1 siRNA was previously described [20].

The immortalized but non-tumorigenic human breast epithelial cell line MCF10A was purchased from ATCC (Manassas, VA) and cultured in DMEM/F12 medium supplemented with 5% horse serum, 20 ng/ml epidermal growth factor (EGF), 10 μg/ml insulin, 0.5 μg/ml hydrocortisone, and 100 ng/ml cholera toxin as described previously [23]. B6/129-TgMMTV-Cre/4 Mam/J (MMTV-Cre hereafter) mice were purchased from the Jackson Laboratory (Bar Harbor, ME).

Culture of MCF10A cells in Matrigel (3-D culture)

Briefly, 40 μl of Matrigel with reduced growth factor were added to each well of eight-well glass chamber slides and spread evenly. After Matrigel was solidified for 15 min, 3000 cells in each well were added in medium containing 5 ng/ml EGF and 2% Matrigel were seeded in each well, and the medium was replaced every three days. Images of spheres with defined scales were subjected to the ImageJ computer program to determine the area covered by each sphere, and the diameter of that sphere was then calculated based on the circle formula.

Mouse breeding and genotyping

Generation of mice containing the floxed Atbf1 allele (Atbf1fl) was previously described [26]. Floxed Atbf1 mice that had been crossed back to the C57BL/6j background were crossed with MMTV-Cre mice, and the F1 mice were then intercrossed. Female mice with the following genotypes, Cre+/Atbf1fl/fl (defined as Atbf1+/−, wild-type), Cre+/Atbf1fl/fl (defined as Atbf1+/−, heterozygous deletion), and Cre+/Atbf1fl/fl (defined as Atbf1−/−, homozygous deletion), were collected. Mice were genotyped by PCR with tail DNA and mammary gland genomic DNA using the following primers: 5'-GGCCCTTGACTGGTCTTTCCGTGTCGTAAGGCTGAACGAGTGATCGCGTCGATGCAACGAGTGAT-3' and 5'-ATTGATGTGAATGGAAGGTGTCAGAAGCAGCTGGAGATCAGACAGTACGTGAGATG3' (+/−) and 5'-CAGGTCGATACACAGAATGCTGGAGATCAGACAGTACGTGAGATG3' (−/−). Whole mount analysis of mammary glands

The fourth abdominal mammary gland on the left was harvested from pubertal mice, placed between two glass slides, and spread by placing weights on top of the slides for 10 min, followed by fixing in 3.7% formalin overnight. Fixed tissues were washed in serially diluted ethanol and stained at room temperature with hematoxylin for 5–10 min. Stained tissues were washed by flowing water, dehydrated progressively in graded alcohol solutions (70%, 95% and 100%; 30 min each), and cleared in xylene for at least 30 min before mounting with cover slips. Slides with whole mount mammary glands were scanned at a 2400 dpi resolution.

Immunohistochemical (IHC) and Immunofluorescence (IF) staining

Tissue sections were prepared from formalin-fixed paraffin-embedded tissues for IHC and IF staining. Briefly, tissue sections were first deparaffinized and rehydrated following the standard procedure. After the treatment with blocking buffer, sections were heated in sodium citrate buffer for antigen retrieval. For IHC staining, sections were incubated with ATBF1 antibody (1:1000) at 4 °C overnight and then with the HRP solution, DAB-chromogen, hematoxylin, dehydrated and finally stored in Permount mounting solution. For IF, the blocking solution was 10% normal goat serum in PBS and the secondary antibody was conjugated with the Alexa Fluor Dye. Nuclei were stained with diamidino-phenyl-indole (DAPI) and sections in the anti-fade mounting solution were stored at −20 °C in the dark.

RNA extraction, RT-PCR and Real time RT-PCR

The fourth abdominal mammary gland on the right, without lymph node, was harvested for RNA and protein analysis. For RNA extraction, harvested tissues were immediately submerged in RNAlater (RNA stabilization reagent). Total RNA from the mammary glands was extracted using the RNeasy Mini Kit following the manufacturer's instructions (Qiagen, Valencia, CA). The first strand cDNA was synthesized by using the iScript cDNA synthesis kit (Bio-Rad, Richmond, CA). Primer sequences for

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semi-quantitative PCR to determine the deletion of Atbf1 at the RNA levels were 5'-GGCCAGATCTTCACCATCC-3' (forward) and 5'-CAGGGAGGAACATGCTACTAGG-3' (reverse). Primer sequences for real time RT-PCR are shown in Table 1.

Protein extraction and western blotting
Western blotting was performed as previously described [27,28]. Briefly, harvested tissues were flash-frozen in liquid nitrogen and lysed in cold RIPA buffer (20 mM Tris pH 8.0, 150 mM NaCl, 0.5% sodium deoxycholate, 0.1% SDS, 1% NP-40, 10 mM sodium pyrophosphate, and 10 mM sodium fluoride) supplemented with protease/phosphatase inhibitors. Cultured cells were washed with PBS and lysed in the same RIPA buffer. Lysates were incubated on ice for 20 minutes with frequent vortexing and cleared by centrifugation. Proteins were separated by 4% SDS-PAGE for ATBF1 and 10% SDS-PAGE for β-actin. The ATBF1 antibody was at 1:800 dilution in 3% BSA/PBS and β-actin antibody was at 1:10,000.

Statistical analysis
Statistical analyses were performed using the SPSS® statistical software (SPSS Inc., Chicago, IL, USA). Student’s t test was used to determine statistical differences between two groups, whereas one-way ANOVA or univariate analysis was used to compare three or more groups, as detailed in figure legends. P values less than 0.05 were considered statistically significant.

Results

Induction of ATBF1 expression during MCF10A cell differentiation
MCF10A cells cultured in Matrigel (3-D) proliferate and differentiate to form spheres with acini, and thus have been used as an in vitro model that mimics mammary epithelial differentiation [25,29,30]. To test whether ATBF1 plays a role in mammary epithelial differentiation, we first examined the expression of ATBF1 in MCF10A cells in both 2-D (plastic plate) and 3-D (Matrigel) cultures. In the 3-D culture, differentiation of MCF10A cells was indicated by the formation of acini (Fig. 1A–1C) and the induction of differentiation markers β-casein in the space surrounding acini (Fig. 1A) and CK18 in the cytoplasm (Fig. 1B, 1D). Induction of β-casein and CK18 was detectable at day 14 but not at day 7. Expression of basal markers CK14 and CK5, on the other hand, was decreased during the differentiation (Fig. 1E). For ATBF1, a significant induction was detected at day 7, and the induction was still demonstrable at day 14, although at a lower level (Fig. 1C, 1D). In the 2-D culture, ATBF1 expression was detected at a lower level (Fig. 1D) without an increase during a 9-day period of culture (data not shown). These results suggest that the induction of ATBF1 is related to cell differentiation.

To determine the role of induced ATBF1 in the differentiation of MCF10A cells in Matrigel, we knocked down ATBF1 by transfecting siRNAs into MCF10A cells before they were plated into Matrigel (Fig. 2A, 2B). Interference with ATBF1 expression

Table 1. Primer sequences for real time RT-PCR.

<table>
<thead>
<tr>
<th>Gene</th>
<th>Forward</th>
<th>Reverse</th>
</tr>
</thead>
<tbody>
<tr>
<td>hATBF1</td>
<td>TGTTCCAGATCGAGATGGGAAT</td>
<td>CTTTCCCCAGATCTCTGAGGTTT</td>
</tr>
<tr>
<td>hCK18</td>
<td>CGCCAGGCCGACCGAGATGAGG</td>
<td>ACTATCAGGGGCTGCTGTTT</td>
</tr>
<tr>
<td>hCD44</td>
<td>CAACCGAGACGAGGATCTC</td>
<td>GATAACCTCGAGTTGCTC</td>
</tr>
<tr>
<td>hCK14</td>
<td>TTCTGAGGAGATGAGGTG</td>
<td>GACGGTGAGGCTGCTGT</td>
</tr>
<tr>
<td>hCK5</td>
<td>TGTGCTCGGCGACCGAGATG</td>
<td>ATTGGGATGGGATGAGGTCTT</td>
</tr>
<tr>
<td>hCD24</td>
<td>TGAAGAACTGAGAGGGTGAC</td>
<td>GAAAGTGAACCTCAGGACAAA</td>
</tr>
<tr>
<td>hCK18</td>
<td>CGCCAGGCCGAGATGAGG</td>
<td>ACTATCAGGGGCTGCTGTTT</td>
</tr>
<tr>
<td>hCK8</td>
<td>GCTGACGACGAGATC</td>
<td>CATGAGACAGACAGAGATG</td>
</tr>
<tr>
<td>hGAPDH</td>
<td>GGTGGTCTCTCTGACAGTTCA</td>
<td>GTTGGCTAGCCAAATTGCTGT</td>
</tr>
<tr>
<td>mAtbf1 (regular)</td>
<td>GGGCCAGATCTCTCCAGGAG</td>
<td>CAGGAGGAAATGCCTACTAGG</td>
</tr>
<tr>
<td>mAtbf1 (real time)</td>
<td>AGAAGGAGGACGGAGCATC</td>
<td>CGGTTACGAGGTTGCTATAC</td>
</tr>
<tr>
<td>mER</td>
<td>TTCTGACGACGAGAATGCTG</td>
<td>CATCTCCAGGAGGAGTACAGAGG</td>
</tr>
<tr>
<td>mCK5</td>
<td>GGAGCTGCTCTCAAAATG</td>
<td>TCCAGACGCTCTGAGT</td>
</tr>
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<td>mCK14</td>
<td>GCTCTTGGTACATGAGGTGT</td>
<td>GAGGAGGAAAGGAGGAGGT</td>
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<tr>
<td>mCD44</td>
<td>TGGATCGGAAATTAGCTGAC</td>
<td>AGCTTTTACTCTGCCACAA</td>
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<tr>
<td>mCK8</td>
<td>ATCGGGATCACCGACCTACCG</td>
<td>TGAGATCCGGAAGATAGGT</td>
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<tr>
<td>mCK18</td>
<td>CGAAGGCTCAGAAAGACTGAC</td>
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<td>mCD24</td>
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<td>mEBag9</td>
<td>GATGGAGCTTCCTCCAGGATGGA</td>
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<td>mIgf-1</td>
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<td>CCACACGAAACTGAAGACAT</td>
</tr>
<tr>
<td>mMyc</td>
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<tr>
<td>mGapdh</td>
<td>CGGTATGACTCCACATCAG</td>
<td>GACTCCAGGACATACCTAGC</td>
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</tbody>
</table>

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resulted in a significant inhibition of mammosphere formation of MCF10A cells in Matrigel, as indicated by the number of spheres with a diameter greater than 75 μm (Fig. 2C–2E). In the 2-D culture, however, MCF10A cell proliferation was slightly enhanced by the knockdown of ATBF1 (Fig. 2F) suggesting that the attenuation of mammosphere formation by ATBF1 knockdown may not be caused by the increase in cell proliferation but likely by a change in cell differentiation. Furthermore, knockdown of ATBF1 reduced the expression of basal cell markers CD44, CK14 and CK5, but not that of luminal cell markers CD24, CK18 or CK8 (Fig. 2G). These results suggest that ATBF1 plays a role in the differentiation of mammary epithelial cells, which might involve the maintenance of the basal cell layer.

Expression of Atbf1 in mouse mammary glands

In order to test the function of Atbf1 in a mouse model, we first evaluated the expression of Atbf1 during mammary gland development in mouse. We collected mammary glands at four different stages (virgin, pregnancy, lactation and involution), and performed real time RT-PCR to measure Atbf1 mRNA expression. The expression of ER was used as a control for gene expression in mammary tissues at different stages. As expected, ER expression was higher during puberty and pregnancy but lower during lactation (Fig. 3A). The level of Atbf1 expression also varied at different stages, with an increase from week 3 to week 9 during puberty but relatively lower levels during pregnancy. During lactation, Atbf1 mRNA was at significantly higher levels, with the highest at day 7 (Fig. 3A). Moreover, IF staining showed that Atbf1 was expressed in the nucleus of both luminal and myoepithelial cells, with strong staining in some luminal cells (Fig. 3B). Dynamic Atbf1 expression during mammary gland development suggests that Atbf1 plays different roles in different stages of developing mammary gland, and is likely more relevant to puberty and lactation.

Conditional knockout of Atbf1 in mouse mammary glands

To better understand the function of Atbf1 in mammary gland development, we bred floxed Atbf1 mice [26] to MMTV-Cre mice to specifically knock out Atbf1 in mouse mammary epithelial cells. To detect Cre-mediated Atbf1 deletion, we designed a pair of PCR primers, of which one was upstream to the first loxP site while the other was downstream to the second loxP site. The primers could produce a small PCR product (289 bp) when the floxed Atbf1 allele was deleted by Cre. As expected, breeding with Cre mice resulted in PCR products indicative of Atbf1’s genomic deletion (Fig. 3C) and truncated Atbf1 mRNA (Fig. 3D) in mammary gland tissues with both heterozygous and homozygous Atbf1 deletion (+/− and −/− respectively). At the protein level, both western blotting and IHC staining revealed that knockout of Atbf1 significantly reduced Atbf1 protein expression in mammary glands even when the deletion occurred in one of the two alleles (Fig. 3E, 3F).
Knockout of Atbf1 promotes ductal elongation and bifurcation in pubertal mammary gland

To investigate the functional contribution of Atbf1 in pubertal mammary gland development, we isolated the fourth abdominal mammary glands from mice at the age of 3, 5, 6 and 8 weeks, performed whole mount analyses, and measured ductal elongation and bifurcation. Atbf1 deletion significantly accelerated ductal elongation and bifurcation, as shown by direct visualization of ductal trees (Fig. 4A) and statistical analysis of ductal invasion and the number of branches at 5- and 6-week old (Fig. 4B, 4C). Atbf1 deletion did not affect the number of terminal end buds (TEBs) though (Fig. 4D). At 8 weeks, which marks the end of puberty in mice, ductal extension and branching equalized, and the effect of Atbf1 deletion disappeared (Fig. 4A, 4E and 4F). Taken together, these findings suggest that, whereas the number of TEBs is not affected, loss of Atbf1 enhances ductal elongation and bifurcation in the mammary tree during puberty.

In parallel, we fixed thoracic mammary glands at the age of 6 week, embedded in paraffin, and prepared HE-stained tissue sections for duct morphometrics (Fig. 4G). The average number of ducts per square millimeter (mm²) in tissue sections was 2.12, 3.56, and 4.75 for wild-type, heterozygous Atbf1 deletion, and homozygous Atbf1 deletion, respectively, and the difference between Atbf1+/− mice and Atbf1+/+ mice was statistically significant (Fig. 4H). Measurement of duct coverage, as defined by the percentage of tissue areas covered by ductal areas, demonstrated that Atbf1 deletion significantly increased duct coverage when compared to normal glands (Fig. 4I). Consistent with the number of ducts per mm², the size of ducts was also significantly increased by the deletion of Atbf1 (Fig. 4J). These findings further indicate that deletion of Atbf1 promotes ductal elongation and bifurcation in pubertal mammary gland.

Knockout of Atbf1 enhances cell proliferation primarily in ER-positive mammary epithelial cells

During puberty, the major activities in mammary gland development are ductal elongation and bifurcation accompanied predominantly by cell proliferation. We evaluated cell proliferation in mammary epithelial cells by measuring the Ki67 proliferation marker. In thoracic mammary glands from 6-week old mice, IF staining of tissue sections from nine mice (3 for each genotype) showed that ducts with Atbf1 knockout (both +/− and −/−) had significantly more Ki67-positive cells than ducts with wild-type Atbf1 (Fig. 5A, 5C). Similarly, TEBs with Atbf1 deletion had significantly more Ki67-positive cells than those without deletion (Fig. 5B, 5C).

Estrogen is the most dominant hormone in mammary epithelial proliferation and differentiation during puberty [23]. Our previous in vitro studies demonstrated that ATBF1 and estrogen-ER signaling formed an autoregulatory feedback loop to regulate cell proliferation in ER-positive cells [18,19,20], and that there was also a correlation between ATBF1 expression and ER positivity in human breast cancer [7]. We therefore evaluated the effect of Atbf1 knockout on mammary epithelial proliferation in the context of ER activity. We co-stained ER and Ki67 in mammary epithelium cells from ducts and TEBs with wild-type and heterozygous or homozygous Atbf1 deletion (Fig. 6A, 6B). Statistical analysis showed that the increase in cell proliferation...
caused by Atbf1 deletion, as indicated by increased Ki67 positive cells, was only statistically significant in ER-positive cells and not in ER-negative cells, in both ducts and TEBs (Fig. 6C, 6D). As expected, the percentage of ER-positive cells among Ki67-positive cells was significantly higher when Atbf1 was deleted (Fig. 6E). These results suggest that, during puberty, Atbf1 functions to inhibit cell proliferation primarily in ER-positive mammary epithelial cells.

Dysregulation of ER target genes and basal cell markers by Atbf1 deletion in mammary glands

Our previous study indicates that ATBF1 inhibits the function of ER in both cell proliferation and ER target gene expression [20]. Therefore, we evaluated whether knockout of Atbf1 in vivo also alters the expression of ER target genes, including amphiregulin (Areg), Igf-1, Ebag9, Cisd, c-Myc [20,31,32]. Areg and Igf-1 are two well-established estrogen-induced genes that regulate terminal end bud formation and ductal morphogenesis in pubertal mammary gland [33,34]. Except for Cisd, all other tested ER
target genes were significantly up-regulated with the deletion of Atbf1 (Fig. 6F). Areg, in particular, was increased by about 8 fold by homozygous deletion of Atbf1. Therefore, Atbf1 also inhibits ER-regulated gene expression in vivo.

The mammary gland is comprised of an adipose-rich stroma embedded with a branching network of epithelial ducts, which are organized in two distinct cell layers: the basal myoepithelial layer and the luminal epithelial layer. Proper organization of myoepithelial and luminal epithelial cells is crucial for the normal structure and function of mammary glands [35]. Moreover, the myoepithelial layer is not only abundant in stem cells but also associated with cell differentiation during mammary gland development [36,37,38] and to suppress tumor development [39,40]. CK5, CK14 and CD44 as myoepithelial cell markers and CK18, CK8 and CD24 as luminal cell markers are generally used as structural and differentiation markers [41,42,43]. Immunofluorescence staining of CK5 and CK18 revealed that knockout of Atbf1 didn’t affect cellular localization of both CK5 and CK18.
Discusssion

Atbf1 regulates pubertal mammary gland development

Mammary gland development is a highly regulated process involving balanced cell proliferation and differentiation under the control of female reproductive hormones, including estrogen, progesterone and prolactin. During puberty, estrogen is the dominant steroid hormone, and multiple factors have been identified for their effects on mammary gland development. Our findings in this study suggest that Atbf1 is one of the regulators controlling pubertal mammary gland development. First, using an in vitro model of mammary epithelial differentiation – the MCF10A cells grown in Matrigel, we found that ATBF1 was induced during cell differentiation, and knockdown of ATBF1 interfered with the differentiation (Fig. 1, Fig. 2). More directly, Atbf1 expression increased from week 3 to week 9 during puberty but fell during pregnancy (Fig. 3A), and knockout of Atbf1 promoted ductal elongation and bifurcation in pubertal mammary glands, which was indicated by extended ductal invasion, total ductal branching, and increased ductal coverage upon the deletion of Atbf1 in pubertal mammary glands (Fig. 4). During puberty, ductal elongation and bifurcation are accompanied predominantly by cell proliferation. Consistent with the ductal phenotype in the pubertal mammary gland, knockout of Atbf1 significantly increased cell proliferation (Fig. 5). Taken together, these findings establish Atbf1 as a novel regulator of pubertal mammary gland development.

The effect of Atbf1 on mammary epithelial proliferation is limited to ER-positive cells

The mammary gland is comprised of ER-positive and ER-negative cells, and estrogen is the most dominant hormone in mammary epithelial proliferation and differentiation during puberty [23]. Several studies have demonstrated that ER-positive cells in normal mammary gland do not proliferate, and estrogen stimulates the proliferation of ER negative cells via a paracrine mechanism [32,44], which is different from ER-positive breast cancer cells, where ER-negative cells do proliferate. Our findings in this study indicated that knockout of Atbf1 primarily makes ER-positive cells proliferate (Fig. 6).

The biological function of estrogen is largely mediated by ER, which is primarily associated with ductal elongation rather than lobuloalveolar formation in the mammary gland [45,46]. Early and complete loss of ER in mammary epithelia prevents the formation of TEBs and severely impairs ductal elongation [47,48]. While it is not known why and how the estrogen-ER signal stimulates the proliferation of ER-negative but not ER-positive cells, it is possible that Atbf1 could be partially responsible for the lack of proliferation in ER-positive cells because knockout of Atbf1 increased the proliferation of ER-positive cells (Fig. 6). Our previous studies in which ATBF1 was identified as a functional inhibitor of ER in gene regulation and cell proliferation control and in which an autoregulatory negative feedback loop was established between ATBF1 and the estrogen-ER signaling in ER-positive human breast cancer cells [18,19], support this prediction. ATBF1 inhibits estrogen-ER signaling via direct interaction with ER, which prevents the binding of the steroid receptor coactivator 3 [SRC3/AIB1] to ER, altering the expression of ER target genes such as CTSD and EBAG9 [20]. In pubertal mammary gland, ablation of Atbf1 upregulated Ebag9 (Fig. 6F), but not Ctsd (data not shown).

Knockout of Atbf1 also significantly upregulated the expression of amphiregulin (Areg) (Fig. 6F), an EGF family member transcriptionally induced by estrogen in pubertal mammary glands
during the exponential expansion of the ductal system [31]. Areg is a key mediator of the estrogen-driven epithelial cell proliferation and ductal elongation at puberty [31,49], and it is possible that Areg mediates the acceleration of mammary gland branching and bifurcation upon the deletion of $Atbf1$.

In addition to Areg, two other ER target genes involved in the regulation of cell proliferation, $Igf-1$ and $c-Myc$ [50,51], were also upregulated by the deletion of $Atbf1$ (Fig. 6F). A number of studies have established the crosstalk between $Igf-1$ and the estrogen-ER signaling [52,53,54]. On one hand, the estrogen-ER signaling stimulates the local synthesis of $Igf-1$ [50,55]; while on the other hand, ER transcriptional activity can be induced by the $Igf-1$ signaling in an estrogen-independent manner [52,54]. $Igf-1$ is essential for TEB formation and ductal morphogenesis in pubertal mammary gland [34,56]. The Myc protein is a well established stimulator of cell proliferation, differentiation and apoptosis [57]. The increase in cell proliferation caused by $Atbf1$ ablation could also involve the upregulation of both IGF-1 and Myc. Taken together, these findings support the view that $Atbf1$ inhibits the proliferation of ER-positive cells by suppressing the estrogen-ER signaling pathway.

Although normal ER-positive cells do not proliferate and estrogen-ER signaling induces the proliferation of ER-negative cells via a paracrine mechanism [44,58], estrogen-ER signaling is clearly mitogenic and promotes cell proliferation in ER-positive breast cancer cells. It is unknown how this biological change occurs between normal and cancer cells. ATBF1 not only inhibits the proliferation of ER-positive cells in both normal mammary gland and breast cancer cells, it is also frequently inactivated by genomic deletion and downregulation in human breast cancers, and its downregulation is associated with worse patient survival in ER-positive breast cancer [4,6,7] (our unpublished data). Therefore, we speculate that molecular machinery exists to limit normal ER-positive cells from proliferating, and ATBF1 is part of this machinery.

$Atbf1$ could be more relevant to the differentiation than the proliferation of mammary epithelial cells

In normal mammary gland, expression of ER is an indication of epithelial differentiation, which could be the reason why ER-positive cells do not proliferate. Based on the observation that $Atbf1$ inhibits ER function [20], increased cell proliferation led by the knockout of $Atbf1$ in ER-positive cells could mean a more
relevant function of Atbf1 in mammary epithelial differentiation. For example, expression of Atbf1 could cooperate with the estrogen-ER signaling to induce and maintain the differentiation status of mammary epithelial cells. Supporting this idea is the induction of ATBF1 during the differentiation of the MCF10A cells grown in Matrigel, which was accompanied by the expression of differentiation markers [Fig. 1]. An increased expression of Atbf1 in lactating mammary glands (Fig. 3A) could also be related to Atbf1’s function in cell differentiation because lactating mammary glands are highly differentiated. A role of ATBF1 in cell differentiation has been suggested by a series of other studies. For example, during brain development Atbf1 expression was increased in the differentiating field when compared to the proliferating stem cells on the ventricular zone [13]. Consistently, when retinoic acid induces neuronal differentiation in the P19 mouse neuroblastoma cell line, Atbf1 expression is elevated by 50-fold within 24 hours [59,60]. The function of ATBF1 in cell cycle arrest, as demonstrated in other studies [3,9], could indirectly indicate a role of ATBF1 in cell differentiation in vivo.

On the other hand, we found that downregulation of ATBF1 reduced the expression of basal cell markers but not that of luminal cell markers, which was detected in both the human MCF10A cells grown in Matrigel in vitro and mouse mammary gland in vivo. These results suggest that Atbf1 is needed for the maintenance of basal cells, which has been shown to play a crucial role in cell differentiation. Bocker and his colleagues described a population of normal human breast cells that express cytokeratin-5/6 (CK5/6) and represent stem cells that give rise to both differentiated luminal cells and myoepithelial cells [43]. The adhesion receptor CD44, which is also a stem cell marker, is expressed in the myoepithelium of the developing mammary gland, mediates epithelial-stromal and cell-cell interactions and modulates ductal development [35]. In another study, three distinct human breast epithelial subsets (basal and myoepithelial cells; luminal progenitor cells, and luminal cells) were identified from the resultant lineage-negative (Lin-) population with antibodies against CD49f and epithelial cell adhesion molecule (EpCAM). CK5/6 is expressed in both the basal (CD49fhiEpCAMlo) and luminal progenitor cells (CD49fhiEpCAMhi) but not in mature luminal cells (CD49fEpCAMhi), which suggests that CK5/6-positive cells are progenitor cells and may contribute to cell differentiation [61]. In breast cancer cells, myoepithelial cells induce growth arrest and apoptosis with secretion regulation of ECM proteins, anti-angiogenic factors, and protease inhibitors [40,62]. When co-cultured with breast cancer cells, myoepithelial cells inhibit expression of MMPs [63] and direct polarization and branching morphogenesis during mammary gland development [64,65].

Atbf1 could regulate mammary gland development during other stages

The majority of mammary gland development takes place after birth under the control of steroid hormones. Ductal elongation and bifurcation during puberty, which is mostly regulated by estrogen, is only the first major phase. After sexual maturation, recurrent estrous cycles trigger side branching with estrogen and progesterone, pregnancy enhances side branching and induces alveologenesis with lactational differentiation via progesterone and...
prolactin, and involution occurs at weaning [23,24]. Our unpublished data suggest that Atbf1 regulates the progesterone-PR pathway and Pg-induced cell differentiation. Moreover, Atbf1 expression was highly induced during the lactating stage (Fig. 3A). These findings suggest a role of Atbf1 in other stages of mammary development, which remains to be determined.

**Atbf1 could be a novel factor that regulates both mammary gland development and tumorigenesis**

Mouse mammary gland development results from balanced cellular activities including proliferation, differentiation, and apoptosis, and disruption of these processes could lead to tumorigenesis. A number of factors have been discovered to have regulatory roles in both mammary gland development and tumorigenesis. For example, the Gata-3 transcription factor, which is specifically located in luminal cells, not only regulates mammary gland morphogenesis and luminal cell differentiation during both puberty and pregnancy [42,66], it has also been implicated in breast cancer [67]. The oncogenic p120-catenin is crucial for E-cadherin function, and ablation of p120 causes a delay in TEB outgrowth during puberty [68]. Loss of the breast cancer 1 (Brca1) tumor suppressor in mammary epithelium alters the estrogenic growth response, and exposure to increased estrogen or ER activity collaborates with Brca1 deficiency to accelerate ductal elongation and TEB differentiation [69]. Conditional knockout of phosphatase and tensin homolog (Pten) leads to excessive ductal branching and precocious lobulo-alveolar development, delayed involution, and severely reduced apoptosis as well as neoplasia in mammary glands [70]. Atbf1 has been established as a tumor suppressor by its frequent mutation in human prostate cancer [3] and induction of precancerous lesions upon deletion in the prostate (Sun et al., unpublished data). It has also been implicated in breast cancer by frequent genomic deletion and downregulation [4,7]. Our findings in this study indicate that Atbf1 is also a regulator of mammary gland development at least during puberty.

In summary, we examined the expression and function of Atbf1 in mouse mammary gland development, and found that Atbf1 was expressed in the nucleus of most mammary epithelial cells with varying levels during different stages of mammary gland development. Knockout of Atbf1 enhanced ductal elongation and bifurcation, and promoted cell proliferation primarily in ER-positive cells during puberty. In addition, knockout of Atbf1 significantly reduced the expression of basal cell markers (CK5, CK14 and CD44) in vitro and in vivo. Taken together, these findings suggest that Atbf1 plays a role in the development of pubertal mammary gland likely by modulating the function of estrogen-ER signaling and the maintenance of basal cells.

**Author Contributions**

Conceived and designed the experiments: JTD ML. Performed the experiments: ML XF GM CX JL. Analyzed the data: ML XS XD TN JTD. Wrote the paper: ML JTD.

**References**

22. Siegel PM, Muller WJ (2010) Transcription factor regulatory networks in mammary gland morphogenesis and lobulo-alveolar development, delayed involution, and severely reduced apoptosis as well as neoplasia in mammary glands [70]. Atbf1 has been established as a tumor suppressor by its frequent genomic deletion and downregulation [4,7]. Our findings in this study indicate that Atbf1 is also a regulator of mammary gland development at least during puberty.